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Water relations and gas exchange of fan bryophytes and their adaptations to microhabitats in an Asian subtropical montane cloud forest

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Abstract Fan life forms are bryophytes with shoots rising from vertical substratum that branch repeatedly in the horizontal plane to form flattened photosynthetic surfaces, which are well suited for intercepting water from moving air. However, detailed water relations, gas exchange characteristics of fan bryophytes and their adaptations to particular microhabitats remain poorly understood. In this study, we measured and analyzed microclimatic data, as well as water release curves, pressure-volume relationships and photosynthetic water and light response curves for three common fan bryophytes in an Asian subtropical montane cloud forest (SMCF). Results demonstrate high relative humidity but low light levels and temperatures in the understory, and a strong effect of fog on water availability for bryophytes in the SMCF. The facts that fan bryophytes in dry air lose most of their free water within 1 h, and a strong dependence of net photosynthesis rates on water content, imply that the transition from a hydrated, photosynthetically active state to a dry, inactive state is rapid. In addition, fan bryophytes developed relatively high cell

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wall elasticity and the osmoregulatory capacity to tolerate desiccation. These fan bryophytes had low light saturation and compensation point of photosynthesis, indicating shade tolerance. It is likely that fan bryophytes can flourish on tree trunks in the SMCF because of substantial annual precipitation, average relative humidity, and frequent and persistent fog, which can provide continual water sources for them to intercept. Nevertheless, the low water retention capacity and strong dependence of net photosynthesis on water content of fan bryophytes indicate a high risk of unbalanced carbon budget if the frequency and severity of drought increase in the future as predicted.

Keywords Desiccation tolerance · Epiphyte · Fan life form · Photosynthesis · Poikilohydry · Water retention capacity

Introduction

Bryophytes are small plants with height ranged from one to a few millimeters up to a decimeter or more, with a few taxa of larger sizes. Most bryophytes have slender stems and unistratose leaves that lack a waxy cuticle (Miller 1982). As a successful group of nonvascular plants, bryophytes mostly avoid competition with tracheophytes by growing on hard, impenetrable substrata (e.g., tree bark and rocks), and tolerate frequent desiccation and nutrient-poor conditions (Bates 1998). Bryophytes are a typical group of poikilohydric plants; they usually dry out and suspend metabolism rapidly under dry conditions, but recover activity quickly when rehydrated (Proctor 2001; Green et al. 2011). Poikilohydric plants show rapid equilibration of tissue water content with the surroundings and adaptation to intermittent metabolism, which are generally optimal characteristics for diminutive Fig. 1 Photos of study site and investigated species. **a** The subtropical montane cloud forest in the Ailao Mountains; **b** abundant epiphytic bryophytes in the understory; **c** fan bryophyte *Plagiochila assamica* Steph. (Photo credit: Liang Song)



plants such as bryophytes (Proctor 2001; Proctor et al. 2007; Green et al. 2011). In contrast, homoiohydric plants maintain high internal water potentials despite a low external water availability (Proctor and Tuba 2002).

Bryophyte life forms, which can be interpreted as recurring arrangements of photosynthetic tissues that minimize evaporative water loss and maximize primary production, are physiologically important for water storage and in determining boundary-layer diffusion resistance to water loss (Proctor 1990; Bates 1998; Kürschner et al. 1999). The life form is a useful concept for the ecophysiology of bryophyte, based on their poikilohydric nature and exceptionally strong dependence upon transient external water supplies (Bates 1998). There should be an ecophysiological basis for bryophyte life forms, but relevant data are very limited. Proctor (2002, 2004b) presented ecophysiological data on two tropical African pendulous epiphytic bryophytes and two Australasian pendulous species of Weymouthia. In another study, León-Vargas et al. (2006) addressed microclimate, light adaptation and desiccation tolerance of six bryophytes of pendulous growth form in two Venezuelan cloud forests. Zotz et al. (2000) found that water relations and carbon gain of moss Grimmia pulvinata (Hedw.) Sm. were related to cushion size. Fan life forms are bryophytes with shoots rising from a vertical substratum that branch repeatedly in the horizontal plane to form flattened photosynthetic surfaces, which are exposed on all sides to ambient air (Bates 1998; Ma et al. 2009). This type of bryophyte usually "stands" on tree trunk surfaces and has a very extensive surface area in contact with the air, and they are thus well suited for intercepting water from moving air (Bates 1998; Kürschner et al. 1999; Ma et al. 2009). However, to our knowledge, detailed water relations and gas exchange characteristics of fan bryophytes and their adaptations to those particular microhabitats are still poorly understood.

The subtropical montane cloud forest (hereafter SMCF) located in the Ailao Mountain National Nature Reserve of Southwest China, also known as montane moist evergreen broadleaf forest, is generally characterized by persistent, frequent cloud cover at the canopy level (Fig. 1a). The forest harbors plentiful epiphytic bryophytes on tree trunks (Fig. 1b) (Shi and Zhu 2009; Chen et al. 2010; Song et al. 2012a; Zhang et al. 2013). The epiphytic bryoflora of the SMCF is not only remarkable in its abundance and richness of species, but also notable for the diversity of life forms represented (Ma et al. 2009; Song et al. 2011). Song et al. (2015) recorded eight life forms in the SMCF of the Ailao

Mountains and 36.2 % of the bryophytes growing on tree trunks at ~1.5 m above the forest floor showed fan forms (Fig. 1c). Considering that a life form spectrum allows one to obtain a realistic impression of the habitat (Frahm and Ohlemüller 2001), the predominance of fan life forms in the bryophyte cover on low tree trunks could indicate suitable conditions for the survival and growth of fans. This phenomenon poses the following two ecological and physiological questions. First, what are the microclimatic conditions on low tree trunks? Second, which water relations and gas exchange characteristics of fan bryophytes help them adapt to low tree trunk microhabitats?

In the present study, we quantified microclimatic characteristics of fan bryophyte microhabitats, and studied water relations (water release curves, pressure-volume relationships) and photosynthetic water and light responses of three representative fan bryophytes in a SMCF of southwest China. We address the following questions: (1) what are the water and light conditions at locations occupied by these fan bryophytes, and how do these environmental conditions influence their water and carbon balances? (2) What are the water retention and desiccation tolerance capacities of these fan bryophytes? (3) How does carbon assimilation of fan bryophytes respond to changes in shoot water content (WC) and photosynthetically active radiation (PAR)? Information regarding water retention, desiccation tolerance, and dependence of carbon assimilation on water content will further elucidate the potential performance of fan bryophytes under a predicted drier climate in the study region (Hijmans et al. 2005; Ramirez and Jarvis 2008; Fan and Thomas 2013).

Materials and methods

Study site

We conducted the study in the Xujiaba region of Yunnan Province (24°32'N, 101°01'E), in a protected section of a pristine 5100-ha SMCF in the Ailao Mountain National Nature Reserve (Fig. 1a; 23°35'-24°44'N, 100°54'-101°01'E), with altitude between 2000 and 2600 m (Wu 1983). The study site is affected by both the southern subcurrent of the western airflow from India and Pakistan and the southwest monsoon, so it experiences a striking alternation of dry and wet conditions (Qiu and Xie 1998). Mean annual temperature of the study site is 11.6 °C, with minimum in December (6.0 °C) and maximum in July (15.8 °C) (Fig. 2a). Mean annual rainfall of the study site is 1859 mm. 86 % of the rainfall occur during the rainy season (May through October), resulting in a pronounced dry period from December through April (Fig. 2a). Mean relative humidity (RH) is 82.6 %, with the minimum in



Fig. 2 a Mean monthly temperatures and rainfall, **b** foggy days and relative humidity, and **c** sunshine duration in the subtropical montane cloud forest during 2007–2013 (data from Ailaoshan Station for Subtropical Forest Ecosystem Studies, elevation 2480 m)

February (62.7 %) and maximum in July (93.3 %; Fig. 2b). Low-level cloud cover and fog are frequent (211 days per year), which compensates for a water shortage in dry months (Figs. 1a, 2b). Sunshine duration in the rainy season is much shorter than that in the dry season (Fig. 2c). The forest is primarily co-dominated by Lithocarpus hancei (Benth.) Rehder, Castanopsis rufescens (Hook.f.et Th.) Huang et Y.T.Chang, and Lithocarpus xylocarpus (Kurz) Markgr., among others (Wu 1983). The forest has four distinct strata, a canopy layer of trees (>20 m), a subcanopy layer of trees (<20 m), a shrub layer and an herb layer, interweaved with numerous lianas and epiphytes (Fig. 1b) (Qiu and Xie 1998). Detailed investigation of the vascular epiphytes and epiphytic bryophytes in this forest was carried out by Xu and Liu (2005) and Ma et al. (2009), and of the epiphytic lichens by Li et al. (2013).

Microclimatic measurements

On a triangular meteorological tower within the study area, two levels of instrument arms were mounted to measure microclimatic variables of the understory (1.5 m) and canopy (24 m). Instruments for measuring air temperature, RH (HMP45C; Vaisala Inc., Helsinki, Finland) and PAR (LQS70-10; Apogee Instruments, Inc., Logan, UT, USA) were installed at these heights, with a sampling interval of 30 min.

Measurements of water relations and gas exchange characteristics

One moss (Homaliodendron flabellatum (Sm.) Fleisch.) belonging to Neckeraceae and two liverworts [Plagiochila arbuscula (Brid. ex Lehm.) Lindenb. and Plagiochila assamica Steph.] belonging to Plagiochilaceae were selected for this study, because they are abundant and representative fan bryophytes and are easily sampled in the study area. All samples of H. flabellatum, P. arbuscula, and P. assamica were collected during the rainy season of 2012, from tree trunks at heights between 1.0 and 2.0 m above the forest floor. Samples were then transferred to the laboratory of the Xujiaba Ecological Field Station for subsequent measurements. After removing dirt and dead biomass, samples were stored outside in the shade for up to 1 week. One day prior to measurements, clean healthy shoots (terminal 1.5–2.5 cm of a stem, including the capitulum) were cut and wetted to ensure recovery of the metabolic processes. After being soaked in rainwater for 60 min so as to be fully hydrated, excess surface water was allowed to drip from the green healthy bryophyte shoots, and these were carefully blotted by placing the samples on blotting paper for 30 s.

Measurement of water release curves

As a first set, wet samples (one shoot each) were weighed using an electronic balance (AL204-IC; Mettler Toledo Instruments Ltd., Shanghai, China). Sample weights were recorded every 2 min during the first 60 min and at incremental intervals (5, 10, 30 min) over the next 260 min, until constant weights were obtained. After measurement, all samples were dried for 48 h at 70 °C to determine the dry weight (DW). Five replicates were measured for each species.

Measurement of pressure-volume curves

As a second set, wet samples (two shoots each) were placed in small polyethylene cups for ease of handling. Then, a succession of measurements of water potential at different WCs was made for each sample. Water potential was measured with the Dewpoint Water PotentiaMeter (WP4-T; Decagon Devices Inc., Pullman, WA, USA), and samples were weighed with an electronic balance (AL204-IC; Mettler Toledo Instruments Ltd., Shanghai, China). After measurement, all samples were dried for 48 h at 70 °C to determine the DW. Four replicates per species were measured.

Measurement of photosynthetic water and light response curves

As a third set, wet samples (three to four shoots each) were placed in a whole plant chamber (LI-6400-17; LI-COR, Lincoln, NE, USA) that connected to a portable infrared gas analyzer (LI-6400XT; LI-COR, Lincoln, NE, USA) to determine net photosynthetic rates (A_N) . The chamber was illuminated by placing a full spectrum light source (LI-6400-18; LI-COR, Lincoln, NE, USA) at its top. For photosynthetic water response curves, a succession of measurements was made on each blotted sample until obtaining constant weight. PAR was set to 250 μ mol m⁻² s⁻¹ (saturation for all species). After each weighing, the samples were allowed to lose up to a few milligrams of water before being resealed in the cuvette. Light response curves of bryophyte shoots were determined following 10 gradients of PAR: 200, 150, 125, 100, 80, 60, 40, 20, 10 and 0 μ mol m⁻² s⁻¹. To maintain relatively high and stable sample water content during the measurement, a piece of moist filter paper was placed in the bottom of the chamber. Empty chamber measurements with moist filter paper did not register significant CO₂ exchange signals, similar to other reports (Romero et al. 2006). The rate of CO_2 emission at zero PAR was assumed to be the respiration rate (R_d, µmol $CO_2 \text{ m}^{-2} \text{ s}^{-1}$). After all measurements were completed, samples were dried for 48 h at 70 °C to determine DW. All A_N is expressed relative to DW. Five replicates per species were measured.

Data analysis

Paired *t*-tests were conducted to compare microclimatic variables between the understory and forest canopy. Frequency distribution of RH and PAR during representative dry and rainy months (February and July, respectively) in 2013 were analyzed following León-Vargas et al. (2006). Here, we used microclimatic data from 2013 as a substitute for those in 2012 because the RH sensor at 1.5 m failed during the rainy season of the latter year, so RH data were not available then.

Evaporative water loss curves were fitted using exponential decay regressions:

$$WC = y_0 + a \cdot e^{(-b \cdot t)},\tag{1}$$

where t is evaporative time and a, b, y_0 are coefficients. Then, the initial (t = 0) WC of the samples can be calculated using ($y_0 + a$). Relative water content (RWC, %) was calculated based on

$$RWC = \frac{M - DW}{BW - DW},$$
(2)

where M, BW, DW refer to real-time, blotted, and ovendried weights, respectively.

Pressure–volume (P–V) curves were analyzed and the corresponding characteristics were calculated according to Beckett (1997). To estimate external capillary water (R_e) from a P–V curve, the water potential reciprocal ($-1/\Psi$, $-MPa^{-1}$) was plotted as a function of 1 – RWC. The straight-line parts of the P–V curves below the turgor loss point (TLP) were fitted by linear regression:

$$-1/\psi = y_0 + a \cdot (1 - RWC).$$
 (3)

Corresponding data points above the TLP were fitted by hyperbolic decay regression:

$$-1/\psi = y_0 + \frac{a \cdot b}{b+1 - RWC},\tag{4}$$

where Ψ is water potential and a, b, and y_0 are coefficients.

Turgor potential (Ψ_p , MPa) was calculated as the difference between the extrapolated linear portion of the curve and actual curve, and $\Psi_{\rm p}$ was then plotted as a function of RWC (Beckett 1997). In most of the cryptogams, $\Psi_{\rm p}$ did not decline with the initial loss of water. Water loss between 100 % RWC and RWC at which turgor began to decrease was assumed to be R_e (Beckett 1997). All RWC data were recalculated to exclude Re. The P-V curves were then replotted and all parameters recalculated. RWC and osmotic potential at TLP (RWC_{TLP} %; $\Psi_{\pi TLP}$ MPa) were calculated according to the point of intersection of beeline and hyperbola (Fig. 3a). Water potential at saturated water content ($\Psi_{\pi s}$) was calculated by the negative reciprocal of intercept of the fitted linear model (Fig. 3a). Apoplastic water (R_a) was calculated from the x-intercept (Fig. 3a), and residual water is called symplast water (R_s). Ψ_p /RWC curves were fitted using exponential regression to calculate bulk elastic modulus (ε):

$$\psi_{\rm p} = \mathbf{a} \cdot \mathbf{e}^{\mathbf{b} \cdot \mathbf{RWC}},\tag{5}$$

where RWC is the corrected relative water content and a and b are coefficients.

 ε can generally increase continuously between the TLP and full turgor (Roberts et al. 1981; Tyree and Karamanos 1981), so it can only be defined precisely for a given Ψ_p . We took values of Ψ_p and ε from the fitted exponential models, calculating ε at an arbitrary Ψ_p of 1.0 MPa (Fig. 3b) following Beckett (1997).

The relationship between A_N and WC was fitted using logistic sigmoidal regression:

$$A_{\rm N} = y_0 + \frac{a}{1 + \left(\frac{WC}{b}\right)^c},\tag{6}$$



Fig. 3 Schematic diagrams showing how parameters of water relations were calculated, based on a pressure-volume curves and b graphs of turgor pressure (Ψ_p) against relative water content (RWC) (data for Plagiochila arbuscula). TLP turgor loss point. Different symbols indicate replicates of specific species. Short dashed lines in upper graph show hyperbolic decay regression and linear regressions for curved and straight-line part of the pressure-volume curves. Short dashed curves in the lower graph show exponential regressions for Ψ_n /RWC curves. Relative water content and osmotic water potential at turgor loss (RWC_{TLP}, $\Psi_{\pi TLP}$) were calculated based on point of intersection of beeline and hyperbola. Osmotic potential at full turgor $(\Psi_{\pi s})$ was calculated by negative reciprocal of intercept of the fitted linear model. Apoplastic water (Ra) was calculated according to the x-intercept of the fitted linear model; residual water content can be used to calculate symplast water (R_s). Bulk elastic modulus (ε) was calculated from the fitted exponential models at an arbitrary $\Psi_{\rm P}$ of 1.0 MPa

where A_N is net photosynthesis rate and a, b, c, y_0 are coefficients.

Relationships between A_N and PAR were fitted by a modified rectangular hyperbola, which showed advantages over the non-rectangular hyperbola, rectangular hyperbola, and Prado–Moraes models in C₃ plants (Ye 2007; Ye and Yu 2008).

$$A_{\rm N} = a \cdot \frac{1 - b \cdot I}{1 + c \cdot I} \cdot I - R_{\rm d}, \tag{7}$$

where I is light intensity; R_d is the dark respiration rate; a, b, c are coefficients.

The main photosynthetic characteristics were calculated according to Ye (2007); Ye and Yu (2008). These included the light-saturated net photosynthetic rate (A_{nmax} ; µmol $CO_2 m^{-2} s^{-1}$), light saturation point (I_{sat} : light intensity on the light curve beyond, which further increases will not augment the rate of photosynthesis; µmol $m^{-2} s^{-1}$), and light compensation point (I_c : light intensity on the light curve where the rate of photosynthesis exactly matches the rate of respiration; µmol $m^{-2} s^{-1}$).

All data were submitted to normality and homoscedasticity tests before applying further statistical methods. Differences in water relations and gas exchange characteristics were compared using one-way ANOVA with Tukey's post hoc tests via SPSS 16.0 software (SPSS Inc., Chicago, IL, USA). All regressions and figures were made using Sigma-Plot 12.5 (Systat Software Inc., San Jose, CA, USA).

Results

Microclimatic conditions

Compared with the forest canopy, significantly lower air temperatures (paired *t*-test, t = 4.21, P = 0.001) and PAR (t = 2.177, P = 0.052), but higher RH (t = 9.027, P < 0.001) were detected in the understory (Fig. 4). The detailed frequency distribution of RH indicated that the understory was a very moist microhabitat, especially during the rainy season (with 93.7 % of RH data recorded in July at more than 90 %; Table 1). In addition, the frequency distribution of PAR further confirmed very low light conditions in the understory (with 56.3 % of the PAR data recorded in February less than 1 µmol m⁻² s⁻¹; Table 2).

Water relations and gas exchange of fan bryophytes

There was a reasonably good exponential decay regression fit between the WC of shoots and time for all three experimental species, with r^2 values 0.98–0.99 (Fig. 5). Average initial water contents of blotted samples of *H. flabellatum*, *P. arbuscula*, and *P. assamica* were 318.8, 402.6, and 451.7 %, respectively. All bryophyte samples lost water very quickly, and all dried up within an hour after being exposed to air with no water supply (Fig. 5). The measured final WCs were very low, with averages 11.6, 12.3, and 11.8 % for *H. flabellatum*, *P. arbuscula*, and *P. assamica*, respectively (model-fit values were much smaller). Further ANOVA results indicated no difference in average final WC (after air drying for 320 min) among different bryophyte species ($F_{2.14} = 0.055$, P = 0.946).



Fig. 4 Comparison of mean **a** air temperature, **b** relative humidity, and **c** photosynthetic active radiation between understory at ~1.5 m and forest canopy at ~24 m in the subtropical montane cloud forest during 2007–2013 (data from Ailaoshan Station for Subtropical Forest Ecosystem Studies, elevation 2480 m)

P–V curves are shown in Fig. 6. According to ANOVA results, significant differences were detected in R_s ($F_{2,11} = 16.330$, P = 0.001), $\Psi_{\pi TLP}$ ($F_{2,11} = 17.992$, P = 0.001), and $\Psi_{\pi s}$ ($F_{2,11} = 7.839$, P = 0.011), but no significant differences in R_a ($F_{2,11} = 1.791$, P = 0.221), RWC_{TLP} ($F_{2,11} = 0.334$, P = 0.724), and ε ($F_{2,11} = 0.167$, P = 0.849) were detected among the three studied bryophytes (Table 3). $\Psi_{\pi TLP}$ ranged from -4.2 MPa in *H. flabellatum* to -2.6 MPa in *P. assamica*, and $\Psi_{\pi s}$ from -2.5 to -1.6 MPa. The moss *H. flabellatum* had lower R_s , $\Psi_{\pi TLP}$, and $\Psi_{\pi s}$ (Table 3) compared with the two liverworts, and ε of the bryophytes was 4.2–4.5 (Table 3).

The response of A_N to WC in the three fan bryophyte species showed similar trends, rising rapidly with increasing WC (Fig. 7). The fitted sigmoidal models (logistic, four parameters) for A_N had r^2 from 0.85 to 0.95 (Fig. 7). A_N increased with WC, reaching 0-nmol CO₂ g⁻¹ DW s⁻¹ (respiration rates equal to photosynthetic rate) as WC approached 80 % DW (82.7, 104.1, and 81.7 % for *H. flabellatum*, *P. arbuscula*, and *P. assamica*, respectively).

Light response of the photosynthetic rate in the three bryophyte species was well described by a modified rectangular hyperbolic function (r^2 between 0.93 and 0.98; Fig. 7). Significant differences were detected in A_N ($F_{2,14} = 6.734$, P = 0.011) and R_d ($F_{2,11} = 5.964$,

Table 1 Percentage frequency distribution of relative humidity (RH)during February and July, 2013 at different heights in the subtropicalmontane cloud forest

RH interval (%)	February		July	
	1.5 m	24 m	1.5 m	24 m
90–100	0.7	4.3	93.7	84.9
80–90	8.9	5.0	5.8	9.8
70-80	14.4	8.3	0.3	3.5
60-70	18.1	13.3	0.1	0.9
50-60	15.8	19.9	0.2	0.2
<50	42.3	49.3	0.0	0.8

Table 2 Percentage frequency distribution of photosynthetically active radiation (PAR) during February and July 2013, at different heights in the subtropical montane cloud forest

PAR interval	February		July		
	1.5 m	24 m	1.5 m	24 m	
>100	1.0	33.6	0.0	15.2	
50-100	2.2	4.5	0.3	14.4	
10-50	14.1	7.8	12.3	18.0	
1-10	26.3	3.9	34.2	7.0	
<1	56.3	50.3	53.1	45.4	



Fig. 5 Water release curves of **a** *Homaliodendron flabellatum*, **b** *Plagiochila arbuscula*, and **c** *P assamica* (N = 5). *WC* water content

P = 0.016), but not in I_c ($F_{2, 14} = 2.912$, P = 0.093) and I_s ($F_{2, 14} = 0.038$, P = 0.963). Light compensation points (all less than 5 µmol m⁻² s⁻¹; Table 4) and light saturation points (all less than 150 µmol m⁻² s⁻¹; Table 4) were low for all three species. The moss *H. flabellatum* showed lower maximum A_N and R_d relative to the two liverworts (Table 4).

Discussion

Microclimatic conditions of the subtropical forest

Microclimate describes well the habitat under which epiphytic bryophytes on trees survive and grow in a SMCF. Similar to León-Vargas et al. (2006), measurements at 1.5 m above the soil surface showed quite high RH but very low temperature and light levels (Fig. 4). These results suggest that epiphytic bryophytes in the understory may remain continuously moist for days during the rainy season. RH was greater than 80 % for most of the time during that season. Even during the driest month, RH was near 60 % (Fig. 4). Frequent and persistent fog provides an important water source, which can alleviate water shortages for epiphytic bryophytes, especially during the dry season (Fig. 2b). However, bryophytes experience dry periods during the dry season and extreme drought in dry years (Song et al. 2012b). In addition, light is limiting for most plants, including bryophytes, in the deep shade of the understory (mean PAR at ~1.5 m above the soil surface was 7.8 μ mol m⁻² s⁻¹; Fig. 4). However, the deep shade minimizes evaporation of poikilohydric plants such as filmy ferns and bryophytes, and nullifies the advantages of typical vascular plants with ventilated mesophyll and stomatal control over water loss (Proctor 2012).

Water relations of fan bryophytes

Because fan bryophytes usually stand on tree trunks, they have very extensive surface areas in contact with air. However, their structure is like a double-edged sword. On one hand, they are better suited for intercepting water from moving air than many other bryophytes (Bates 1998; Kürschner et al. 1999). On the other hand, with their loose and open structures, fan bryophytes inevitably have weak resistance to water loss, owing to limited external capillary space (Bates 1998). Our results show that the three fan bryophytes lost most of their free water within 1 h when exposed to air without moisture supply, suggesting weak water retention capacity of these Fig. 6 Pressure–volume curves (*left panel*) and graphs of turgor pressure against relative water content (*right panel*) of **a**, **b** *H*. *flabellatum*, **c**, **d** *P*. *arbuscula*, and **e**, **f** *P*. *assamica* (N = 4). Abbreviations and symbols are in Fig. 3



Table 3 Parameters of water relations in (a, b) Homaliodendron flabellatum, (c, d) Plagiochila arbuscula, and (e, f) P. assamica (N = 4)

Species	R _s (% DW)	R _a (% DW)	RWC _{TLP} (%)	$\Psi_{\pi TLP}$ (MPa)	$\Psi_{\pi s}$ (MPa)	ε
Homaliodendron flabellatum	$112.0\pm9.1a$	$19.6 \pm 3.3a$	$64.3 \pm 3.7a$	-4.2 ± 0.3 a	$-2.5\pm0.2a$	$4.2 \pm 0.3a$
Plagiochila arbuscula	$194.5\pm13.1\mathrm{b}$	$19.5\pm5.8a$	$67.4\pm2.2a$	$-2.6\pm0.2\mathrm{b}$	$-1.7\pm0.1\mathrm{b}$	$4.5\pm0.5a$
P. assamica	$173.4\pm9.2\text{b}$	$10.2 \pm 2.0a$	$63.9\pm3.8a$	$-2.6\pm0.2\text{b}$	$-1.6\pm0.2\text{b}$	$4.5\pm0.6a$

Values are mean \pm SE. Different lowercase letters indicate significant difference for $\alpha = 0.05$

bryophytes. The high abundance of fan bryophytes with low water retention capacity in the SMCF (Ma et al. 2009) is likely facilitated by the heavy annual precipitation (1859 mm), high average RH (82.6 %), and frequent and long duration of fog (211 foggy days annually; Fig. 2), which may provide a continuous water source for the bryophytes to intercept. The fog could be a very important water source for fans, especially in the dry season (Fig. 2b). The present study provides some of the scarce empirical evidence of the low water retention capacity of fan bryophytes and implies that these organisms are often associated with continuous water supply from frequent rain, clouds or fogs. The water relations characteristics of fan bryophytes in this study are generally consistent with those of Santarius (1994), Beckett (1997), Proctor et al. (1998), and Proctor (1999). Therefore, our results add evidence for the generally mesic pattern of physiological adaption of bryophytes. Nevertheless, fan bryophytes show tolerance to desiccation, which may occur at any time of year (Proctor 2004a; León-Vargas et al. 2006). ε is of ecophysiological interest in plants subject to periodic water stress. In this study, generally low ε (4.2–4.5) and thus high cell elasticity were detected for the fan bryophytes relative to other studies (9–34) (Beckett 1997). This could be because fan bryophytes need high cell Fig. 7 Photosynthetic water and light response curves of **a**, **b** *H*. *flabellatum*, **c**, **d** *P*. *arbuscula*, and **e**, **f** *P*. *assamica* (N = 5). Symbol shapes refer to different samples. Dotted lines in figures indicate zero net photosynthetic rates (A_N). WC and I in figures refer to water content and light intensity, respectively



Table 4 Light saturation point (I_s), light compensation point (I_c), maximum net photosynthetic rate (A_{Nmax}), and dark respiration (R_d) derived from photosynthetic light response curves of (a) *H. flabellatum*, (b) *P. arbuscula*, and (c) *P. assamica* (N = 5)

Species	$I_{s} (\mu mol m^{-2} s^{-1})$	$I_c (\mu mol \; m^{-2} \; s^{-1})$	$A_{Nmax} (nmol CO_2 g^{-1} DW s^{-1})$	$R_d \text{ (nmol CO}_2 \text{ g}^{-1} \text{ DW s}^{-1}\text{)}$
Homaliodendron flabellatum	$101.9 \pm 8.5a$	$3.6 \pm 0.3a$	$17.1 \pm 1.2a$	$7.5\pm0.9a$
Plagiochila arbuscula	$104.5\pm4.9a$	$4.2 \pm 0.3a$	$20.8 \pm 0.5 \mathrm{b}$	9.8 ± 0.7 ab
P. assamica	$103.3\pm6.9a$	$4.7 \pm 0.3a$	$21.7\pm1.0b$	$10.9 \pm 0.4 \mathrm{b}$

Values are mean \pm SE. Different lowercase letters indicate significant difference for $\alpha = 0.05$

wall elasticity to tolerate desiccation during dry periods, owing to their weaker water retention capacity. Additionally, the osmotic potentials at turgor loss point of the fan bryophytes were lower than co-occurring angiosperm species (Zhang YJ, unpublished data), suggesting greater drought tolerance of fan bryophytes compared with angiosperms. Thus, fan bryophytes have weak water retention abilities but high desiccation tolerance, for surviving dry periods. Further experiments should be done to explore the desiccation tolerance of different life form bryophyte species in various microhabitats.

Gas exchange of fan bryophytes

Net photosynthetic rates of fan bryophytes are strongly dependent on WC, which increases sigmoidaly with increasing WC. A decline in photosynthetic rate at high WC has been detected in some bryophyte species (Dilks and Proctor 1979), suggesting depression of carbon gain at such WC. However, this was not observed for any of the three bryophyte species in the present study. These fan bryophytes may have structural or physiological adaptations to facilitate gas exchange at high WC during the rainy season. Nevertheless, the WC of the blotted bryophyte materials in this study (with maximum <500 %; see Fig. 7) did not reach the maximum WC of the saturated bryophyte materials, which was between 700 and 2000 % (Dilks and Proctor 1979).

It has been reported that the light compensation points of 18 tropical bryophyte species were from 22 μ mol m⁻² s⁻¹ in Octoblepharum pulvinatum (Dozy & Molk.) Mitt. to 159 µmol m⁻² s⁻¹ in Frullania mirabilis J.B. Jack & Stephani (Wagner et al. 2013). In addition, Marschall and Proctor (2004) found that the light saturation points of 39 mosses and 16 liverworts varied from 79 μ mol m⁻² s⁻¹ in Jubula hutchinsiae (Hook.) Dumort. to 2549 μ mol m⁻² s⁻¹ piliferum Hedw., with Polytrichum median in 583 μ mol m⁻² s⁻¹ for mosses and 214 μ mol m⁻² s⁻¹ for liverworts. The latter may be overestimated because the light saturation points were mainly taken from light curves of relative electron transport rate (not CO₂ exchange), based on measurements of (modulated) chlorophyll florescence (Marschall and Proctor 2004). In our study, we measured light-response curves of gas exchange for three fan bryophyte species and found relatively small values of light compensation points (<5 μ mol m⁻² s⁻¹; Table 4) and light saturation points (<105 μ mol m⁻² s⁻¹; Table 4). Similarly, some filmy fern species from comparable habitats and of comparable life forms show comparable photosynthetic light saturation points (Proctor 2012). Low light compensation and saturation points are adaptations to low light levels.

Interspecific comparisons

It had been repeatedly confirmed that the ratio of species richness of liverworts to mosses correlate well with water availability, because liverworts usually favor wet habitats (Wolf 1993; Frahm and Ohlemüller 2001). For example, there was a distinct increase of liverworts with increasing water input from eastern to western Europe (Duell 1984). In addition, liverworts accounted for 80 % of bryophytes in the Chocó region, one of the heaviest precipitation regions with nearly 15,000 mm $vear^{-1}$ (Frahm 1994). Our results provide a physiological explanation for these patterns. According to P–V relationships, the moss (H. flabellatum) in our study had higher osmoregulation capacity (more negative $\Psi_{\pi TLP}$ and $\Psi_{\pi s}$), and therefore greater desiccation tolerance than the liverworts (P. arbuscula and P. assam*ica*). This suggests that liverworts may have less tolerance to drought compared with mosses. Further study is needed to confirm this hypothesis, considering that very few species have been included in the present study.

Ecological implications

Our results suggest that net photosynthesis rates of fan bryophytes are greatly dependent on WC. Because fan bryophytes lost most of their free water within 1 h in the dry air without water supply (Fig. 2), the implication is that the transition from a hydrated, photosynthetically active state to a dry, inactive state is rapid. In other words, fan bryophytes dry up and suspend photosynthesis quickly when water is unavailable. These fan bryophytes developed desiccation tolerance, which can be considered a special adaptive strategy to unpredictable dry periods or extreme drought in moist cloud forests (Proctor 2000, 2001). Considering abundant desiccation-tolerant epiphytic bryophytes investigated in SMCFs (Ma 2009; Song et al. 2011), we suggest that desiccation-tolerant bryophytes are among the most conspicuously successful plants in habitats with reasonably regular moisture supply.

Considering that carbon gain and growth are restricted to periods of sufficient hydration, capturing and preserving water are thus suggested to be crucial adaptations for bryophytes (Norris 1990; Wagner et al. 2013). The low water retention capacity and strong dependence of net photosynthesis on the WC of fan bryophytes thus imply a high risk of carbon imbalance under dry conditions. This situation would be exacerbated by the fact that faster drying could lead to more frequent rewetting, causing carbon loss through carbohydrate leaching (Coxson et al. 1992). The study area of Yunnan is currently experiencing drought (Fan and Thomas 2013). There was a severe drought from the end of 2009 to early 2010, which can be considered the worst over the past century (Qiu 2010; Wang 2010). This drought has been proven to result in remarkably reduced rates of growth and detrimental effects on the health of several epiphytic bryophytes (Song et al. 2012b). Since climate conditions in the SMCF are predicted to become drier by the 2050s (Hijmans et al. 2005; Ramirez and Jarvis 2008), the results of this study, together with those of a previous transplantation experiment (Song et al. 2012b), suggest a dim future for epiphytic bryophytes.

Conclusions

Our results suggest that fog helps maintain relatively high water availability during most of the year for tree trunk fan bryophytes in an Asian subtropical cloud forest. The fan bryophyte species studied have limited water retention capacities, but have developed relatively high cell wall elasticity and osmoregulation capacity so as to tolerate desiccation. In addition, relatively low light saturation and compensation points may help them tolerate shaded tree trunk microhabitats. The carbon gain of these fan bryophytes is highly water-dependent. Therefore, fan bryophytes can flourish on tree trunks in the subtropical cloud forest because of heavy annual precipitation, high average relative humidity, and frequent and long duration of fog, which provide continuous water supplies for them to intercept. However, the low water retention capacity and strong dependence of net photosynthesis on the water content of fan bryophytes suggest a high risk of carbon imbalance for them under increasing severity and frequency of drought. Therefore, this study points to an uncertain future for climate-sensitive epiphytic bryophytes under ongoing climatic change.

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