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Dysoxylentin A, the first 21-nortriterpenoid bearing a 2-(propan-2-ylidenyl)furan-3(2H)-one, from Dysoxylum lenticellatum

Ting Tang^{a,d}, Shang-Gao Liao^{b,*}, Zhi Na^a, Yan Li^c, You-Kai Xu^{a,*}

^a Key Laboratory of Tropical Forest Ecology, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Mengla 666303, Yunnan, China ^b Guiyang Medical University, Guiyang 550004, Guizhou, China

^c State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, Yunnan, China ^d Graduate University of Chinese Academy of Sciences, Beijing 100049, China

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Introduction

The genus Dysoxylum (Meliaceae) comprises about 200 species, which are mainly distributed in India, Southeast Asia, and Polynesia. Among these species, fourteen are distributed in China and ten have been found to grow in Yunnan Province.¹ Many species have been traditionally used as medicines by the indigenous people in Fiji, Papua New Guinea, and New Zealand to relieve fever, convulsions, hemorrhage, rigid limbs, and facial distortion in children.² Phytochemical investigations of more than twenty species in this genus have led to the isolation of a number of structurally diverse and biologically active compounds including cytotoxic alkaloids,³ antifeeding limonoids,⁴ cytotoxic diterpenoids,⁵ anti-leukemic triterpenoid glucosides,⁶ antibacterial and cytotoxic trit-erpenoids,⁷ sesquiterpenoids,^{2b,8} oxyneolignans,⁹ bioflavonoids,¹⁰ and a sulfur-containing compound.¹¹ Dysoxylum lenticellatum is a plant endemic to southwest Yunanan province. Previous investigation of the twigs and leaves of D. lenticellatum has revealed the presence of triterpenoids, diterpenoids, and ceramids in this plant.¹² As part of our ongoing project aiming at structurally diverse natural products, the stems of D. lenticellatum have been chemically investigated. As a result, dysoxylentin A, a new 21nortriterpenoid with a 2-(propan-2-ylidenyl)furan-3(2H)-one unit, was isolated. In this Letter, we report the isolation, structural

ABSTRACT

Dysoxylentin A (1), the first 21-nortriterpenoid bearing a 2-(propan-2-ylidenyl)furan-3(2H)-one functional group was isolated from the stem of *Dysoxylum lenticellatum*. Its structure was elucidated by extensive spectroscopic analysis. A plausible biosynthetic pathway was postulated. Dysoxylentin A exhibited selective cytotoxicity against HL-60 tumor cell line.

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elucidation, and biological activity of the new compound. A plausible biosynthetic pathway was also postulated.



Dysoxylentin A (**1**)¹³ was isolated as a white amorphous powder. Its molecular formula was determined to be $C_{29}H_{42}O_3$ by the HREIMS ion at *m*/*z* 438.3138 (calcd 438.3134) showing 9° of unsaturation. The UV spectrum of dysoxylentin A (**1**) showed absorption maxima at 204, 260, and 307 nm, suggesting the presence of a conjugated system. The IR spectrum indicated the presence of hydroxyl (3440 cm⁻¹), conjugated carbonyl (1707 cm⁻¹) and double bond (1641, 1591 cm⁻¹) groups. In the ¹H spectrum (Table 1), signals for seven quaternary methyls [$\delta_{\rm H}$ 0.79 (s, 6H), 0.91, 0.93, 1.06, 1.98, 2.26], two olefinic protons [$\delta_{\rm H}$ 5.30 (t-like) and $\delta_{\rm H}$ 5.63 (s)], one oxygenated methine proton [$\delta_{\rm H}$ 3.47 (br s)], and one methine



^{*} Corresponding authors. Tel.: +86 691 8715910/851 6908468 8712; fax: +86 691 8715070.

E-mail addresses: lshangg@163.com (S.-G. Liao), xyk@xtbg.org.cn (Y.-K. Xu).

Table 1 1 H-(400 MHz) and 13 C-(100 MHz) NMR data of 1 in CDCl₃

11-(400 WITZ) and		
No.	$\delta_{\rm H}$ (mult, J, in Hz)	δ_{C}
1α	1.48 ^{NR}	31.1
1β	1.39 ^{NR}	
2α	1.91 ^{NR}	25.3
2β	1.62 ^{NR}	
3	3.47 (br s)	76.2
4		37.4
5	1.78 ^{NR}	44.5
6α	1.93 ^{NR}	23.7
6β	2.04 ^{NR}	
7	5.30 (t-like)	119.0
8		144.6
9	2.35 (m)	48.3
10		34.8
11α	1.66 ^{NR}	17.2
11β	1.60 ^{NR}	
12α	1.96 ^{NR}	31.3
12β	1.70 ^{NR}	
13		46.2
14		50.8
15α	1.75 ^{NR}	34.0
15β	1.64 ^{NR}	
16α	2.07 ^{NR}	23.9
16β	1.89 ^{NR}	
17	2.91 (t, 9.0 Hz)	48.5
18	0.79 (3H, s)	23.1
19	0.79 (3H, s)	12.9
20		184.0
22	5.63 (s)	107.3
23		187.2
24		144.9
25		130.2
26	1.98 (3H, s)	19.9
27	2.26 (3H, s)	17.0
28	0.91 (3H, s)	21.7
29	0.93 (3H, s)	27.7
30	1.06 (3H, s)	27.1

^{NR} Indicates that the proton signal was either partially overlapped or unresolved.

 $[\delta_{\rm H} 2.91 (t, J = 9.0 \text{ Hz})]$ adjacent to a methylene were easily identified. The ¹³C NMR (Table 1) and DEPT spectra exhibited 29 carbon resonances, which corresponded to seven methyls, seven methylenes, six methines (one oxygenated and two olefinic), and nine quaternary carbons (one ketone and four olefinic). The above functionalities accounted for 4° of unsaturation, the remaining 5° of unsaturation suggested the presence of a pentacyclic system in dysoxylentin A. The aforementioned data were very different from those of the compounds isolated from the same plant,¹² suggesting that dysoxylentin A (1) possibly possessed a new structural skeleton. Extensive 2D NMR experiments (¹H–¹H COSY, HSQC, HMBC, and ROESY) were thus performed and the structure of 1 was established. All proton signals were unambiguously assigned to their respective carbon atoms by using HSQC experiment. Four spin



Figure 1. Key HMBC and ¹H-¹H COSY correlations of 1.

systems drawn with bold bonds were established on the basis of ¹H–¹H COSY spectrum (Fig. 1). Further HMBC correlation analysis (Fig. 1) led to the assembly of the four subunits with the quaternary carbons and other functionalities. In particular, HMBC correlations of H₃-19 to C-1, C-5, C-9, and C-10, of H₃-18 to C-12, C-13, C-14, and C-17, of H₃-28/H₃-29 to C-3, C-4, and C-5, of H₃-30 to C-8, C-13, C-14, and C-15, and of H-7 to C-14 indicated that dysoxylentin A (1) possessed a carbon skeleton with an ABCD-ring system same as that of a 3-hydroxy tirucallane-type triterpenoid.^{12b,14} The tetracyclic feature of the above structure required the presence of an additional ring in the rest part of dysoxylentin A. The remaining ¹H- and ¹³C-NMR data for one ketone ($\delta_{\rm C}$ 187.2), one trisubstituted double bond ($\delta_{\rm H}$ 5.63, $\delta_{\rm C}$ 107.3; $\delta_{\rm C}$ 130.2, 144.9, 184.0), and two olefinic methyls ($\delta_{\rm H}$ 1.98, 2.26, $\delta_{\rm C}$ 17.0, 19.9) suggested the presence of a 2-(propan-2-ylidenvl)furan-3(2H)-one-5-vl group in dvsoxvlentin A.¹⁵ HMBC correlations from H-22 to C-17, C-20, C-23, and C-24, and from H₂-26/ 27 to C-23, C-24, and C-25 further confirmed this conclusion, while correlations from H-17 to C-20 and C-22 indicated that the group was located at C-17. Thus, the planar structure of dysoxylentin A was established as shown in Figure 1.

The relative configuration of dysoxylentin A (1) was determined by the coupling patterns and ROESY correlations. The broad singlet of H-3 suggested the β -orientation of H-3,^{3a} which was further confirmed by the ROESY correlation of H-3 with H₃-28. The α -orientations of H-5 and H-9 were deduced from



Figure 2. Key ROESY correlations of 1.



Scheme 1. Plalusible biosynthetic pathway proposed for 1.

the correlations of H-5/H₃-29, H-9/H₃-18. The β -orientation of H-17 was determined by the correlations of H-17/H₃-30. Consequently, the relative stereochemistry of dysoxylentin A was determined as depicted (Fig. 2).

To the best of our knowledge, this is the first 21-nortirucallane triterpenoid that bears a 2-(propan-2-ylidenyl)furan-3(2H)-one-5yl group, which has never been reported to be present in nature. Although two endophytic fungi¹⁵ and one plant¹⁶ were reported to produce natural products with such a structural unit, this unit was only present as a part of benzofuranone¹⁵ or as a simple aglycone.¹⁶ The biogenetic origin of dysoxylentin A (1) could be traced back to 24,25-epoxytirucall-7-ene-3,23-dione (2),^{3a} a tirucallane triterpenoid also isolated in this investigation (Scheme 1). Reduction of the 3-ketone group of **2** to a α -hydroxyl would yield I. subsequent dehvdration between H-21 and OH-20 of the hydroxylated product of which would afford III. Oxidative cleavage of the $\Delta^{20(21)}$ double bond followed by reduction of the newly generated ketone group would give V, nucleophilic attack of the 21-OH of which on C-24 of the 24,25-epoxide would furnish the dihydrofuranone derivative VI. Dehydration between H-24 and OH-25 of VI followed by hydroxylation of H-22 would yield VIII. In the last step, dehydration would occur between the OH-22 and H-20 to give 1.

Dysoxylentin A (1) was assayed for its cytotoxicity against five human tumor cell lines (human myeloid leukemia HL-60, hepatocellular carcinoma SMMC-7721, lung cancer A-549, breast cancer MCF-7, and colon cancer SW480) by using MTT method¹⁷ with cisplatin as the positive control (IC₅₀ 1.94 μ M against HL-60). Dysoxylentin A showed selective activity (IC₅₀ 34.61 μ M) against the HL-60 cell line but did not show any obvious cytotoxicity (IC₅₀ >100 μ M) against the other cell lines.

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Supplementary data

Supplementary data (experimental details, NMR spectrum, MS data) associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2011.12.109.

References and notes

- 1. Yunnan Institute of Botany. In *Flora Yunnanica*. Science Press, Beijing, 1977, Tomus 1, p. 243.
- (a) Aalbersberg, W.; Singh, Y. Phytochemistry **1991**, 30, 921–926; (b) Russell, G. B.; Hunt, M. B.; Bowers, W. S.; Blunt, J. W. Phytochemistry **1994**, 35, 1455–1456; (c) Chen, J. L.; Kernan, M. R.; Jolad, S. D.; Stoddart, C. A.; Bogan, M.; Cooper, R. J. Nat. Prod. **2007**, 70, 312–315.
- (a) Zhang, X. Y.; Li, Y.; Wang, Y. Y.; Cai, X. H.; Feng, T.; Luo, X. D. J. Nat. Prod. 2010, 73, 1385–1388; (b) Ismail, I. S.; Nagakura, Y.; Hirasawa, Y.; Hosoya, T.; Lazim, M. I. M.; Lajis, N. H.; Shiro, M.; Morita, H. J. Nat. Prod. 2009, 72, 1879–1883.
- Luo, X. D.; Wu, S. H.; Wu, D. G.; Ma, Y. B.; Qi, S. H. Tetrahedron 2002, 58, 7797– 7804.
- G. Duh, C. Y.; Wang, S. K.; Cheng, I. S. J. Nat. Prod. 2000, 63, 1546–1547; (b)
 Fujioka, T.; Yamamoto, M.; Kashiwada, Y.; Fujii, H.; Mihashi, K.; Ikeshiro, Y.;
 Chen, I. S.; Lee, K. H. Bioorg. Med. Chem. Lett. 1998, 8, 3479–3482.
- (a) Kashiwada, Y.; Fujioka, T.; Chang, J. J.; Chen, I. S.; Mihashi, K.; Lee, K. H. J. Org. Chem. **1992**, 57, 6946–6953; (b) Kashiwada, Y.; Fujioka, T.; Mihashi, K.; Chen, I. S.; Katayama, H.; Ikeshiro, Y.; Lee, K. H. J. Nat. Prod. **1997**, 60, 1105–1114.
- (a) He, X. F.; Wang, X. N.; Yin, S.; Dong, L.; Yue, J. M. Bioorg. Med. Chem. Lett. 2011, 21, 125–129; (b) He, X. F.; Wang, X. N.; Yin, S.; Dong, L.; Yue, J. M. Eur. J. Org. Chem. 2009, 4818–4824; (c) He, X. F.; Wang, X. N.; Gan, L. S.; Yin, S.; Dong, L.; Yue, J. M. Org. Lett. 2008, 10, 4327–4330; (d) Ismail, I. S.; Nagakura, Y.; Hirasawa, Y.; Hosoya, T.; Lazim, M. I. M.; Lajis, N. H.; Morita, H. Tetrahedron Lett. 2009, 50, 4830–4832.
- Mulholland, D. A.; Iourine, S.; Taylor, D. A. H. Phytochemistry 1998, 47, 1421– 1422.
- 9. Xie, B. J.; Yang, S. P.; Yue, J. M. Phytochemistry 2008, 69, 2993-2997.
- 10. He, K.; Timmermann, B. N.; Aladesanmi, A. J.; Zeng, L. *Phytochemistry* **1996**, *42*, 1199–1201.
- 11. Jogia, M. K.; Andersen, R. J.; Mantus, E. K.; Clardy, J. Tetrahedron Lett. 1989, 30, 4919–4920.
- (a) Qi, S. H.; Wu, D. G.; Zhang, S.; Luo, X. D. Z. *Naturforsch. B* 2003, 58, 1128–1132; (b) Huang, H. L.; Wang, C. M.; Wang, Z. H.; Yao, M. J.; Han, G. T.; Yuan, J. C.; Gao, K.; Yuan, C. S. J. Nat. Prod. 2011, 74, 2235–2242.
- 13. Dysoxylentin A (1): white amorphous powler; $[\alpha]_D^{5.8} = -2.42$ (c 0.0015, CH₃OH); UV (CH₃OH) λ_{max} (log ε) 307 (3.53), 260 (3.86), 204 (3.89) nm; CD (CH₃OH) λ_{max} ($\delta\varepsilon$) 213 (-6.07), 259 (+11.68), 290 (-0.92), 340 (+1.19); IR (KBr) ν_{max} 3440, 2950, 1707, 1641, 1591, 1459, 1384, 1135 cm⁻¹; ¹H and ¹³C NMR data, see Table 1; EIMS *m*/z (relative intensity) 438 [M]⁺ (58), 423 (36), 405 (49), 321 (20), 298 (45), 255 (20), 187 (23), 150 (100), 83 (71), 69 (59), 55 (49); HREIMS *m*/z 438.3138 (calcd for C₂₉H₄₂O₃, 438.3134).
- (a) Mohamad, K.; Martin, M. T.; Litaudon, M.; Gaspard, C.; Sévenet, T.; Païs, M. *Phytochemistry* **1999**, *52*, 1461–1468; (b) Liu, H. M.; Heilmann, J.; Rali, T.; Sticher, O. J. Nat. Prod. **2001**, *64*, 159–163; (c) Luo, X. D.; Wu, S. H.; Ma, Y. B.; Wu, D. G. *Phytochemistry* **2000**, *54*, 801–805.
- (a) Lu, Z. Y.; Wang, Y.; Miao, C. D.; Liu, P. P.; Hong, K.; Zhu, W. M. J. Nat. Prod. 2009, 72, 1761–1767; (b) Ding, G.; Zheng, Z. H.; Liu, S. C.; Zhang, H.; Guo, L. D.; Che, Y. S. J. Nat. Prod. 2009, 72, 942–945; (c) Ogawa, A.; Murakami, C.; Kamisuki, S.; Kuriyama, I.; Yoshida, H.; Sugawara, F.; Mizushina, Y. Bioorg. Med. Chem. Lett. 2004, 14, 3539–3543.
- Comte, G.; Allais, D. P.; Chulia, A. J.; Vercauteren, J.; Delage, C. *Phytochemistry* 1996, 41, 1329–1332.
- 17. Mosmann, T. J. Immunol. Methods 1983, 65, 55-63.