

FLAVONOIDS FROM *Millettia cubitti*

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The genus *Millettia* (Leguminosae) is comprised of more than 200 species of climbers and trees distributed in tropical Africa, Asia, and Australia [1]. *Millettia* plants have been known to be a rich source of flavonoids [2–5]. *Millettia cubitti* Dunn is a tree distributed in the south of Yunnan Province, China [6]. There have been no reports on the chemical composition of this plant so far. As part of our continuing studies on bioactive compounds from *Millettia* plants, we have first carried out an investigation of the chemical constituents of the title plant.

The twigs of *Millettia cubitti* were collected from Xishuangbanna, Yunnan Province, P. R. China in April 2013. It was dried (7.5 kg), ground, and macerated with 95% EtOH. The ethanol extract was then successively extracted with chloroform (CHCl₃) and ethyl acetate (EtOAc) to give a CHCl₃ extract (128 g) and an EtOAc extract (29 g), respectively. The CHCl₃ extract was subjected to silica gel column chromatography (CC) with petroleum ether (PE)–EtOAc step-gradient elution (9:1→4:6) to yield compounds **2** (25 mg), **4** (28 mg), and 7 fractions (A–G). Fraction C (2.2 g) was separated by reversed-phase C₁₈ (RP-18) CC eluted with MeOH–H₂O (80→90%) to give compound **1** (15 mg). Fraction D (5.0 g) was chromatographed over silica gel CC eluted with PE–EtOAc (4:1, 3:1, 2:1, 6:4) to afford compound **6** (20 mg) and fractions D1–D6. Fraction D4 was subjected to RP-18 CC eluted with MeOH–H₂O (70→90%) to obtain compounds **7** (18 mg) and **9** (16 mg). Fraction E (5.6 g) was subjected to silica gel CC eluted with CHCl₃–MeOH (40: 1, 20:1, 10:1) to give compound **8** (17 mg) and fractions E1–E5. Fraction E2 was further purified by CC on Sephadex LH-20 eluted with MeOH to afford compound **5** (16 mg). Fraction E3 was further separated by RP-18 CC eluted with MeOH–H₂O (60→90%) to give compounds **3** (14 mg) and **10** (19 mg).

The isolated compounds were identified as 5-hydroxy-6'',6''-dimethylchromene-[2'',3'':7,8] flavone (**1**) [7], maximaisoflavone B (**2**) [8], cuneatin methyl ether (**3**) [9], isoerythrinin A 4'-(3-methylbut-2-enyl) ether (**4**) [10], 6-methoxy-7-hydroxy-3',4'-methylenedioxyisoflavone (**5**) [11], isopongaflavone (**6**) [12], formononetin (**7**) [13], fujikinetin methyl ether (**8**) [9], durmillone (**9**) [10], and 3',4'-methylenedioxy-[2'',3'':7,8]furanoisoflavone (**10**) [14] by spectral analysis using NMR spectrometers, including 2D NMR, and comparison with values reported in the literature. All compounds were isolated from *Millettia cubitti* for the first time. As far as we know, prior to this study, ¹³C NMR data of compounds **1** and **2** and PMR and ¹³C NMR data of **10** have not been reported. This is the first time that the data mentioned have been confirmed.

5-Hydroxy-6'',6''-dimethylchromene-[2'',3'':7,8]flavone (1). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 182.5 (C-4), 163.5 (C-2), 161.7 (C-7), 159.6 (C-5), 151.9 (C-8a), 131.8 (C-4'), 131.4 (C-1'), 129.1 (C-3', 5'), 127.6 (C-5''), 126.2 (C-2', 6'), 114.8 (C-4''), 105.8 (C-3), 105.5 (C-8), 101.4 (C-4a), 100.4 (C-6), 78.1 (C-6''), 28.2 (C-7'', 8'').

Maximaisoflavone B (2). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 175.7 (C-4), 163.2 (C-7), 157.8 (C-8a), 152.2 (C-2), 147.6 (C-3', 4'), 139.4 (C-3''), 127.5 (C-5), 125.7 (C-1'), 124.9 (C-3), 122.3 (C-6'), 118.5 (C-2''), 118.1 (C-4a), 115.0 (C-6), 109.7 (C-2'), 108.3 (C-5'), 101.1 (OCH₂O), 100.8 (C-8), 65.4 (C-1''), 25.8 (C-4''), 18.3 (C-5'').

3',4'-Methylenedioxy-[2'',3'':7,8]furanoisoflavone (10). ¹H NMR (400 MHz, CDCl₃, δ, ppm, J/Hz): 8.24 (1H, d, J = 8.8, H-5), 8.06 (1H, s, H-2), 7.76 (1H, d, J = 2.0, H-5''), 7.58 (1H, d, J = 8.8, H-6), 7.14 (1H, d, J = 1.7, H-2'), 7.14 (1H, d, J = 2.0, H-4''), 7.02 (1H, dd, J = 8.0, 1.7, H-6'), 6.90 (1H, d, J = 8.0, H-5'), 6.01 (2H, s, OCH₂O). ¹³C NMR (100 MHz, CDCl₃, δ, ppm): 176.1 (C-4), 158.1 (C-7), 151.7 (C-2), 150.8 (C-8a), 147.7 (C-3', 4'), 145.7 (C-5''), 125.6 (C-3), 125.5 (C-1'), 122.5 (C-6'), 122.4 (C-5), 119.9 (C-4a), 116.9 (C-8), 110.3 (C-6), 108.4 (C-5'), 104.2 (C-4''), 101.1 (OCH₂O).

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REFERENCES

1. F. Ngandeu, M. Bezabih, D. Ngamga, A. T. Tchinda, B. T. Ngadjui, B. M. Abegaz, H. Dufat, and F. Tillequin, *Phytochemistry*, **69**, 258 (2008).
2. M. Chatsumpun, B. Sritularak, and K. Likhitwitayawuid, *Chem. Nat. Compd.*, **46**, 634 (2010).
3. Z. Na, Q. S. Song, and H. B. Hu, *Rec. Nat. Prod.*, **7**, 307 (2013).
4. B. Sritularak, K. Likhitwitayawuid, J. Conrad, B. Vogler, S. Reeb, I. Claiber, and W. Kraus, *J. Nat. Prod.*, **65**, 589 (2002).
5. B. Sritularak and K. Likhitwitayawuid, *Phytochemistry*, **67**, 812 (2006).
6. Editorial Committee of Flora Reipublicae Popularis Sinicae, *Flora of China*, Vol. 40, Science Press, Beijing, 1994, 135 p.
7. M. C. Do Nascimento, R. L. de Vasconcellos Dias, and W. B. Mors, *Phytochemistry*, **15**, 1553 (1976).
8. E. V. Rao and M. S. R. Murthy, *Phytochemistry*, **24**, 875 (1985).
9. N. C. Veitch, P. S. E. Sutton, G. C. Kite, and H. E. Ireland, *J. Nat. Prod.*, **66**, 210 (2003).
10. A. Yenesew, J. O. Midiwo, and P. G. Waterman, *Phytochemistry*, **41**, 951 (1996).
11. E. V. Rao, M. S. R. Murthy, and R. S. Ward, *Phytochemistry*, **23**, 1493 (1984).
12. C. C. Andrei, D. T. Ferreira, M. Faccione, L. A. B. de Moraes, M. G. de Carvalho, and R. Braz-Filho, *Phytochemistry*, **55**, 799 (2000).
13. A. A. Drenin, E. Kh. Botirov, and E. V. Petrulyak, *Chem. Nat. Compd.*, **44**, 24 (2008).
14. K. Fukui, M. Nakayama, and K. Okazaki, *Nippon Kagaku Zasshi*, **85**, 446 (1964).