












RESEARCH ARTICLE OPEN ACCESS

Identification of Priority Areas for Conservation of Threatened Agamid Lizards of Sri Lanka Using Species Distribution Modeling

Iresha Wijerathne^{1,2}  | Suranjan Karunarathna³  | Thilina Sursinghe⁴  | Dulan R. Vidanapathirana⁵  |
Kanishka Ukuwela⁶  | Sriyani Wickramasinghe⁶  | Chaya Sarathchandra⁶  | Jagath Gunatilake^{2,7}  |
Aiwu Jiang¹  | Eben Goodale⁸  | Suranjan Fernando⁹ 

¹Guangxi Key Laboratory of Forest Ecology and Conservation, College of Forestry, Guangxi University, Nanning, China | ²Postgraduate Institute of Science, University of Peradeniya, Peradeniya, Sri Lanka | ³Nature Exploration & Education Team, Moratuwa, Sri Lanka | ⁴Department of Biological Sciences, Bridgewater State University, Bridgewater, Massachusetts, USA | ⁵Herpetological Foundation of Sri Lanka, Wattala, Sri Lanka | ⁶Faculty of Applied Sciences, Rajarata University of Sri Lanka, Mihintale, Sri Lanka | ⁷Department of Geology, Faculty of Science, University of Peradeniya, Peradeniya, Sri Lanka | ⁸Department of Health and Environmental Sciences, Xi'an Jiaotong-Liverpool University, Suzhou, China | ⁹Freelance Biodiversity Specialist in Kandy, Kandy, Sri Lanka

Correspondence: Iresha Wijerathne (ireshaw888@gmail.com) | Aiwu Jiang (aiwuu@gxu.edu.cn)

Received: 14 October 2024 | **Revised:** 15 March 2025 | **Accepted:** 24 March 2025

Editor-in-Chief & Handling Editor: Ahimsa Campos-Arceiz

Funding: Suranjan Karunarathna appreciates the Rufford Foundation for partially funding for fieldwork (Grant 23951-1).

Keywords: agamid lizards | conservation priority | land-use patterns | protected areas | species diversity | threatened species

ABSTRACT

To ensure the survival of threatened species, identifying biodiversity hotspots is essential for determining where and how conservation should be prioritized. Sri Lanka along with Western Ghats of India is a global biodiversity hotspot, with reptiles among its most threatened taxa. Agamid lizards in Sri Lanka are both threatened and evolutionarily distinct, with 19 of 22 species endemic to the island. We assessed the distribution of 14 threatened agamid lizard species using Species Distribution Modeling (SDM). The generated distribution maps were used to identify highly diverse target areas for agamid conservation, both within and outside protected areas. We assigned conservation priority ranks for species using different criteria such as IUCN status, the number of occurrence records, and the size of the estimated range. Our results indicated that agamid hotspots are primarily concentrated in the wet zone of Sri Lanka; specifically, Sinharaja Forest Reserve, the Peak Wilderness Sanctuary, and the Knuckles Mountain Range emerged as high priority conservation targets. Although all species targeted in our SDM had some portion of their range within protected areas, over 40% of high-priority agamid habitats remain unprotected. We advocate for the expanded use of SDM for hotspot identification and conservation planning, particularly for threatened and endangered species with poorly documented population status and geographic distribution.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Integrative Conservation* published by John Wiley & Sons Australia, Ltd on behalf of Xishuangbanna Tropical Botanical Garden (XTBG).

Summary

Sri Lanka's unique agamid lizards, many restricted to this tropical island nation, face threats from habitat loss and climate change. To protect these endangered reptiles, we implemented a machine-learning approach, species distribution modeling to map their habitat suitability and identify priority conservation areas. Focusing on 14 threatened species and their known species-presence information, we combined numerous bioclimatic environmental predictors and forest cover to predict their spatial distribution in Sri Lanka based on habitat suitability. Results revealed that while all species have some habitat in protected zones like Sinharaja Forest Reserve, the Central Highlands and the Knuckles Range, over 40% of their most critical habitats remain unprotected. These unprotected areas, often in high-elevation forests of the Central Highlands and Knuckles Mountain Range, are vital for species with tiny ranges. The research also highlighted that some human-modified landscapes, like agroforests, could support conservation if managed sustainably. By pinpointing gaps in Sri Lanka's current network of protected areas, this study provides a roadmap for expanding the country's conservation network. The methods used here, particularly the modeling approach, could help protect other poorly studied species worldwide, ensuring limited resources target the most urgent needs.

• Practitioner Points

- Species distribution modeling (SDM) is a powerful tool for identifying conservation gaps/hotspots, including unprotected critical habitats, even for species with limited data, guiding targeted conservation actions.
- Prioritize wet and montane zones: Key conservation lands like Sinharaja Forest, the Knuckles Range, and Central Highlands require expanded and enhanced protection and habitat connectivity to safeguard Ceylonese endemic agamids.
- Integrate human-modified landscapes: Secondary forests and agroforests can complement protected areas if restored and managed to support biodiversity, offering a buffer against habitat loss.

1 | Introduction

Accelerated rates of habitat deterioration and natural-resource exploitation continue to imperil biodiversity worldwide (Hanski 2011; Lovejoy 2006). Systematic conservation planning plays an important role in addressing the biodiversity crisis to protect ecosystem integrity and functions (Margules and Pressey 2000), as well as endemic and threatened species (Myers et al. 2000). Establishing Protected Areas (hereafter PAs) that sufficiently represent variable levels of biological organization and spatial extents has become a critical goal for conservation policies (Lovejoy 2006; Margules and Pressey 2000; Pearson 2016). Establishment and maintenance of PAs remain the most widespread and historic countermeasures against biodiversity erosion (Gray et al. 2016). PAs safeguard against the ill-effects of demographic and genetic stochasticity and support the ecological infrastructure necessary for long-term population viability despite anthropogenic stressors (Pressey et al. 2007; Rodrigues et al. 2004). Despite the increase in global PA cover within the past decade, only 17% of the planet

surface is currently protected (UNEP-WCMC and IUCN 2020). Filling these gaps via systematic conservation planning requires spatially explicit data on species distribution, but scientific knowledge of geographic patterns of biodiversity remains incomplete (Lovejoy 2006). Both information paucity and conservation gaps are particularly notable in the tropical realm, especially with respect to less charismatic, nongame vertebrates, such as reptiles (Rodrigues et al. 2004).

Sri Lanka, a tropical island in Indian ocean, exemplifies such conservation and information gaps, as PAs in the country are primarily designed for charismatic megafauna and economically valuable ecosystems (Miththapala 2016). Sri Lanka is recognized as one of richest and herpetologically most diverse countries of the world with a high number of threatened species (Böhm et al. 2013), and out of 247 reptile species, 67.2% are endemics (Gibson et al. 2020). Therefore, it is worthwhile examining whether its PAs protect reptile diversity, as well as that of other taxa. In particular, we selected agamid lizards as focal species, for several reasons. First, agamid species are ecologically sensitive and evolutionarily distinct, with 19 of 22 species being endemic to Sri Lanka. They are distributed throughout the island, yet accurate data on their populations and distributions remain limited. Second, a high proportion (72.7%) of Sri Lankan agamid lizards are classified as threatened on the IUCN Red List (MOE 2012). Third, agamids are often used as biodiversity indicators (Ishwar et al. 2003; Megia-Palma et al. 2020). Similarly to other taxa, agamid lizards are threatened by land-use changes and other anthropogenic disturbances (Miththapala 2016), such as habitat loss from agricultural conversion, forest degradation and over-exploitation (Böhm et al. 2013). Overall, identifying conservation targets for reptile diversity and endemism in Sri Lanka may help implement proactive policies to halt habitat modification and curb biological homogenization.

Assessment of species distribution ranges is crucial for identifying conservation targets, which typically requires spatially and temporally extensive field surveys (Pearce and Ferrier 2001). Species Distribution Modeling (hereafter SDM) is useful in mapping the potential geographic range of species, especially in regions where comprehensive field surveys are lacking (Hirzel and Le Lay 2008; Peterson 2001). By analyzing known species occurrence data alongside environmental variables, SDMs can predict species distributions across geographic space and time. Multiple approaches to SDMs exist, including generalized regressions, classification trees, discriminant functions, and machine-learning algorithms (Hirzel and Le Lay 2008; Peterson 2001). In recent years, SDM applications have been commonly used by conservation professionals and have been featured prominently in the scientific literature (Feng et al. 2019). In applications of conservation biology, SDM is a valuable tool and has been applied to a wide range of taxa, from herbaceous plants (Abdelal et al. 2019) to numerous vertebrate species such as mammals (Razgour et al. 2011), amphibians and reptiles (Pawar et al. 2007). The maps generated by SDMs delineate geographic areas where species are most likely to occur, that is, their potential distributions. When these spatial predictions are compared with maps of existing PAs, we can identify "conservation gaps," regions of high-priority conservation areas that currently lack formal protection (Ahmadi et al. 2023; Margules and Pressey 2000). Recognizing these gaps is essential for guiding reserve design and establishing conservation priorities.

The explicit development and application of SDMs in conservation planning is increasingly recognized as best practice, particularly when detailed occurrence data are limited (Feng et al. 2019; Johnson and Gillingham 2005).

In this study, with the overarching goal of guiding conservation planning for agamid lizards in Sri Lanka using an SDM-based approach, our specific objectives were to (1) estimate the potential distribution ranges of selected agamid lizard species, (2) identify conservation targets based on SDM-generated species ranges, and (3) highlight conservation gaps outside of the existing PAs. We believe that our study will provide insights for future conservation planning in Sri Lanka for agamid lizards and assist conservation authorities to prioritize high-value targets and to efficiently use limited financial and logistic resources.

2 | Materials and Methods

2.1 | Focal Species

Agamid lizards (Family Agamidae, dragon lizards) in Sri Lanka are both ecologically unique and evolutionarily distinct (Karunaratna et al. 2020; Schulte et al. 2002). Their exceptional degree of endemism, as well as their restricted geographical ranges (Karunaratna et al. 2020), combine to give Sri Lankan agamids a high conservation value, both nationally and globally. Recently, the project “Assess to Plan: Conservation Action Planning for the Snakes and Lizards of Sri Lanka” (Gibson et al. 2020) identified four critically endangered, nine endangered, and two vulnerable agamid species.). However, we excluded *Sitana ponticeriana* because this species is no longer considered a threatened species according to the latest categorization by the IUCN (www.iucn.org) and we didn't include recently described data deficient species *Ceratophora ukuwelai*. Hence, we selected 14 agamid species for this study (Table 1).

2.2 | MaxEnt-Based Distribution Modeling Approach

The prediction of a species' potential distribution based on georeferenced presence-only records is possible through MaxEnt, a spatially explicit, machine-learning algorithm based on maximum entropy theory (Phillips and Schapire 2004). To estimate the probability distribution of a species across a certain geographical area, MaxEnt produces the least biased solution, that is, the one that maximizes its entropy using information on confirmed associations between the species' georeferenced points and abiotic and biotic geospatial variables (e.g., elevation, climate, hydrological features, and land-use or tree cover data (Elith et al. 2006; Pawar et al. 2007). The resulting SDM is a probability distribution prediction over a geographical area of interest, which generates a scaled potential habitat map, wherein each mapped pixel carries a potential value generated as a function of the predictor variables. Higher values correspond to greater predicted potential distribution for the species of interest. Maxent is robust when modeling with presence-only occurrence data, even for small sample sizes, and has outperformed many alternative conventional techniques (Elith et al. 2006; Pearson et al. 2007) in delivering stable, reliable, and high-accuracy predictions across a

range of sample sizes (Wisz et al. 2008). MaxEnt has been adopted across a diverse range of ecological, evolutionary, conservation and biosecurity applications (e.g., to determine environmental drivers of species presence, map current and future distributions, and design protected-area networks).

2.3 | Data Sources: Species Distribution Records and Environmental Predictors

To incorporate the environmental heterogeneity of Sri Lanka, and to accommodate a multi-species perspective, we included the entire country as the study region. We obtained georeferenced species occurrences from both peer-reviewed publications and unpublished field observations by field biologists (Table S1) which were collected using random (Greig-Smith 1964) and adaptive sampling (Krebs 1989) methods. To reduce spatial autocorrelation, we identified clusters of occurrence points within the same 1 km² pixel, the smallest spatial unit in our geospatial environmental space. From each cluster containing multiple points, we randomly selected one occurrence record and removed the rest. This process minimized the influence of spatially autocorrelated data points on model predictions (Tobea et al. 2016). To apply this method, occurrence points for the selected agamid species were spatially rarefied using SDMtoolbox 2.0 (Brown et al. 2017).

2.4 | Species Distribution Modeling

Initially, we used all 19 bioclimatic variables (BIO 01–BIO 19) as climatic predictors of agamid distribution, encompassing annual, monthly, and quarterly climatic trends (<http://www.worldclim.org/>). These variables include central tendencies (e.g., mean annual temperature), seasonal variations (annual range in temperature and precipitation), and extreme measurements (e.g., mean temperature of coldest quarter), and hence are biologically meaningful proxies of species' potential distributions (Fick and Hijmans 2017; Hijmans et al. 2005). These bioclimatic predictors are spatially explicit (spatial resolution: 1 km) and derived from monthly rainfall and temperature records for the period of 30 years (1970–2000; Fick and Hijmans 2017; Hijmans et al. 2005). We excluded BIO 8, 9, 18, and 19, as their use in SDMs is discouraged due to spatial artifacts (Booth 2022).

In addition to the bioclimatic variables, we included tree density (Crowther et al. 2015) as a proxy for habitat structure and forest cover (spatial resolution: 1 km). Further, an elevation layer (Digital Elevation Model) was added, which was produced by the Shuttle Radar Topography mission (Farr et al. 2007). For land use, we used current layers at 1 km² resolution (digitized from 30-m spatial resolution map; <https://earthexplorer.usgs.gov/>), with 31 land-use types, including natural and anthropogenic habitats (Table S1). To avoid over prediction in the modeling process due to collinearity among predictors, we calculated Pearson correlation tests and removed variables that were highly correlated with others ($|r| \geq 0.7$), retaining those variables that have been used in previous SDM studies conducted in Sri Lanka (Wijerathne et al. 2025). The final SDM contained BIO 2, BIO 3, BIO 5, BIO 12, BIO 14, BIO 15, elevation, land use and tree density layer.

TABLE 1 | List of threatened agamid lizards of Sri Lanka used for the study.

Species	Conservation status	Range	Qualitative estimate of abundance
<i>Calotes desilvai</i> Bahir & Maduwage, 2005	CR	Rakwana Hills	Very Rare
<i>Calotes liocephalus</i> Günther, 1872	EN	Peak Wilderness	Rare
<i>Calotes manamendrai</i> Amarasinghe & Karunarathna, 2014	EN	Knuckles Range	Rare
<i>Calotes nigrilabris</i> Peters, 1860	EN	Horton Plains and Nuwara Eliya areas of Central Highlands	Common
<i>Calotes pethiyagodai</i> Amarasinghe, Karunarathna, Hallermann, Fujinuma, Grillitsch & Campbell, 2014	EN	Knuckles Range	Rare
<i>Ceratophora aspera</i> Günther, 1864	EN	Southwestern wet zone	Uncommon
<i>Ceratophora erdeleni</i> Pethiyagoda & Manamendra-Arachchi, 1998	CR	Rakwana Hills	Rare
<i>Ceratophora karu</i> Pethiyagoda & Manamendra-Arachchi, 1998	CR	Rakwana Hills	Rare
<i>Ceratophora stoddartii</i> Gray, 1835	EN	Central Highlands	Uncommon
<i>Ceratophora tennentii</i> Günther & Gray, 1861	CR	Knuckles Range	Uncommon
<i>Cophotis ceylanica</i> Peters, 1861	EN	Central Highlands	Rare
<i>Cophotis dumbara</i> Samarawickrama, Ranawana, Rajapaksha, Ananjeva, Orlov & Ranasinghe, 2006	CR	Knuckles Range	Rare
<i>Lyriocephalus scutatus</i> Linnaeus, 1758	VU	Wet zone and some inselberg mountains	Uncommon
<i>Sitana devakai</i> Amarasinghe, Ineich & Karunarathna, 2015	VU	North west coastal areas	Common

Note: The column “Range” identifies the largest single area specifically occupied by each species. The qualitative estimate of abundance is based on Bahir and Surasinghe (2005), which describes the abundance of this species in sites where it is present. Abbreviations: CR, critically endangered; EN, endangered; NT, near threatened; VU, vulnerable.

Considering the rarity and low coverage of some of the species, we combined several methods for data processing suggested in Jeliaskov et al. (2022). Specifically, we used the SDM + pseudo absence (Barbet-Massin et al. 2012) and bias-corrected SDM approaches (Fourcade et al. 2014). Since our study area is an island, we adopted an unlimited dispersal scenario, which assumes there are no significant dispersal barriers, although we recognize this assumption may not hold true for some species that cannot navigate across valleys between mountain ranges.

Regarding the specifics of the SDM, background points and occurrence data can exhibit geographical bias (Gafna et al. 2023). Multiple methods have been employed in different studies to mitigate this bias (Acevedo et al. 2012; Valavi et al. 2022). We adopted a method similar to that used by Fourcade et al. (2014), generating restricted background points as pseudo absences for all the species to minimize the bias of presence-only data. Initially, we evaluated nine different approaches using a grid graph (Whitford et al. 2024; Supporting Information S1: Figure S1) and assessed performance based on Area Under the Curve (AUC) and Boyce’s index, which compares predicted potential distribution with actual occurrences (Hirzel et al. 2006). This evaluation indicated that using 1000

background points with unlimited buffer distance was the optimal approach for SDMs of these species.

We used R package “ENMeval,” specifically designed for optimizing the MaxEnt approach to SDMs (Kass et al. 2021; Muscarella et al. 2014). Various combinations of feature classes were explored including linear, linear-quadratic, hinge, and linear-quadratic-hinge models (Merow et al. 2013; Radosavljevic and Anderson 2014). Regularization multipliers were applied in increments of 0.5, ranging from 0.5 to 4, to control model complexity. To determine the optimal model for each species, we utilized a spatial block cross-validation approach (Roberts et al. 2017). This method involved dividing the data into four blocks, using three for training and one for testing, as outlined by Hirzel et al. (2006) and Valavi et al. (2022). Initially, the model with the lowest average 10.0% omission rate was selected. If multiple models displayed the same omission rate, the one with the highest average validated AUC value was chosen instead (Zhang et al. 2021). Permutation importance (PI) values for all the predictor variables were obtained to evaluate their influence for the model (Schnase et al. 2021). Subsequently, the consequent drop in area under the receiver operating characteristics curve (AUC) was measured (> 0.8 were considered “good”; Fielding

and Bell 1997) where a greater drop in AUC signifies greater biological importance of the randomized predictor to the overall model performance (Phillips and Schapire 2004). We used Boyce Index together with the AUC to measure the model performance of the SDMs (Table S2). Using QGIS version 3.24.1 (QGIS.org, 2024), we reclassified the distribution maps into a binary raster maps depicting either presence (1) or absence (0) of the focal species using the maximum training sensitivity and specificity cumulative threshold values obtained through the model, as a standard approach to selecting thresholds of occurrence in the prediction of species (Table S2, Liu et al. 2005).

2.5 | Conservation Targets and Gaps

Ranking species based on their conservation priority should account for numerous parameters such as rarity, population trends and density, their current range, and threats. To incorporate these factors, we developed a conservation-priority ranking system, using three main criteria (Table S3): the conservation status in the IUCN Red List 2019 (Gibson et al. 2020; Karunarathna et al. 2020), the number of georeferenced points in the data set, and the total area of the MaxEnt-predicted distribution range. The ranking system ranges between 1 and 5, with 5 representing the highest conservation priority. To determine the final conservation priority for each agamid species, we summed across the three criteria and then reclassified the summed rank into the same 1–5 scale (Table S3).

We then ranked regions of Sri Lanka in their conservation importance. We first excluded fragments < 5 km² from SDMs since such small habitats are unlikely to sustain minimum viable populations, and thus are inadequate for conservation. Then we superimposed the SDM maps for all species in ArcMap. Geographic areas where the distribution ranges of > 3 species overlapped were

considered “conservation targets.” As these conservation targets included protected as well as unprotected landscapes, we superimposed conservation targets with the current PAs of Sri Lanka, to separate areas receiving statutory protection from those without such protection (“conservation gaps”).

Next we ranked PAs in their conservation importance and identified areas for conservation outside of PAs. Based on agamid species present at each PA, we summed species-specific conservation-priority ranks (Table 2) to generate PA-specific conservation priority rankings (Table S4: e.g., a PA with three threatened species each of rank 5, was valued at 15), where PAs with the greatest concentrations of threatened agamid species are valued the highest. Likewise, conservation gaps were defined as areas outside of PAs in which the distribution ranges of at least three agamid species overlapped. Next, by superimposing the SDM results with a land-use layer of Sri Lanka (Land use/land cover image of Sri Lanka using Landsat-7 ETM+C₂L₁ images acquired on 2017 with 30 m spatial resolution, <https://earthexplorer.usgs.gov/>), we identified areas outside of PAs with land-cover types suitable for agamid conservation. Referring to published chronicles of agamid autecology (Karunarathna and Amarasinghe 2011; Karunarathna and Amarasinghe 2012; Pethiyagoda and Manamendra-Arachchi 2012), we determined the conservation potential of land-cover types, as is shown in Table S5.

3 | Results

3.1 | Importance of Predictor Variables

The average AUC values for all SDMs were over 95% (0.95 ± 0.03), and Boyce Index (0.72 ± 0.21) values indicated satisfactory model performance for all selected agamid species. Among all the predictors, BIO 5 (Maximum temperature of the

TABLE 2 | Species-specific conservation priority ranks assigned using three criteria: conservation status in Gibson et al. (2020) and www.iucn.org, the number of georeferenced points in the data set, and the total area of the MaxEnt-predicted distribution range.

Species	Code	Red list status	Rank	Occurrence points	Rank	Area of potential distribution (km ²)	Rank	Total	Overall rank
<i>Calotes desilvai</i>	<i>cades</i>	CR	3	12	5	63.6	5	13	5
<i>Calotes liocephalus</i>	<i>calioc</i>	EN	2	15	5	210.4	5	12	4
<i>Calotes manamendrai</i>	<i>caman</i>	EN	2	17	4	640.3	4	10	3
<i>Calotes nigrilabris</i>	<i>canig</i>	EN	2	35	3	457.2	5	10	3
<i>Calotes pethiyagodai</i>	<i>capet</i>	EN	2	21	4	558.5	5	11	4
<i>Ceratophora aspera</i>	<i>ceasp</i>	EN	2	40	2	3372.0	3	7	2
<i>Ceratophora erdeleni</i>	<i>ceerd</i>	CR	3	12	5	38.2	5	13	5
<i>Ceratophora karu</i>	<i>cekar</i>	CR	3	12	5	37.2	5	13	5
<i>Ceratophora stoddartii</i>	<i>cesto</i>	EN	2	41	2	2454.7	4	8	2
<i>Ceratophora tennentii</i>	<i>ceten</i>	CR	2	17	4	307.2	5	11	4
<i>Cophotis ceylanica</i>	<i>cocey</i>	EN	2	17	4	415.4	5	11	4
<i>Cophotis dumbara</i>	<i>codum</i>	CR	2	16	5	302.2	5	12	4
<i>Lyriocephalus scutatus</i>	<i>lyscu</i>	VU	1	84	1	9965.7	2	4	1
<i>Sitana devakai</i>	<i>sidev</i>	VU	1	23	4	11710.9	1	6	1

Note: Both criterion-specific and final ranks (summed ranks from all criteria) for conservation prioritization ranged between 1 and 5, indicating lowest and highest priorities for conservation.

warmest month) and Elevation had the most influence on the resulting SDMs (Supporting Information S1: Figure S2).

3.2 | Predicted Distribution Ranges of Selected Species and Their Conservation-Priority Ranks

According to SDM maps we generated (Figure 1, Table 2), *Sitana devakai* showed the broadest distribution range (117,10.90 km²), followed by *Lyriocephalus scutatus* (9965.76 km²), while *Ceratophora karu* (37.21 km²) and *Ceratophora erdeleni* (38.2 km²) had the most restricted ranges, exclusively limited to the Rakwana

Hills. In addition, *Calotes desilvai* (63.60 km²) was among the species with the smallest ranges, also restricted to the Rakwana Hills. In contrast, the predicted distribution of *Lyriocephalus scutatus* covered lowlands as well as the three montane areas of southwestern Sri Lanka. For all the species collectively, the wet zone encompassed 45.0% of the predicted distribution ranges, the intermediate zone covered 17.1%, and the remaining portion of the distribution range was in the dry zone (Supporting Information S1: Figure S3). However, these figures overemphasize the importance of the dry-zone, as only *Sitana devakai* was widely distributed there. All other species had their primary distribution range within the wet and montane zones (see Figure 1).

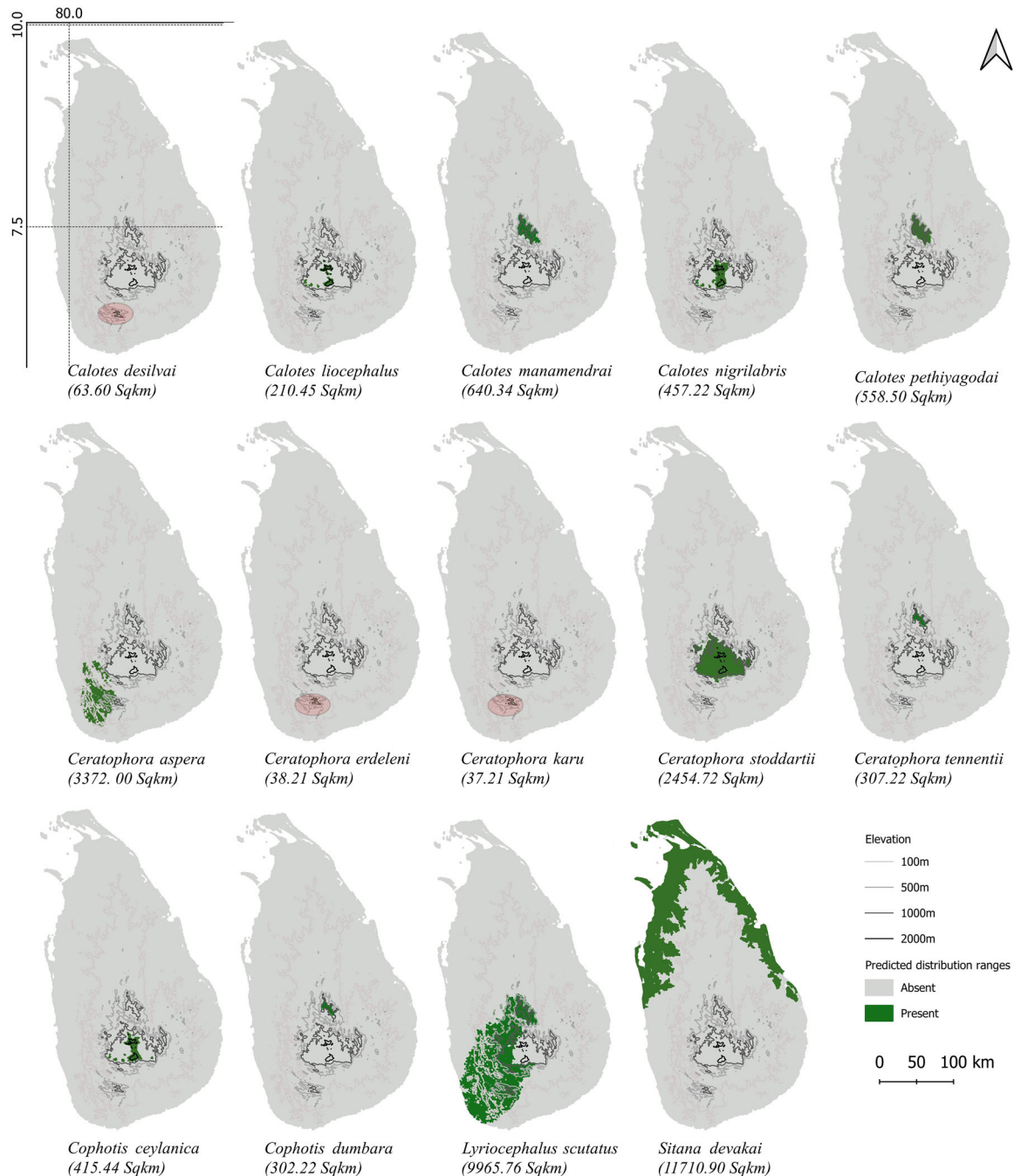


FIGURE 1 | Predicted geographic distribution ranges of 14 threatened agamid species in Sri Lanka. Small pink ellipses highlight areas of distribution that are barely visible.

Three species were given the highest priority conservation rank (5; Table 2), with all these species showing the most restricted predicted distribution ranges ($< 65 \text{ km}^2$) and being classified as CR, according to the IUCN reptile assessment of 2019 (Gibson et al. 2020). Likewise, five species were ranked as high conservation priorities (rank 4). In contrast, several species were ranked as low conservation priorities (two species at rank 1, and two further species at rank 2).

3.3 | Conservation Targets and Gaps

3.3.1 | Conservation Targets Based on Number of Species Present

We identified three geographically distinct clusters as conservation targets in Sri Lanka (Figure 2). These include the Central Highlands, the Rakwana Hills, and the Knuckles Range. Nested in the southwestern wet zone, and characterized by moist, evergreen montane forest ecosystems, the Rakwana Hills showed both high overall agamid species richness and endemism. The predicted distribution of *Ceratophora karu*, *Ceratophora erdeleni*, and *Calotes desilvai* were restricted to the Rakwana Hills, at elevations $> 900 \text{ m}$ above mean sea level. In addition, the predicted distribution of *Ceratophora aspera* included the Rakwana Hills, while also including parts of the lowland ($< 900 \text{ m}$) wet zone. The Knuckles Range, a humid montane ecosystem dominated by evergreen rainforests with high vegetation heterogeneity, included the predicted distribution ranges for five threatened agamid species (*Calotes manamendrai*, *Calotes pethiyagodai*, *Ceratophora tennenti*, *Cophotis dumbera* and *Lyriocephalus scutatus*). Finally, the distribution of five agamid species included the Central Highlands ($> 900 \text{ m}$), with four species (*Ceratophora stoddartii*, *Calotes nigrilabris*, *Cophotis ceylanica* and *Calotes lyriocephalus*) restricted to that mountain range (Table 3).

3.3.2 | Conservation Targets Inside PAs, and Their Prioritization

Conservation targets, defined as regions with high species richness of agamids (> 3 species), were restricted to the higher elevations ($> 900 \text{ m}$) of the Central Highlands and other mountainous regions of southwestern Sri Lanka (Figure 2). A substantial proportion (53%) of these targets is already within a designated PA (Figure 3). PAs covered 461.33 km^2 of these target distribution range of threatened agamids, without *Sitana devakai* (this species was not included in the conservation target analysis because of its discontinuous distribution). Many protected areas $> 900 \text{ m}$ elevation also were highly ranked (with a score of more than 10) as conservation priorities (Table S4). Among all PAs we analyzed, Sinharaja Forest Reserve in the Rakwana Hills ($> 900 \text{ m}$ elevation range of eastern Sinharaja) gained the highest total priority rank (18), as it encompassed the distribution of five threatened agamid species (Table S4). Numerous PAs in the Central Highlands, particularly those located at the higher elevations ($> 1500 \text{ m}$, e.g., Peak Wilderness, Samanala Adawiya, Piduruthalagala, Kandapola, Hakgala, and Horton Plains) also received high ranks (Table S3). With a richness of five threatened agamid species, the Knuckles Range also ranked highly among the PAs, receiving a total score of 16.

3.3.3 | Conservation Gaps Outside PAs

Located within the Central Highlands and Knuckles Range, conservation targets that amount to an area of 403.32 km^2 lie outside the PAs (Figure 3 and Supporting Information S1: Figure S4). In contrast, all conservation targets found in the Rakwana Hills were found within PAs, and no more than two of the selected agamid species were found in unprotected conservation targets there. Aside from the areas west of the Knuckles Range and a few habitat patches along the Central Highlands, the remaining unprotected conservation targets are located above 1200 m elevation (Supporting Information S1: Figure S4). Much of the Central Highlands, as well as the eastern slopes of the Knuckles Range, offer sufficiently large, contiguous, high-priority habitats for agamid conservation, though these areas are currently unprotected. These unprotected conservation targets include a small portion (16.07 km^2) of highly fragmented habitats ($< 5 \text{ km}^2$) attached to PAs. However, much (387.25 km^2) of these unprotected high-priority habitat patches are sufficiently large in spatial extent ($> 5 \text{ km}^2$) and are found along the PA boundaries (Supporting Information S1: Figure S5).

3.3.4 | Conservation Targets After Land-Cover Analysis

After conducting the land-use analysis (Figure 4), nearly 172.61 km^2 of conservation target areas outside of PAs were identified as suitable for threatened agamid conservation, being composed of isolated patches of forests, scrublands and agroforests with high agamid diversity (Table S5). This corresponds to nearly 42.8% of the total acreage of conservation targets outside the PAs. In contrast, 230.71 km^2 of the conservation targets outside of PAs corresponded to habitat areas moderately or highly unsuitable for the focal species.

4 | Discussion

4.1 | Distribution-Range Predictions and Species-Level Conservation Prioritization

Our study marks a cornerstone in Sri Lanka's systematic conservation planning where we produced SDMs for multiple species of conservation interest to estimate their current distribution, thereby identifying conservation target areas, including conservation gaps. We identified that nearly ~45% of agamid conservation targets remain unprotected, requiring immediate conservation action. These findings align with global-scale conservation gap analyses for reptiles, which indicate that a high proportion of conservation priority areas remains unprotected (Cox et al. 2022; Rodrigues et al. 2004a; Rodrigues 2004b).

Overall, our predicted distribution ranges exceeded historical estimates from previous publications for both widely distributed and range restricted species. For example, historical range sizes for *Lyriocephalus scutatus* and *Ceratophora aspera* were estimated at 700 km^2 and 800 km^2 , respectively (Bahir and Surasinghe 2005), while our analyses suggest ranges of 3300 km^2 and 9000 km^2 . Similarly, historical estimates for range-restricted species such as *Calotes desilvai*, *Ceratophora karu*, and *Ceratophora erdeleni* were $< 10 \text{ km}^2$, whereas our models predict ranges of $30\text{--}60 \text{ km}^2$. Recent

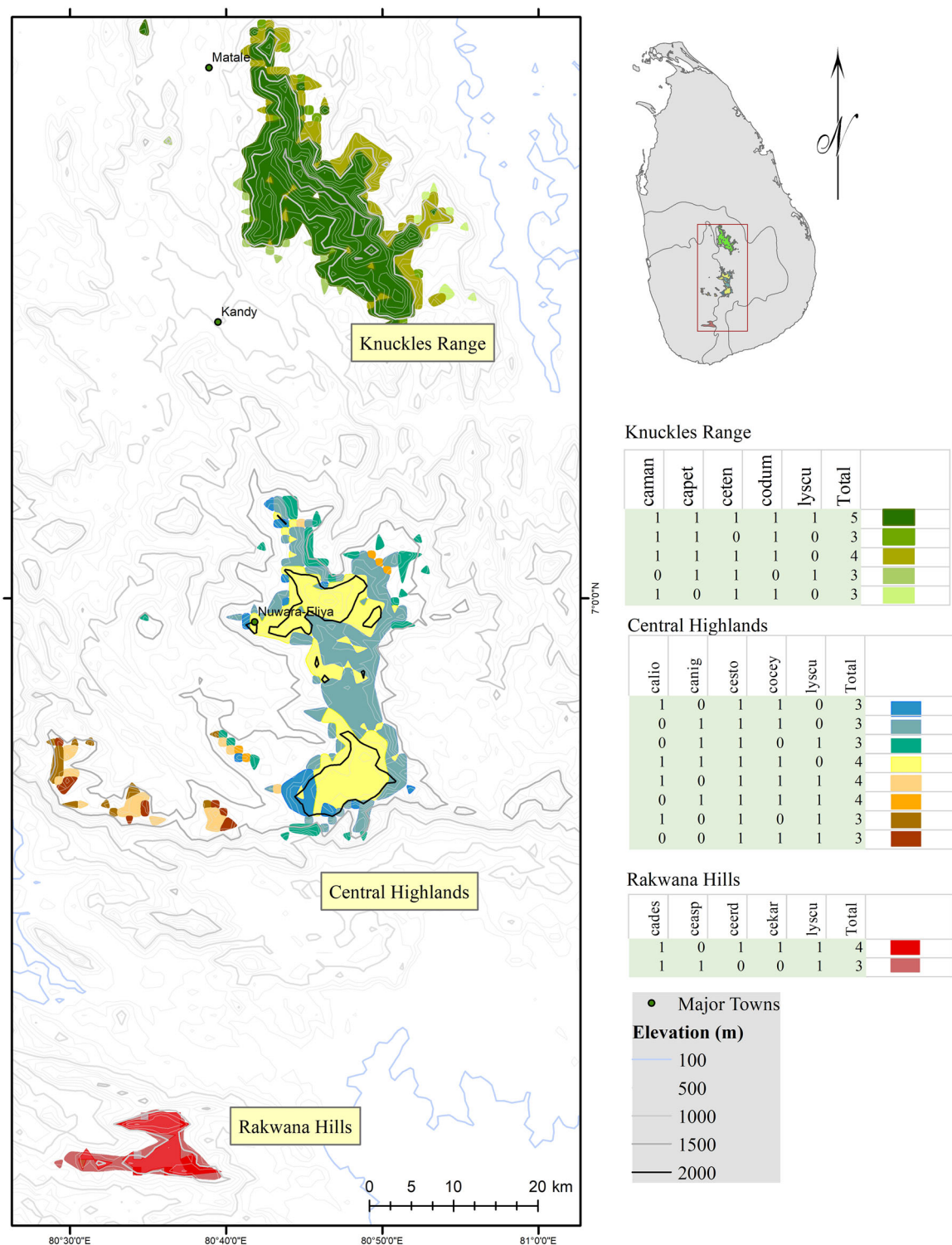


FIGURE 2 | Conservation target areas for agamids: including (from south to north) Rakwana Hills, Central Highlands and the Knuckles Range. Each colors corresponds to an area with a specific combination of species. Binary number tables of presence “1” and absence “0” of each species are shown, and also the total number of species present. Species codes are shown as in Table 2.

studies (Gibson et al. 2020) suggest that, contrary to historical records and natural history accounts (Deraniyagala 1955), geographical relicts such as *Lyriocephalus scutatus* and *Ceratophora aspera* may have had healthy, stable, widely distributed populations across the mid-elevations and lowlands of the wet zone. Our predicted distributions are also corroborated by recent surveys, in

which these threatened agamid species were found in isolated secondary forest fragments, forest plantations, and home gardens (Karunaratna and Amarasinghe 2011). Structural complexity of vegetation, cool-moist microclimatic features, and food availability have been identified as the critical determinants for the occupancy of range-restricted agamids (Jayasekara et al. 2018; Somaweera

TABLE 3 | Range of agamid species overlaid within the most important high-priority conservation areas.

Region	Total no. of species present	Species confined to mountain range	Maximum species overlapping in one place	Total high-priority conservation area (km ²)
Knuckles Range	5	<i>Calotes manamendrai</i> <i>Calotes pethiyagodai</i> <i>Ceratophora tennentii</i> <i>Cophotis dumbara</i> <i>Lyriocephalus scutatus</i>	5	387.22
Central Highlands	5	<i>Calotes liocephalus</i> <i>Calotes nigrilabris</i> <i>Ceratophora stoddartii</i> <i>Cophotis ceylanica</i> <i>Lyriocephalus scutatus</i>	5	418.99
Rakwana Hills	5	<i>Calotes desilvai</i> <i>Ceratophora erdeleni</i> <i>Ceratophora aspera</i> <i>Ceratophora karu</i> <i>Lyriocephalus scutatus</i>	4	63.34

Note: The total number of species present within the three regions are shown, although all the species are not evenly distributed throughout the regions.

et al. 2015). Novel ecosystems such as secondary and degraded forests and abandoned monocrop plantations that characterize Sri Lanka's wet zone are utilized by threatened and endemic fauna for at least part of their life cycle (Pethiyagoda 2012). Our study adds credence to the importance of these novel ecosystems as shelters for native biodiversity, although not as replacements for primary forests.

The species that we found to be conservation priorities have also been highlighted as conservation targets in previous, location-specific studies (Jayasekara et al. 2018; Somaweera et al. 2015), as well as in island-wide conservation assessments (Bahir and Surasinghe 2005). Moreover, the agamid species we ranked for high-priority conservation also have phylogenetic and evolutionary uniqueness. For instance, the endemic, horned-lizard genus *Ceratophora* reveals multiple, independent evolutionary events involving their rostral appendage (Johnston et al. 2013; Schulte et al. 2002). Likewise, the endemic *Calotes* have undergone insular radiation (Macey et al. 2000).

4.2 | Prioritization of Conservation Target Areas

We identified forests in both wet and intermediate zones as critical targets for protecting threatened agamid species. These tropical rain forests are critical for conservation of endemic reptiles (Erdelen 1988), amphibians (Surasinghe 2009), freshwater fish (Pethiyagoda 1994), woody plants (Gunatilleke and Ashton 1987) and the avifauna of Sri Lanka (Abeyarama and Seneviratne 2017). Among these, the montane forests of the Central Highlands, the Knuckles Range, and the Rakwana Hills are particularly notable for their biodiversity (Gunatilleke, Pethiyagoda, et al. 2017). The elevation gradient characteristic of these regions begets a climatic gradient, which then shapes

differential vegetation layering of the forest. The resulting environmental heterogeneity is largely responsible for the high overall faunal diversity characteristic to these mountainous regions. Ecological distinctiveness of these regions has been used to recognize “key biodiversity areas” at the global scale, based on taxonomic uniqueness, exceptional biodiversity and endemism, evolutionary and phylogenetic phenomena, and rarity of habitat types (Olson and Dinerstein 1998; Eken et al. 2004). These recognitions highlight the vulnerability and irreplaceability of the conservation targets identified in our study, both of which are fundamental criteria in systematic conservation planning (Margules and Pressey 2000). Thus, the conservation targets we identified for agamids may provide multiple benefits, encompassing a range of taxa, safeguarding globally and regionally rare ecoregions, and preserving unique biogeographic histories and evolutionary processes.

Beyond their high endemism, the conservation targets we identified face numerous anthropogenic threats. Except for small tracts in inaccessible terrains (e.g., high elevations, steep slopes), both lowland and montane rainforests have historically been cleared for commercial-scale export agriculture and selectively logged for plywood and hardwood timber for decades (Perera 2001). Although logging is now banned in PAs, these ecosystems continue to bear the legacy of past exploitation (Perera 2001). Furthermore, amidst the growing human population, forest cover of the wet and intermediate zone has dwindled to less than a tenth of its original cover and the remaining forests suffer from severe fragmentation (Aukema et al. 2017). Even today, illicit resource extractions (timber felling, non-timber forest products extraction, mining, poaching, and collecting for international pet trade) continue despite strict legislative frameworks (Gunatilleke, Pethiyagoda, et al. 2017). Enigmatic forest diebacks have also been reported

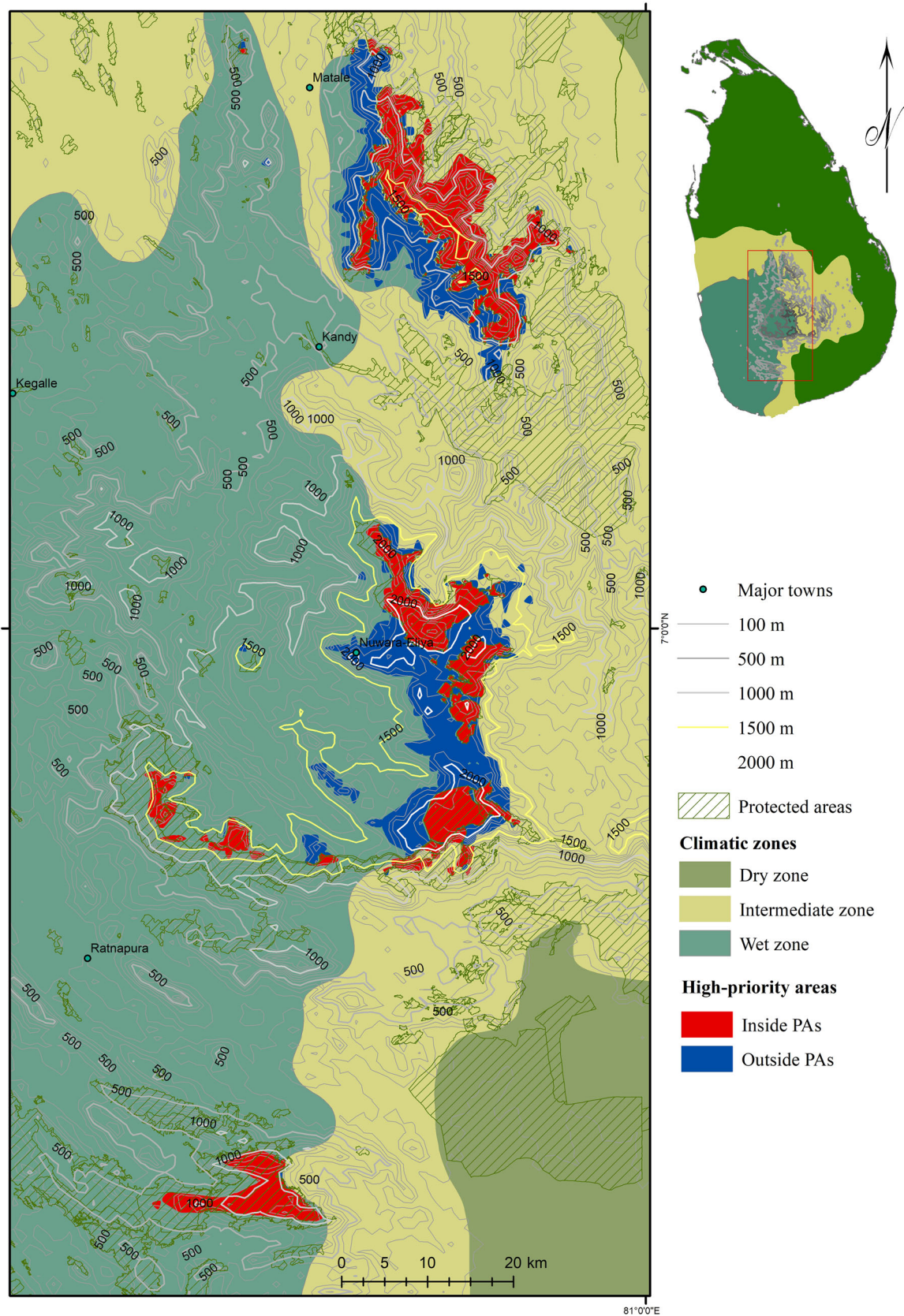


FIGURE 3 | Predicted high-priority target areas for conservation of threatened agamids, compared with the protected area (PA) network in south-western and central Sri Lanka. Red patches of high-priority conservation areas within PAs are mostly concentrated in Knuckles Range, Rakwana Hills and Central Highlands, with few others scattered outside the above three regions and are demarcated as PAs. Blue colored areas are high-priority areas currently outside the PAs.

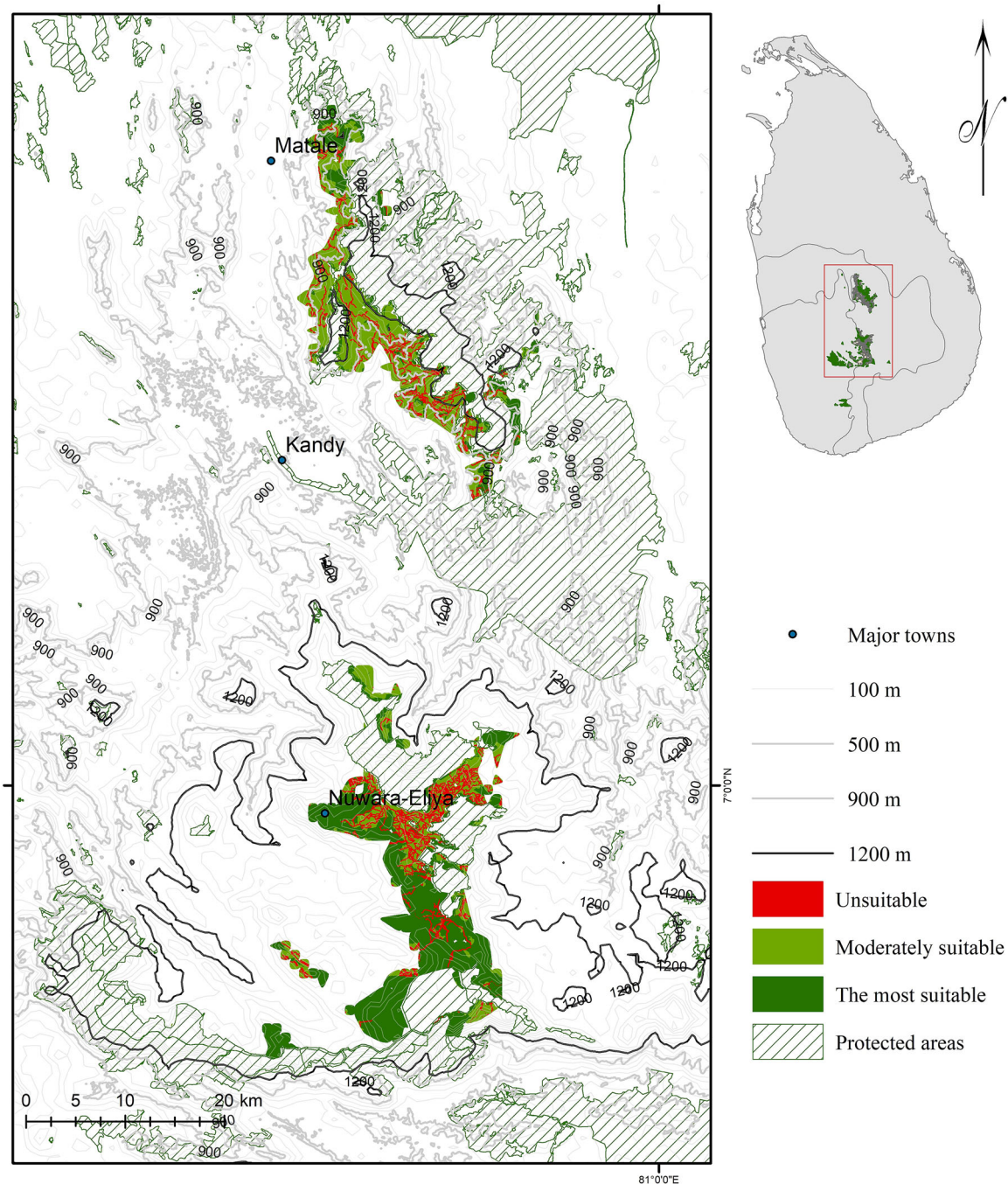


FIGURE 4 | Colored areas indicate high-priority areas outside of PAs where agamids are not protected, and are further categorized as to how suitable the habitat is for agamid conservation.

from montane forests, which may alter unique forest structural complexity and microclimatic features rendering these habitats less suitable for environmentally sensitive agamids (Ranasinghe et al. 2007).

Our predicted critical agamid habitats are not immune to the adversities of climate change. Altitudinal shifts of species (i.e., moving to higher elevations) seeking wet and cool climatic conditions have been documented worldwide (Neate-Clegg and Tingley 2023; Parmesan and Yohe 2003; Rubenstein et al. 2023; Walther et al. 2002). Several critically endangered agamids (e.g., *Ceratophora erdeleni*, *Calotes desilvai*, *Ceratophora karu*)

are already confined to higher elevations as substantiated through our SDMs. Species with restricted high-altitude ranges are at the greatest risk of extinction (Aukema et al. 2017; Parmesan and Yohe 2003; Tobeña et al. 2016; Wijerathne et al. 2025). Increased drought frequency, altered moisture regimes, reduced water availability, and elevated temperatures pose significant threats to the ecological integrity of optimal agamid habitats (Kottawa-Arachchi and Wijeratne 2017). Climatic anomalies are already evident in Sri Lanka (Burt and Weerasinghe 2014). If these anomalies continue to intensify, they will significantly impact Sri Lanka's agamid species through various ecological changes (Dayananda et al. 2021).

These imminent threats underscore the high-priority we have assigned to both the wet and intermediate zones.

4.3 | Effectiveness of SDMs in Conservation Planning

The potential distribution ranges we have predicted reflect physiological and behavioral adaptation of a species to abiotic and environmental conditions. The underestimation of geographic ranges in previous studies may result from imperfect detection or spatial biases during field surveys, leading to false negatives (Nichols et al. 2008; Otto and Roloff 2011). In addition, it is both logistically infeasible and prohibitively expensive to cover all potential habitats via conventional surveys, which can still underestimate the true distribution range (MacKenzie and Nichols 2004; Wilson et al. 2005). Since MaxEnt delivers reliable predictions despite small field samples (Costa et al. 2010), the SDM approach is of great value when targeting such species for protection (Abdelal et al. 2019; Guisan and Thuiller 2005; Remya et al. 2015).

The SDM-based predictions we made indicate agamid predicted current geographic distribution, not their long-term persistence. In practice, large portions of species' distribution ranges are suboptimal for species persistence. Due to inadequate management actions, even PAs can suffer from diminished ecological integrity (Rodrigues et al. 2004). Therefore, the increased distribution range sizes of agamids we reported does not warrant downgrading their conservation status. The long-term suitability of heavily modified and disturbed, suboptimal habitats for threatened and endangered agamids remains questionable (Erdelen 2012). For instance, compared to primary and natural forests, agroecosystems of Sri Lanka's wet zone are dominated by common agamid species, whereas the occurrence of highly specialized, threatened and endemic species are infrequent (Gamage et al. 2011). These suboptimal habitats are also prone to biological invasions, frequent human disturbances (Perera 2001), and extractions (Chokkalingam et al. 2001) that may alter critical habitat structure, rendering them less suitable for conservation-dependent agamids. Subsidized predation by human commensals or competition against disturbance-tolerant generalists can also challenge long-term persistence of sensitive agamid species (Oro et al. 2013). Nonetheless, with sustainable management actions, these suboptimal habitats can offer opportunities to promote conservation.

4.4 | Limitations of the Study

Our study has several limitations commonly encountered in SDM studies. For example, SDMs may lack microhabitat information (McCoy and Bell 1991), and neglect the effect of various biotic interactions, such as competition and predation (Dunne et al. 2002), which can lead to inaccurate predictions. Another prevalent issue is the reliance on coarse resolution data, which may overlook critical habitat features (Guisan and Thuiller 2005). Also, a lack of updated data layers, such as land use and vegetation, hinders the modeling efforts, and a deficiency of occurrence data poses a significant challenge in Sri Lanka, particularly for rare species restricted to specific

geographic patches. Thus, future research should focus on exploring specific microclimatic layers for SDMs and also recommended to follow new sampling and modeling technique to enhance the clarity of research.

Another potential issue is over-prediction, which is a well-documented issue that can lead to inflated estimates of species distributions (Mendes et al. 2020). One major assumption of our SDM is unlimited dispersal within the island; however, climatic, habitat, and anthropogenic factors can constrain dispersal. Certain species, endemic to specific montane regions such as the Knuckles Range and Rakwana Hills, have historically struggled to move across unfavorable low-elevation habitats that separate these montane areas; traversing such areas may be even more difficult now due to habitat loss and fragmentation. Small populations may die out due to demographic stochasticity or anthropogenic disturbance, and then this lack of dispersal may mean that they are not colonized again. Hence, it is very important to follow-up our modeling with more ground-truthing, and our maps (e.g., Figure 4, showing high diversity agamid target areas outside of protected areas) can guide such surveys.

4.5 | Recommended Conservation Actions for Conservation Targets

Habitat loss due to human settlements, agriculture, and natural resource overexploitation are the most impactful threats to reptiles (Böhm et al. 2013), especially in continental islands like Sri Lanka. Existing conservation lands we ranked with high-priority (Sinharaja and the Rakwana Hills, the Knuckles Range) can be targeted in management plans. But there is an inadequacy of conservation actions outside conventional PAs in Sri Lanka (Bambaradeniya et al. 2004; Erdelen 2012). Addressing such challenges is a critical necessity for agamids and other taxa of conservation interest (e.g., mammals, Fernando and Ranasinghe 2009; Nekaris et al. 2015, and avifauna, Goodale et al. 2014).

The sizable proportion (44.6%) of high-priority conservation gaps we identified calls for strategizing conservation outside PAs. With effective restoration (e.g., introduction of native flora, invasive species removal, species re-introductions), abandoned croplands, forest plantations (Ashton et al. 2001), and mixed-crop eco-agricultural landscapes will provide opportunities for agamid conservation (Scherr and McNeely 2008). The conservation gaps we identified can also be managed as reserve buffers, wildlife corridors, or components of the landscape matrix, thereby enhancing landscape-scale conservation and habitat connectivity (Pressey et al. 2007). While conventional land protection is crucial for agamid conservation, Sri Lankan authorities must extend management efforts beyond PA boundaries to address anthropogenic stressors, such as habitat degradation, fragmentation, and human-wildlife conflict. A landscape-scale approach, including buffer zones, wildlife corridors, and habitat restoration, will improve connectivity between protected areas, mitigate ecological isolation, and enhance long-term species survival (Lovejoy 2006; Pressey et al. 2007). As PAs in the wet and intermediate zones are severely fragmented, landscape-scale connectivity is essential to rekindle metapopulation dynamics and gene flow in isolated

habitat fragments (Chokkalingam et al. 2001; Erdelen 2012; Miththapala 2016). Lowland rainforests that mostly remain as scattered small fragments (> 100 ha) with varying degrees of protection benefit from landscape-scale connectivity and reserve buffers to ameliorate edge effects.

The rare and endemic flora and fauna provides biodiversity conservation potential (Gunatilleke et al. 2005; Gunatilleke, Gunatilleke, et al. 2017), ecosystem values (Surasinghe et al. 2020), and carbon sequestration (Senadheera et al. 2019), and is therefore compatible with holistic, multi-purpose conservation. Reptiles broadly mirror distribution patterns of other taxa and are sensitive to habitat degradation, and thus paint a representative view of biodiversity, albeit being greatly overlooked in reserve selection and design (Böhm et al. 2013; Roll et al. 2017). Therefore, we propose threatened, endemic reptiles as a biodiversity surrogate for systematic conservation planning in Sri Lanka. Our taxon-specific approach is comparable to similar efforts applied for other taxa, such as Important Bird Areas (Fishpool and Evans 2003), Endemic Bird Areas (Brooks et al. 2017), and Important Plant Areas (Darbyshire et al. 2017) which led to successful on-ground conservation actions at variable spatial scales. Furthermore, the fine-grained, spatially explicit approach we used to identify conservation targets can assist Sri Lanka's natural resource managers and conservation officials in expanding existing PAs, designating new conservation areas, enhancing legislative protection for current PAs, and reforming land management policies with a focus on conservation.

5 | Concluding Remarks

Tropical wet evergreen lowland rainforests and moist montane and submontane evergreen forests are critical for conservation of the range-restricted, threatened agamid lizards of Sri Lanka. These forest ecoregions are irreplaceable, ecologically distinct, and vulnerable to anthropogenic pressures, and thus must be prioritized in systematic conservation planning. Yet high-priority agamid habitats in Sri Lanka span across protected and unprotected landscapes, and therefore conservation actions must also include novel ecosystems such as secondary and regenerating forests. Thus, we recommend a landscape-scale, holistic approach that accounts for habitat connectivity, science-based management, and forest restoration for agamid conservation in Sri Lanka. As pressure mounts on natural resources due to anthropocentric demands, effective management of human-modified landscapes is imperative for agamid conservation, which presents a daunting task for Sri Lankan conservation authorities. A broad range of eco-evolutionary and socioeconomic factors that are temporally and spatially dynamic will define and contextualize the ecological integrity and future status of conservation targets. As geographic patterns in biodiversity, species vulnerabilities, and anthropogenic legacies vary in time, the prospects for biodiversity protection should identify and accommodate spatial and temporal heterogeneity in conservation needs and opportunities. Biodiversity threats such as climate change and environmental pollution will add complexity to future conservation planning. Predictive tools such as SDMs that account for both natural and anthropogenic dynamics will be increasingly useful to make scientifically informed conservation decisions.

Author Contributions

Iresha Wijerathne: conceptualization, methodology, software, formal analysis, writing – original draft. **Suranjan Karunarathna:** investigation, data curation, validation. **Thilina Sursinghe:** writing – original draft, writing – review and editing, validation. **Dulan R. Vidanapathirana:** investigation, validation, data curation, visualization. **Kanishka Ukuwela:** investigation, validation, data curation. **Sriyani Wickramasinghe:** writing – review and editing, conceptualization, resources. **Chaya Sarathchandra:** visualization, software. **Jagath Gunatilake:** resources, visualization, project administration. **Aiwu Jiang:** writing – review and editing, visualization, resources. **Eben Goodale:** writing – review and editing, visualization, validation. **Suranjan Fernando:** conceptualization, methodology, supervision.

Acknowledgments

Our special thanks to Bud Chapmen and Channaka Jayathilake, Janaki Manori, Malika Gunawardhana, Pavithra Panduwawala and Shyamala Nirmani for their technical assistance to the project. ILW is sincerely grateful to her family for encouragement, and also thanks the Idea Wild team for providing field equipment. Finally, our sincere gratitude to the reviewers and editorial board of the journal for valuable comments and suggestions for improving the manuscript. Iresha Wijerathne appreciates the support of a Chinese Government Scholarship from the Chinese Scholarship Council (CSC-2021SLJ009558).

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

References

- Abdelaal, M., M. Fois, G. Fenu, and G. Bacchetta. 2019. "Using MaxEnt Modeling to Predict the Potential Distribution of the Endemic Plant *Rosa arabica* Crép in Egypt." *Ecological Informatics* 50: 68–75. <https://doi.org/10.1016/j.ecoinf.2019.01.003>.
- Abeyarama, D. K., and S. S. Seneviratne. 2017. "Evolutionary Distinctness of Important Bird Areas (IBAs) of Sri Lanka: Do the Species-Rich Wet Zone Forests Safeguard Sri Lanka's Genetic Heritage?" *Ceylon Journal of Science* 46, no. 5: 89. <https://doi.org/10.4038/cjs.v46i5.7456>.
- Acevedo, P., A. Jiménez-Valverde, J. M. Lobo, and R. Real. 2012. "Delimiting the Geographical Background in Species Distribution Modelling." *Journal of Biogeography* 39, no. 8: 1383–1390. <https://doi.org/10.1111/j.1365-2699.2012.02713.x>.
- Ahmadi, M., M. R. Hemami, M. Kaboli, and F. Shabani. 2023. "MaxEnt Brings Comparable Results When the Input Data Are Being Completed; Model Parameterization of Four Species Distribution Models." *Ecology and Evolution* 13, no. 2: 1–13. <https://doi.org/10.1002/ece3.9827>.
- Ashton, M. S., C. V. S. Gunatilleke, B. M. P. Singhakumara, and I. A. U. N. Gunatilleke. 2001. "Restoration Pathways for Rain Forest in Southwest Sri Lanka: A Review of Concepts and Models." *Forest Ecology and Management* 154, no. 3: 409–430. [https://doi.org/10.1016/S0378-1127\(01\)00512-6](https://doi.org/10.1016/S0378-1127(01)00512-6).
- Aukema, J. E., N. G. Pricope, G. J. Husak, and D. Lopez-Carr. 2017. "Biodiversity Areas Under Threat: Overlap of Climate Change and Population Pressures on the World's Biodiversity Priorities." *PLoS One* 12, no. 1: e0170615. <https://doi.org/10.1371/journal.pone.0170615>.

- Bahir, M. M., and T. D. Surasinghe. 2005. "A Conservation Assessment of the Sri Lankan Agamidae (Reptilia: Sauria)." *Raffles Bulletin of Zoology* 12: 407–412.
- Bambaradeniya, C. N. B., J. P. Edirisinghe, D. N. De Silva, C. V. S. Gunatilleke, K. B. Ranawana, and S. Wijekoon. 2004. "Biodiversity Associated With an Irrigated Rice Agro-Ecosystem in Sri Lanka." *Biodiversity and Conservation* 13, no. 9: 1715–1753. <https://doi.org/10.1023/B:BIOC.0000029331.92656.de>.
- Barbet-Massin, M., F. Jiguet, C. H. Albert, and W. Thuiller. 2012. "Selecting Pseudo Absences for Species Distribution Models: How, Where and How Many?" *Methods in Ecology and Evolution* 3, no. 2: 327–338. <https://doi.org/10.1111/j.2041-210X.2011.00172.x>.
- Böhm, M., B. Collen, J. E. M. Baillie, et al. 2013. "The Conservation Status of the World's Reptiles." *Biological Conservation* 157: 372–385. <https://doi.org/10.1016/j.biocon.2012.07.015>.
- Booth, T. H. 2022. "Checking Bioclimatic Variables That Combine Temperature and Precipitation Data Before Their Use in Species Distribution Models." *Austral Ecology* 47, no. 7: 1506–1514. <https://doi.org/10.1111/aec.13234>.
- Brooks, M. E., K. Kristensen, K. J. van Benthem, et al. 2017. "Glmmtmb Balances Speed and Flexibility Among Packages for Zero-Inflated Generalized Linear Mixed Modeling." *R Journal* 9, no. 2: 378–400. <https://doi.org/10.32614/rj-2017-066>.
- Brown, J. L., J. R. Bennett, and C. M. French. 2017. "SDMtoolbox 2.0: The Next Generation Python-Based GIS Toolkit for Landscape Genetic, Biogeographic and Species Distribution Model Analyses." *PeerJ* 5, no. 12: e4095. <https://doi.org/10.7717/peerj.4095>.
- Burt, T., and K. Weerasinghe. 2014. "Rainfall Distributions in Sri Lanka in Time and Space: An Analysis Based on Daily Rainfall Data." *Climate* 2, no. 4: 242–263. <https://doi.org/10.3390/cli2040242>.
- Chokkalingam, U., D. M. Bhat, and E. Kalimantan. 2001. "Secondary Forests Associated With the Rehabilitation of Degraded Lands in Tropical Asia: A Synthesis Promotion for Sustainable Forest Management in." *Journal of Tropical Forest Science* 13, no. 4: 816–831. <https://cgspage.cgiar.org/items/a21418e1-e000-4c65-b3fa-48b73428c651>.
- Costa, G. C., C. Nogueira, R. B. Machado, and G. R. Colli. 2010. "Sampling Bias and the Use of Ecological Niche Modeling in Conservation Planning: A Field Evaluation in a Biodiversity Hotspot." *Biodiversity and Conservation* 19, no. 3: 883–899. <https://doi.org/10.1007/s10531-009-9746-8>.
- Cox, N., B. E. Young, P. Bowles, et al. 2022. "A Global Reptile Assessment Highlights Shared Conservation Needs of Tetrapods." *Nature* 605, no. 7909: 285–290. <https://doi.org/10.1038/s41586-022-04664-7>.
- Crowther, T. W., H. B. Glick, K. R. Covey, et al. 2015. "Mapping Tree Density at a Global Scale." *Nature* 525: 201–205. <https://doi.org/10.1038/nature14967>.
- Darbyshire, I., S. Anderson, A. Asatryan, et al. 2017. "Important Plant Areas: Revised Selection Criteria for a Global Approach to Plant Conservation." *Biodiversity and Conservation* 26, no. 8: 1767–1800. <https://doi.org/10.1007/s10531-017-1336-6>.
- Dayananda, B., S. B. Bezeng, S. Karunaratna, and R. A. Jeffree. 2021. "Climate Change Impacts on Tropical Reptiles: Likely Effects and Future Research Needs Based on Sri Lankan Perspectives." *Frontiers in Ecology and Evolution* 9: 688723. <https://doi.org/10.3389/fevo.2021.688723>.
- Deraniyagala, P. E. P. 1955. *A Colored Atlas of Some Vertebrates From Ceylon*. Ceylon Government Press.
- Dunne, J. A., R. J. Williams, and N. D. Martinez. 2002. "Network Structure and Biodiversity Loss in Food Webs: Robustness Increases With Connectance." *Ecology Letters* 5, no. 4: 558–567. <https://doi.org/10.1046/j.1461-0248.2002.00354.x>.
- Eken, G., L. Bennun, T. M. Brooks, et al. 2004. "Key Biodiversity Areas as Site Conservation Targets." *Bioscience* 54, no. 12: 1110–1118. [https://doi.org/10.1641/0006-3568\(2004\)054\[1110:KBAASC\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2004)054[1110:KBAASC]2.0.CO;2).
- Elith, J., C. H. Graham, R. P. Anderson, et al. 2006. "Novel Methods Improve Prediction of Species' Distributions From Occurrence Data." *Ecography* 29, no. 2: 129–151. <https://doi.org/10.1111/j.2006.0906-7590.04596.x>.
- Erdelen, W. 1988. "Forest Ecosystems and Nature Conservation in Sri Lanka." *Biological Conservation* 43, no. 2: 115–135. [https://doi.org/10.1016/0006-3207\(88\)90086-9](https://doi.org/10.1016/0006-3207(88)90086-9).
- Erdelen, W. R. 2012. "Conservation of Biodiversity in a Hotspot: Sri Lanka's Amphibians and Reptiles." *Amphibian & Reptile Conservation* 5, no. 2: 33–51.
- Farr, T. G., P. A. Rosen, E. Caro, et al. 2007. "The Shuttle Radar Topography Mission." *Reviews of Geophysics* 45, no. 2: RG2004. <https://doi.org/10.1029/2005RG000183>.
- Feng, X., D. S. Park, C. Walker, A. T. Peterson, C. Merow, and M. Papeş. 2019. "A Checklist for Maximizing Reproducibility of Ecological Niche Models." *Nature Ecology & Evolution* 3, no. 10: 1382–1395. <https://doi.org/10.1038/s41559-019-0972-5>.
- Fernando, G. W. A. R., and P. N. Ranasinghe. 2009. "Dieback in Tropical Montane Forests of Sri Lanka: Anthropogenic or Natural Phenomenon?" *Journal of Geological Society of Sri Lanka* 13: 27–52.
- Fick, S. E., and R. J. Hijmans. 2017. "WorldClim 2: New 1-km Spatial Resolution Climate Surfaces for Global Land Areas." *International Journal of Climatology* 37, no. 12: 4302–4315. <https://doi.org/10.1002/joc.5086>.
- Fielding, A. H., and J. F. Bell. 1997. "A Review of Methods for the Assessment of Prediction Errors in Conservation Presence/Absence Models." *Environmental Conservation* 24, no. 1: 38–49. <https://doi.org/10.1017/S0376892997000088>.
- Fishpool, L. D. C., and I. M. Evans. 2003. *The Canadian Field-Naturalist* (Vol. 117, 4). Biology and Wildlife Department, University of Alaska. <https://doi.org/10.22621/cfn.v117i4.786>.
- Fourcade, Y., J. O. Engler, J. Secondi, and J. Secondi. 2014. "Mapping Species Distributions With MaxEnt Using a Geographically Biased Sample of Presence Data: A Performance Assessment of Methods for Correcting Sampling Bias." *PLoS One* 9, no. 5: e97122. <https://doi.org/10.1371/journal.pone.0097122>.
- Gafna, D. J., J. A. Obando, J. M. Kalwij, K. Dolos, and S. Schmidtlein. 2023. "Climate Change Impacts on the Availability of Anti-Malarial Plants in Kenya." *Climate Change Ecology* 5, no. 1: 30–44. <https://doi.org/10.1016/j.ecochg.2023.100070>.
- Gamage, S. N., D. K. Weerakoon, and A. Gunawardena. 2011. "Current Status of Vertebrate Diversity in Anthropogenic and Natural Ecosystems in South-western Sri Lanka." *Journal of the National Science Foundation of Sri Lanka* 39, no. 4: 383–389. <https://doi.org/10.4038/jnsfsr.v39i4.3886>.
- Gibson, C., A. de Silva, M. F. Tognelli, and S. Karunaratna. 2020. *Assess to Plan: Conservation Action Planning for the Snakes and Lizards of Sri Lanka*. IUCN Conservation Planning Specialist Group.
- Goodale, E., S. W. Kotagama, T. R. S. Raman, et al. 2014. "The Response of Birds and Mixed-Species Bird Flocks to Human-Modified Landscapes in Sri Lanka and Southern India." *Forest Ecology and Management* 329: 384–392. <https://doi.org/10.1016/j.foreco.2013.08.022>.
- Gray, C. L., S. L. L. Hill, T. Newbold, et al. 2016. "Local Biodiversity Is Higher Inside Than Outside Terrestrial Protected Areas Worldwide." *Nature Communications* 7: 12306. <https://doi.org/10.1038/ncomms12306>.
- Greig-Smith, P. 1964. *Quantitative Plant Ecology*. Butterworths.
- Guisan, A., and W. Thuiller. 2005. "Predicting Species Distribution: Offering More Than Simple Habitat Models." *Ecology Letters* 8, no. 9: 993–1009. <https://doi.org/10.1111/j.1461-0248.2005.00792.x>.
- Gunatilleke, C. V. S., and P. S. Ashton. 1987. "New Light on the Plant Geography of Ceylon. II. The Ecological Biogeography." *Journal of Biogeography* 14, no. 4: 295–327. <https://doi.org/10.2307/2844940>.

- Gunatilleke, N., S. Gunatilleke, and P. S. Ashton. 2017. "South-West Sri Lanka: A Floristic Refugium in South Asia." *Ceylon Journal of Science* 46, no. 5: 65–78. <https://doi.org/10.4038/cjs.v46i5.7454>.
- Gunatilleke, N., B. Madurapperuma, I. A. U. N. Gunatilleke, C. V. S. Gunatilleke, and M. A. A. B. Dilhan. 2005. "Plant Biogeography and Conservation of the South-Western Hill Forests of Sri Lanka." *Raffles Bulletin of Zoology* 12: 9–22.
- Gunatilleke, N., R. Pethiyagoda, and S. Gunatilleke. 2017. "Biodiversity of Sri Lanka." *Journal of National Science Foundation* 36: 25–62. <https://doi.org/10.4038/jnsfr.v36i0.8047>.
- Hanski, I. 2011. "Habitat Loss, the Dynamics of Biodiversity, and a Perspective on Conservation." *Ambio* 40, no. 3: 248–255. <https://doi.org/10.1007/s13280-011-0147-3>.
- Hijmans, R. J., S. E. Cameron, J. L. Parra, P. G. Jones, and A. Jarvis. 2005. "Very High Resolution Interpolated Climate Surfaces for Global Land Areas." *International Journal of Climatology* 25, no. 15: 1965–1978. <https://doi.org/10.1002/joc.1276>.
- Hirzel, A. H., and G. Le Lay. 2008. "Habitat Suitability Modelling and Niche Theory." *Journal of Applied Ecology* 45, no. 5: 1372–1381. <https://doi.org/10.1111/j.1365-2664.2008.01524.x>.
- Hirzel, A. H., G. Le Lay, V. Helfer, C. Randin, and A. Guisan. 2006. "Evaluating the Ability of Habitat Suitability Models to Predict Species Presences." *Ecological Modelling* 199, no. 2: 142–152. <https://doi.org/10.1016/j.ecolmodel.2006.05.017>.
- Ishwar, N. M., R. Chellam, A. Kumar, and B. R. Noon. 2003. "The Response of Agamid Lizards to Rainforest Fragmentation in the Southern Western Ghats, India." *Conservation and Society* 1, no. 2: 69–86.
- Jayasekara, D., D. Mahaulpatha, E. Jayasekara, and A. De Silva. 2018. "Habitat Utilization of Endangered Rhino Horned Lizard (*Ceratophora stoddartii*) (Sauria: Agamidae) in the Horton Plains National Park, Sri Lanka." *Journal of Entomology and Zoology Studies* 6, no. 4: 1544–1549.
- Jeliazkov, A., Y. Gavish, C. J. Marsh, et al. 2022. "Sampling and Modelling Rare Species: Conceptual Guidelines for the Neglected Majority." *Global Change Biology* 28, no. 12: 3754–3777. <https://doi.org/10.1111/gcb.16114>.
- Johnson, C. J., and M. P. Gillingham. 2005. "An Evaluation of Mapped Species Distribution Models Used for Conservation Planning." *Environmental Conservation* 32, no. 2: 117–128. <https://doi.org/10.1017/S0376892905002171>.
- Johnston, G. R., M. S. Y. Lee, and T. D. Surasinghe. 2013. "Morphology and Allometry Suggest Multiple Origins of Rostral Appendages in Sri Lankan Agamid Lizards." *Journal of Zoology* 289, no. 1: 1–9. <https://doi.org/10.1111/j.1469-7998.2012.00962.x>.
- Karunarathna, D. M. S. S., and A. A. T. Amarasinghe. 2011. "Reptile Diversity of a Fragmented Lowland Rain Forest Patch in Kukulugala, Ratnapura District, Sri Lanka." *Taprobanica* 2, no. 2: 86–94. <https://doi.org/10.4038/tapro.v2i2.3146>.
- Karunarathna, D. M. S. S., and A. A. T. Amarasinghe. 2012. "Reptile Diversity in Beraliya Mukalana Proposed Forest Reserve, Galle District, Sri Lanka." *Taprobanica* 4, no. 1: 20–26. <https://doi.org/10.4038/tapro.v4i1.4378>.
- Karunarathna, S., N. A. Poyarkov, C. Amarasinghe, et al. 2020. "A New Species of the Genus *Ceratophora* Gray, 1835 (Reptilia: Agamidae) From a Lowland Rainforest in Sri Lanka, With Insights on Rostral Appendage Evolution in Sri Lankan Agamid Lizards." *Amphibian & Reptile Conservation* 14, no. 3: 103–126. <https://doi.org/10.5281/zenodo.13258697>.
- Kass, J. M., R. Muscarella, P. J. Galante, et al. 2021. "ENMeval 2.0: Redesigned for Customizable and Reproducible Modeling of Species' Niches and Distributions." *Methods in Ecology and Evolution* 12, no. 9: 1602–1608. <https://doi.org/10.1111/2041-210X.13628>.
- Kottawa-Arachchi, J. D., and M. A. Wijeratne. 2017. "Climate Change Impacts on Biodiversity and Ecosystems in Sri Lanka: A Review." *Nature Conservation Research* 2, no. 3: 2–22. <https://doi.org/10.24189/ncr.2017.042>.
- Krebs, C. J. 1989. *Ecological Methodology*, 654. Harper and Row Publishers Inc.
- Liu, C., P. M. Berry, T. P. Dawson, and R. G. Pearson. 2005. "Selecting Thresholds of Occurrence in the Prediction of Species Distributions." *Ecography* 28, no. 3: 385–393. <https://doi.org/10.1111/j.0906-7590.2005.03957.x>.
- Lovejoy, T. E. 2006. "Protected Areas: A Prism for a Changing World." *Trends in Ecology & Evolution* 21, no. 6: 329–333. <https://doi.org/10.1016/j.tree.2006.04.005>.
- Macey, J. R., J. A. Schulte, A. Larson, et al. 2000. "Evaluating Trans-Tethys Migration: An Example Using Acrodont Lizard Phylogenetics." *Systematic Biology* 49, no. 2: 233–256. <https://doi.org/10.1093/sysbio/49.2.233>.
- MacKenzie, D. I., and J. D. Nichols. 2004. "Occupancy as a Surrogate for Abundance Estimation." *Animal Biodiversity and Conservation*, 27, no. 1: 461–467. <https://doi.org/10.32800/abc.2004.27.0461>.
- Margules, C. R., and R. L. Pressey. 2000. "Systematic Conservation Planning." *Nature* 405: 243–253. <https://doi.org/10.1038/35012251>.
- McCoy, E. D., and S. S. Bell. 1991. *Habitat Structure: The Evolution and Diversification of a Complex Topic*. Integrative Biology Faculty and Staff Publications. https://doi.org/10.1007/978-94-011-3076-9_1.
- Megía-Palma, R., L. Arregui, I. Pozo, et al. 2020. "Geographic Patterns of Stress in Insular Lizards Reveal Anthropogenic and Climatic Signatures." *Science of the Total Environment* 749: 141655. <https://doi.org/10.1016/j.scitotenv.2020.141655>.
- Mendes, P., S. J. E. Velazco, A. F. A. Andrade, and P. De Marco. 2020. "Dealing With Overprediction in Species Distribution Models: How Adding Distance Constraints Can Improve Model Accuracy." *Ecological Modelling* 431: 109180. <https://doi.org/10.1016/j.ecolmodel.2020.109180>.
- Merow, C., M. J. Smith, and J. A. Silander. 2013. "A Practical Guide to MaxEnt for Modeling Species' Distributions: What It Does, and Why Inputs and Settings Matter." *Ecography* 36, no. 10: 1058–1069. <https://doi.org/10.1111/j.1600-0587.2013.07872.x>.
- Miththapala, S. 2016. "Conservation Revisited." *Ceylon Journal of Science* 44, no. 2: 1–26. <https://doi.org/10.4038/cjsbs.v44i2.7347>.
- MOE. 2012. *The National Red List 2012 of Sri Lanka: Conservation Status of the Fauna and Flora*. Ministry of Environment.
- Muscarella, R., P. J. Galante, M. Soley-Guardia, et al. 2014. "ENMeval: An R Package for Conducting Spatially Independent Evaluations and Estimating Optimal Model Complexity for Maxent Ecological Niche Models." *Methods in Ecology and Evolution* 5, no. 11: 1198–1205. <https://doi.org/10.1111/2041-210X.12261>.
- Myers, N., R. A. Mittermeier, C. G. Mittermeier, G. A. B. da Fonseca, and J. Kent. 2000. "Biodiversity Hotspots for Conservation Priorities." *Nature* 403, no. 6772: 853–858. <https://doi.org/10.1038/35002501>.
- Neate-Clegg, M. H. C., and M. W. Tingley. 2023. "Building a Mechanistic Understanding of Climate-Driven Elevational Shifts in Birds." *PLoS Climate* 2, no. 3: e0000174. <https://doi.org/10.1371/journal.pclm.0000174>.
- Nekaris, K., A. Arnell, and M. Svensson. 2015. "Selecting a Conservation Surrogate Species for Small Fragmented Habitats Using Ecological Niche Modelling." *Animals* 5, no. 1: 27–40. <https://doi.org/10.3390/ani5010027>.
- Nichols, J. D., L. L. Bailey, A. F. O'Connell Jr., et al. 2008. "Multi-Scale Occupancy Estimation and Modelling Using Multiple Detection Methods." *Journal of Applied Ecology* 45, no. 5: 1321–1329. <https://doi.org/10.1111/j.1365-2664.2008.01509.x>.

- Olson, D. M., and E. Dinerstein. 1998. "The Global 200: A Representation Approach to Conserving the Earth's Most Biologically Valuable Ecoregions." *Conservation Biology* 12, no. 3: 502–515. <https://doi.org/10.1046/j.1523-1739.1998.012003502.x>.
- Oro, D., M. Genovart, G. Tavecchia, M. S. Fowler, and A. Martínez-Abraín. 2013. "Ecological and Evolutionary Implications of Food Subsidies From Humans." *Ecology Letters* 16, no. 12: 1501–1514. <https://doi.org/10.1111/ele.12187>.
- Otto, C. R. V., and G. J. Roloff. 2011. "Using Multiple Methods to Assess Detection Probabilities of Forest-floor Wildlife." *Journal of Wildlife Management* 75, no. 2: 423–431. <https://doi.org/10.1002/jwmg.63>.
- Parmesan, C., and G. Yohe. 2003. "A Globally Coherent Fingerprint of Climate Change Impacts Across Natural Systems." *Nature* 421: 37–42. <https://doi.org/10.1038/nature01286>.
- Pawar, S., M. S. Koo, C. Kelley, M. F. Ahmed, S. Chaudhuri, and S. Sarkar. 2007. "Conservation Assessment and Prioritization of Areas in Northeast India: Priorities for Amphibians and Reptiles." *Biological Conservation* 136, no. 3: 346–361. <https://doi.org/10.1016/j.biocon.2006.12.012>.
- Pearce, J., and S. Ferrier. 2001. "The Practical Value of Modelling Relative Abundance of Species for Regional Conservation Planning: A Case Study." *Biological Conservation* 98, no. 1: 33–43. [https://doi.org/10.1016/S0006-3207\(00\)00139-7](https://doi.org/10.1016/S0006-3207(00)00139-7).
- Pearson, R. G. 2016. "Reasons to Conserve Nature." *Trends in Ecology & Evolution* 31, no. 5: 366–371. <https://doi.org/10.1016/j.tree.2016.02.005>.
- Pearson, R. G., C. J. Raxworthy, M. Nakamura, and A. Townsend Peterson. 2007. "Original article: Predicting Species Distributions From Small Numbers of Occurrence Records: A Test Case Using Cryptic Geckos in Madagascar." *Journal of Biogeography* 34, no. 1: 102–117. <https://doi.org/10.1111/j.1365-2699.2006.01594.x>.
- Perera, A. 2001. "Secondary Forest Situation in Sri Lanka: A Review." *Journal of Tropical Forest Science* 13, no. 4: 768–785.
- Peterson, A. T. 2001. "Predicting Species' Geographic Distributions Based on Ecological Niche Modeling." *Condor* 103, no. 3: 599–605. <https://doi.org/10.1093/condor/103.3.599>.
- Pethiyagoda, R. 1994. "Threats to the Indigenous Freshwater Fishes of Sri Lanka and Remarks on Their Conservation." *Hydrobiologia* 285: 189–201. https://doi.org/10.1007/978-94-011-0958-1_19.
- Pethiyagoda, R. 2012. "Biodiversity Conservation in Sri Lanka's Novel Ecosystems." *Ceylon Journal of Science* 41, no. 1: 1–10. <https://doi.org/10.4038/cjsbs.v41i1.4532>.
- Pethiyagoda Jr., R. S., and K. Manamendra-Arachchi. 2012. "Endangered Anurans in a Novel Forest in the Highlands of Sri Lanka." *Wildlife Research* 39, no. 7: 641–648. <https://doi.org/10.1071/WR12079>.
- Phillips, S. J., and R. E. Schapire. 2004. "A Maximum Entropy Approach to Species Distribution Modeling." In *Proceedings of the Twenty-First International Conference on Machine Learning*. Association for Computing Machinery. <https://doi.org/10.1145/1015330.1015412>.
- Pressey, R. L., M. Cabeza, M. E. Watts, R. M. Cowling, and K. A. Wilson. 2007. "Conservation Planning in a Changing World." *Trends in Ecology & Evolution* 22, no. 11: 583–592. <https://doi.org/10.1016/j.tree.2007.10.001>.
- Radosavljevic, A., and R. P. Anderson. 2014. "Making Better Maxent Models of Species Distributions: Complexity, Overfitting and Evaluation." *Journal of Biogeography* 41, no. 4: 629–643. <https://doi.org/10.1111/jbi.12227>.
- Ranasinghe, P. N., C. B. Dissanayake, D. V. N. Samarasinghe, and R. Galappatti. 2007. "The Relationship Between Soil Geochemistry and Die Back of Montane Forests in Sri Lanka: A Case Study." *Environmental Geology* 51, no. 6: 1077–1088. <https://doi.org/10.1007/s00254-006-0399-6>.
- Razgour, O., J. Hanmer, and G. Jones. 2011. "Using Multi-Scale Modelling to Predict Habitat Suitability for Species of Conservation Concern: The Grey Long-Eared Bat as a Case Study." *Biological Conservation* 144, no. 12: 2922–2930. <https://doi.org/10.1016/j.biocon.2011.08.010>.
- Remya, K., A. Ramachandran, and S. Jayakumar. 2015. "Predicting the Current and Future Suitable Habitat Distribution of *Myristica dactyloides* Gaertn. Using MaxEnt Model in the Eastern Ghats, India." *Ecological Engineering* 82: 184–188. <https://doi.org/10.1016/j.ecoleng.2015.04.053>.
- Roberts, D. R., V. Bahn, S. Ciuti, et al. 2017. "Cross-Validation Strategies for Data With Temporal, Spatial, Hierarchical, or Phylogenetic Structure." *Ecography* 40, no. 8: 913–929. <https://doi.org/10.1111/ecog.02881>.
- Rodrigues, A. S. L., H. R. Akçakaya, S. J. Andelman, et al. 2004. "Global Gap Analysis: Priority Regions for Expanding the Global Protected-Area Network." *Bioscience* 54, no. 12: 1092–1100. [https://doi.org/10.1641/0006-3568\(2004\)054\[1092:GGAPRF\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2004)054[1092:GGAPRF]2.0.CO;2).
- Rodrigues, A. S. L., S. J. Andelman, M. I. Bakarr, et al. 2004. "Effectiveness of the Global Protected Area Network in Representing Species Diversity." *Nature* 428, no. 6983: 640–643. <https://doi.org/10.1038/nature02422>.
- Roll, U., A. Feldman, M. Novosolov, et al. 2017. "The Global Distribution of Tetrapods Reveals a Need for Targeted Reptile Conservation." *Nature Ecology & Evolution* 1, no. 11: 1677–1682. <https://doi.org/10.1038/s41559-017-0332-2>.
- Rubenstein, M. A., S. R. Weiskopf, R. Bertrand, et al. 2023. "Climate Change and the Global Redistribution of Biodiversity: Substantial Variation in Empirical Support for Expected Range Shifts." *Environmental Evidence* 12, no. 7: 7. <https://doi.org/10.1186/s13750-023-00296-0>.
- Scherr, S. J., and J. A. McNeely. 2008. "Biodiversity Conservation and Agricultural Sustainability: Towards a New Paradigm of 'Eco Agriculture' Landscapes." *Philosophical Transactions of the Royal Society, B: Biological Sciences* 363, no. 1491: 477–494. <https://doi.org/10.1098/rstb.2007.2165>.
- Schnase, J. L., M. L. Carroll, R. L. Gill, et al. 2021. "Toward a Monte Carlo Approach to Selecting Climate Variables in MaxEnt." *PLoS One* 16, no. 3: e0237208. <https://doi.org/10.1371/journal.pone.0237208>.
- Schulte, J. A., J. R. Macey, R. Pethiyagoda, and A. Larson. 2002. "Rostral Horn Evolution Among Agamid Lizards of the Genus *Ceratophora* Endemic to Sri Lanka." *Molecular Phylogenetics and Evolution* 22, no. 1: 111–117. <https://doi.org/10.1006/mpev.2001.1041>.
- Senadheera, D. K. L., W. M. P. S. B. Wahala, and S. Weragoda. 2019. "Livelihood and Ecosystem Benefits of Carbon Credits Through Rainforests: A Case Study of Hiniduma Bio-Link, Sri Lanka." *Ecosystem Services* 37: 100933. <https://doi.org/10.1016/j.ecoser.2019.100933>.
- Somaweera, R., N. Wijayathilaka, G. Bowatte, and M. Meegaskumbura. 2015. "Conservation in a Changing Landscape: Habitat Occupancy of the Critically Endangered Tennent's Leaf-nosed Lizard (*Ceratophora tennentii*) in Sri Lanka." *Journal of Natural History* 49, no. 31/32: 1961–1985. <https://doi.org/10.1080/00222933.2015.1006280>.
- Surasinghe, T., R. Kariyawasam, H. Sudasinghe, and S. Karunarathna. 2020. "Challenges in Biodiversity Conservation in a Highly Modified Tropical River Basin in Sri Lanka." *Water* 12, no. 1: 26. <https://doi.org/10.3390/w12010026>.
- Surasinghe, T. D. 2009. "Conservation and Distribution Status of Amphibian Fauna in Sri Lanka." *Biodiversity* 10, no. 1: 3–17. <https://doi.org/10.1080/14888386.2009.9712632>.
- Tobefia, M., R. Prieto, M. Machete, and M. A. Silva. 2016. "Modeling the Potential Distribution and Richness of Cetaceans in the Azores From Fisheries Observer Program Data." *Frontiers in Marine Science* 3: 202. <https://doi.org/10.3389/fmars.2016.00202>.
- UNEP-WCMC and IUCN. 2020. *Protected Planet Report 2020*. UNEP-WCMC and IUCN. <https://protectedplanetreport2020.protectedplanet.net/>.

- Valavi, R., G. Guillera-Arroita, J. J. Lahoz-Monfort, and J. Elith. 2022. "Predictive Performance of Presence-Only Species Distribution Models: A Benchmark Study With Reproducible Code." *Ecological Monographs* 92, no. 1: e01486. <https://doi.org/10.1002/ecm.1486>.
- Walther, G. R., E. Post, P. Convey, et al. 2002. "Ecological Responses to Recent Climate Change." *Nature* 416: 389–395. <https://doi.org/10.1038/416389a>.
- Whitford, A. M., B. R. Shipley, and J. L. McGuire. 2024. "The Influence of the Number and Distribution of Background Points in Presence-Background Species Distribution Models." *Ecological Modelling* 488: 110604. <https://doi.org/10.1016/j.ecolmodel.2023.110604>.
- Wijerathne, I., Y. Deng, E. Goodale, et al. 2025. "The Vulnerability of Endemic Vertebrates in Sri Lanka to Climate Change." *Global Ecology and Conservation* 59: e03515. <https://doi.org/10.1016/j.gecco.2025.e03515>.
- Wilson, K. A., M. I. Westphal, H. P. Possingham, and J. Elith. 2005. "Sensitivity of Conservation Planning to Different Approaches to Using Predicted Species Distribution Data." *Biological Conservation* 122, no. 1: 99–112. <https://doi.org/10.1016/j.biocon.2004.07.004>.
- Wisz, M. S., R. J. Hijmans, J. Li, et al. 2008. "Effects of Sample Size on the Performance of Species Distribution Models." *Diversity and Distributions* 14, no. 5: 763–773. <https://doi.org/10.1111/j.1472-4642.2008.00482.x>.
- Zhang, Z., J. M. Kass, S. Mammola, et al. 2021. "Lineage-Level Distribution Models Lead to More Realistic Climate Change Predictions for a Threatened Crayfish." *Diversity and Distributions* 27, no. 4: 684–695. <https://doi.org/10.1111/ddi.13225>.

Supporting Information

Additional supporting information can be found online in the Supporting Information section.