RESEARCH ARTICLE



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The reconstructed evolutionary history of the *Engelhardia* spicata complex highlights the impact of a three-tiered landform in the Indo-Burma ecoregion

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Abstract

Premise: The lateral displacement of the Indochina Peninsula, driven by the Indian—Asian plate collision, significantly altered the topography of the Indo-Burma ecoregion, affecting its climate and biological evolution. Despite the renowned biodiversity of the region, spatiotemporal patterns of evolution remain poorly understood.

Methods: We analyzed the *Engelhardia spicata* complex, which has a continuous distribution across Indo-Burma, based on a robust phylogenetic framework comprising 778 individuals from 80 populations, to elucidate spatiotemporal and paleogeological patterns of evolution. We used ancestral area reconstruction to reconstruct the historical biogeography of the species complex and to understand the broader evolutionary history of the Indo-Burma ecoregion.

Results: An initial divergence within the *E. spicata* complex approximately 26.62 million years ago (Ma) separated a lineage in the Truong Son Mountain Range from one in the Hengduan Mountains and the Shan Plateau. The Shan Plateau and Hengduan Mountain lineages subsequently diverged around 23.03 Ma. These results highlight a three-tiered landform in the Indo-Burma ecoregion, characterized by high-elevation northern regions (Hengduan Mountains, Yunnan-Guizhou Plateau), intermediate-elevation central plateau (Shan Plateau), and low-elevation southern ranges (southern Truong Son Mountains).

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Conclusions: Our findings support the tectonic hypothesis that crustal thickening and lateral extrusion of Indochina occurred simultaneously during the Late Oligocene, which led to the formation of the Indo-Burma ecoregion and highlights the biological significance of the resulting three-tiered landform (north-to-south altitudinal gradients) in these regions, providing novel insights into biogeographic patterns in Southeast Asia.

KEYWORDS

biogeographic patterns, *Engelhardia spicata* complex, Indo-Burma ecoregion, Juglandaceae, spatiotemporal history, three-tiered landform

During the Cenozoic era, one of the most significant tectonic events on Earth was the collision between the Eurasian and Indian plates, which led to the formation of the Qinghai-Tibet Plateau (QTP), encompassing the Hengduan Mountains, the plateau platform (including the Pamir Mountains, Karakoram Mountains, Kunlun Mountains, Qaidam Basin, and Qilian Mountains), and the Himalayas (Mao et al., 2021). This collision also caused lateral displacement of the Indochina Peninsula (Yin and Harrison, 2000; Li et al., 2024; Huang et al., 2025). The collision between the Indian and Eurasian tectonic plates and the initial rapid uplift of the QTP are widely accepted to have occurred during the Paleocene (Ding et al., 2022), with subsequent localized uplift during the Oligocene, Miocene and even Pliocene epochs (Luo et al., 2022; Lai et al., 2025a, 2025b). This sustained uplift induced clockwise rotation and southeastward extrusion of the Indochina Peninsula along the Ailao Shan-Red River shear zone, as described by a two-phase acceleration-deceleration model (Liu et al., 2012; Li et al., 2024). This extrusion may also have contributed to the north-high-south-low topographic structure of the Indochina Peninsula. The ongoing extension of the Ailao Shan-Red River shear zone toward southern Vietnam may have played a significant role in shaping the modern Truong Son Mountain Range. During the same period, widespread uplift in the western part of China resulted in high western and low eastern terrain features that have since characterized the region's topography (Che et al., 2010). This geomorphological pattern prompts a critical question: Did similar tectonic processes such as crustal thickening and lateral extrusion similarly shape the adjacent Indo-Burma ecoregion?

The Indo-Burma ecoregion, situated in northern Southeast Asia, is recognized as a distinct bioregion that encompasses the Indochina Peninsula and surrounding areas. The northern boundary of the region is defined by the Eastern Himalayas and subtropical regions (Meng et al., 2025) and the southern boundary by the Isthmus of Kra (Meng and Song, 2023). This biological subregion serves as a crucial link between Eurasia and the Malay Archipelago, facilitating regional biodiversity convergence, diffusion, and migration (Meng and Song, 2023). The tiered topography (i.e., high-elevation northern ranges, intermediate plateaus, and southern lowlands) of the region mirrors the geomorphological gradient in western China. The complex topography of this crucial biogeographic corridor

between Eurasia and the Malay Archipelago has facilitated biodiversity convergence, diffusion, and migration (Meng and Song, 2023). This global biodiversity hotspot (Myers et al., 2000) is home to over 13,500 plant species, of which 7000 are endemic (van Dijk et al., 1999; Mittermeier et al., 2005) and consequently, of significant importance for conserving global biodiversity. The extraordinary biodiversity can be attributed to a combination of complex geological and climatic histories, combined with habitat diversity driven by variations in topography, climate, and latitude (Mittermeier et al., 2002). The intricate geological events since the Paleocene and extending well into the Miocene have profoundly influenced climate and vegetation changes across Southeast Asia (Kooyman et al., 2019), shaping the origin, evolution, and geographical distribution patterns of biodiversity in the region. Understanding the geographic distribution patterns of organisms in the Indo-Burma ecoregion better should enhance biodiversity conservation efforts from evolutionary and ecological perspectives (Woodruff, 2010). Biogeographical studies in the Indo-Burma ecoregion to date have focused primarily on animals, including the families Ichthyophiidae (Gower et al., 2002), Dicroglossidae (Bossuyt and Milinkovitch, 2001; Che et al., 2010), and Ranidae (Wu et al., 2020). Phytogeographic research has focused on the extrusion of the Indo-Burma ecoregion (Zhao et al., 2016; Meng et al., 2022b; Huang et al., 2023; Li et al., 2024; Zhang et al., 2024). However, the role of topographical dynamics in shaping biogeographic processes across the broader Indo-Burma ecoregion, particularly its elevational gradient from the northern highlands to the southern lowlands, is poorly understood. This elevational gradient parallels the "three-tiered" topography of China (Figure 1), where stepwise geological uplift has profoundly influenced species diversification (Lee, 1939; Clark et al., 2005). Whether similar geomorphological processes shaped the Indo-Burma ecoregion is debated, with limited biological evidence linking extant species distributions to tectonic history (Maneerat and Bürgmann, 2022). We hypothesized that the biogeographic patterns of Engelhardia Leschenault ex Blume a lineage of trees within the Juglandaceae (walnut family) now recognized as a species complex (Manos et al., 2007; Meng et al., 2022b)—potentially mirror the phased uplift of the Indo-Burma terrain. By analyzing the spatial genetic structure in this complex, we aimed to disentangle how Cenozoic geological evolution may have driven diversification in this ecoregion.

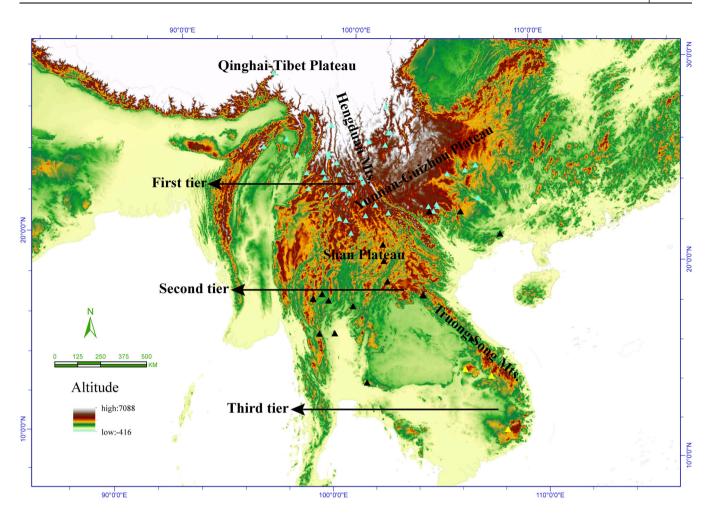


FIGURE 1 Sample distribution and results for plastid DNA sequences. Colored triangles indicate different lineages (blue: group A, black: group B, yellow: group C. The three-tiered landform of the Indo-Burma ecoregion is shown (average elevations a.s.l.: first tier, 4000–5000 m; second tier, 1000–2000 m; third tier, <500 m.

Engelhardia has a long evolutionary history in the region and provides critical insights for reconstructing biogeographic patterns across tropical and subtropical Asia (Song et al., 2020; Huang et al., 2024). Molecular and morphological data indicate that there are nine species in Engelhardia, six of which are found in the Indo-Burma ecoregion (Zhang et al., 2020; Meng et al., 2022a). Engelhardia spicata is the most widely distributed species. Our previous research demonstrated that the relationship between E. spicata and its constituent taxa is complex; the complex includes E. spicata var. aceriflora, E. spicata var. colebrookeana, and E. rigida (Zhang et al., 2020). The Engelhardia spicata complex is primarily distributed in southwestern China (including Tibet, Yunnan, Sichuan, Guangxi, and Guizhou), Myanmar, Thailand, Vietnam, Laos, Indonesia, Malaysia, Nepal, and the Philippines (Zhang et al., 2020). The complex lives at elevations ranging from 142 to 1880 m a.s.l. and is commonly found on slopes within mixed broad-leaf forests or at the forest edges (Appendix S1: Table S1). The distinctive distribution pattern of the complex, spanning diverse topographical gradients and geological formations provides a valuable model for investigating the impact of geological events in the Indo-Burma ecoregion.

An exploration of spatial and temporal changes in the staircase landform in the Indo-Burma ecoregion should enhance our understanding of causes and mechanisms related to the formation and evolution of this natural environment during the Cenozoic era. Our study addressed two key questions: (1) When did the three-tiered landform of the Indo-Burma ecoregion begin to form and influence distribution patterns of Engelhardia spicata complex? (2) How did the geological processes contribute to the formation of the three-tiered landform and to the distribution patterns of E. spicata complex in particular? To address these questions, we characterized the phylogeography of the E. spicata complex based on alignments of five plastid DNA regions and nuclear ribosomal (nrDNA) data from 80 populations across the Indo-Burma ecoregion, to illuminate the spatiotemporal evolutionary patterns of plants in the Indo-Burma ecoregion and investigate the topographic structure of the region from a biological perspective.

MATERIALS AND METHODS

Sample collection and DNA analysis

We collected a total of 778 individuals of the E. spicata complex from 80 populations across the Indo-Burma ecoregion (Figure 2; Appendix S1: Table S1). The sampling locations span the Indo-Burma ecoregion and parts of southern China and encompass a diverse range of topographical gradients and geological formations. We obtained meta-data relevant to these areas from various published (Manning, 1966; Flora of China, Lu et al., 1999) and online sources (Chinese Virtual Herbarium [CVH; https://www.cvh. ac.cn]; Global Biodiversity Information Facility [GBIF; https://www.gbif.org]). We extracted total genomic DNA for these samples using a Plant Genomic DNA Kit (Tiangen Biotech, Beijing, China). We recovered five plastid DNA regions (psbA-trnH, trnL-trnF, rps16, trnS-trnG, and rpl32trnL) based on laboratory protocols detailed in Appendix S1: Table S2. We concatenated the five plastid fragments using Geneious v6.1.2 (https://www.geneious.com/). We then used

DnaSP v6 (Rozas et al., 2017) to identify and determine haplotypes and ITS genotypes. Table S1 in Appendix S1 details the number and distribution of different haplotypes and ribotypes across populations. We determined haplotype diversity (Hd) and the selective neutrality parameters Tajima's D (Tajima, 1989) and Fu and Li's D^* (Fu and Li, 1993; Fu, 1997) using DnaSP v6 for the combined plastid DNA haplotype (H) and ITS ribotype (R). And employed Arlequin v3.1 (Excoffier et al., 2007) for an analysis of molecular variance (AMOVA) (Excoffier et al., 1992) using both the concatenated plastid DNA sequences and ITS data. We assessed genetic variation within and among populations using Φ - and R-statistics. We assessed the significance of fixation indices using 1000 permutations and tested a null hypothesis of spatial expansion using mismatch distribution analysis (MDA) in DnaSP v6. We assessed the goodness-offit of population demographic history by calculating the sum of squared deviations between the observed and expected mismatch distributions and the raggedness index (HRag). These calculations were based on 1000 parametric bootstrap replicates (Harpending, 1994).

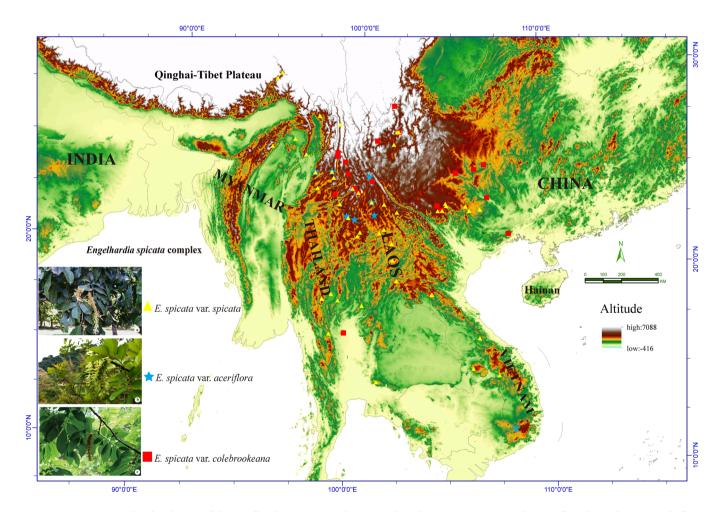


FIGURE 2 Geographic distribution of the *Engelhardia spicata* complex across the Indo-Burma ecoregion and parts of southern China. A total of 80 locations were studied across. Sample localities: red triangles, *E. spicata* var. *spicata*; black stars, *E. spicata* var. *aceriflora*; blue squares, *E. spicata* var. *colebrookeana*.

DNA sequence phylogenetic analyses

We inferred the lineage relationships between *H* and *R* from a median-joining network using NETWORK v2.0 (Bandelt et al., 1999). Separately, based on previous phylogenetic inferences in Juglandaceae (Manos and Stone, 2001; Manos et al., 2007; Zhang et al., 2020; Meng et al., 2022a), we selected Engelhardia fenzelii, E. roxburghiana, E. hainanensis, E. villosa, E. serrata, E. apoensis, and Rhoiptelea chiliantha as outgroups. We inferred phylogenies using Bayesian inference for the plastid DNA and ITS data sets with MrBayes v3.1.2 (Huelsenbeck and Ronquist, 2001). We determined the best-fit DNA substitution model for each data set using the Akaike information criterion (AIC) and PAUP* version 4.0b10 (Swofford, 2002) and MrModeltest 2.4 (Nylander, 2004). The optimal substitution models proposed for the combined plastid DNA and nrDNA data were GTR + I + G and GTR + G, respectively. For the Bayesian analyses, we ran the MCMC algorithm for 5×10^6 generations, using a cold chain and three heated chains, starting from random trees, sampling every 5000 generations and discarding the first 10% as burn-in. All effective sample size (ESS) values were >200. We also used IQ-TREE v2.1.1 (Minh et al., 2020) for maximum likelihood (ML) analyses, including a model selection analysis for each data set. To identify the best ML tree, we ran five independent analyses, each searching for the optimal tree topology. Subsequently, we ran a single analysis with 1000 ultrafast bootstrap replicates (Minh et al., 2020).

Divergence time estimation

To avoid potential biases due to recombination and incomplete concerted evolution in nuclear sequences on age estimates (Álvarez and Wendel, 2003; Emadzade and Hörandl, 2011), we used the plastid data rather than the combined plastid and nuclear data. For comparative purposes, we also estimated divergence times using the ITS data set. We analyzed these data sets using BEAST v1.8.1, which implements the Bayesian MCMC algorithm for phylogenetic inference and molecular dating (Drummond Rambaut, 2007). For these analyses, we used nucleotide substitution models identical to those described for the Bayesian phylogenetic analysis above. We compared four clock models (strict, uncorrelated relaxed, random, and fixed) and two tree speciation priors (Yule and birth-death) using Bayes factors (Kass and Raftery, 1995) calculated from marginal likelihoods derived from path sampling (Baele et al., 2013). The uncorrelated relaxed clock model (Appendix S1: Table S3) and Yule tree prior model (Appendix S1: Table S4) were identified as the best-fit models. For the uncorrelated relaxed clock model, we used a normal distribution to define the prior for the mean substitution rate parameter, and for the GTR + G model for each DNA region separately, we considered mean values for parameters of the distribution (Ho and Phillips, 2009). For

the MCMC analysis, we set the combined plastid DNA and ITS chain length to 3×10^8 generations, sampling every 10,000 generations. All options were configured using BEAUti v1.8.1 in BEAST.

We assessed convergence in Tracer v1.6 (Rambaut et al., 2014), and all ESS values were >200. After discarding the first 10% sampled trees as burn-in, we generated a maximum clade credibility (MCC) tree with median ages and 95% highest posterior density (HPD) intervals on nodes using TreeAnnotator v1.8.1 (Drummond and Rambaut, 2007), which we visualized using FigTree v1.4.4 (Rambaut, 2018). Based on phylogenetic analyses of both extant and extinct Engelhardioideae (Meng et al., 2015), we calibrated the dated tree using fossil records of Alatonucula ignis (Hermsen and Gandolfo, 2016) and Paleooreomunnea stoneana (Dilcher et al., 1976). Specifically, we used (1) Alatonucula ignis, the most reliable and oldest known fruit fossil record, found in South America during the Eocene (Hermsen and Gandolfo, 2016) with its ${}^{40}\text{Ar}/{}^{39}\text{Ar}$ age = 52.22 ± 0.22 Ma (Wilf, 2012; Barreda et al., 2020), to calibrate the stem group age of Engelhardioideae; (2) Paleooreomunnea stoneana, discovered in North America during the Eocene with an estimated age of 40.4-48.6 Ma, to calibrate the Engelhardioideae crown group (Dilcher et al., 1976; Sauquet et al., 2012; Zhang et al., 2013).

Ancestral range reconstruction and elevation analysis

We inferred ancestral distribution ranges of E. spicata complex using the R package BioGeoBEARS version (Matzke, 2013), along with plastid DNA fossil-calibrated tree data sets. Based on the weighted AIC corrected for sample size (AICc wt), we compared six biogeographical models: DEC, DEC+j, DIVALIKE, DIVALIKE+j, BAYAR-EALIKE, and BAYAREALIKE+j. The DEC model was determined to be optimal, exhibiting the highest AICc_wt (Appendix S1: Table S5), and was thus used for ancestral area reconstruction. To account for phylogenetic uncertainties, we conducted ancestral range reconstruction using the statistical DEC model as implemented in RASP v.4.2 (Ree and Smith, 2008; Yu et al., 2015). We loaded the MCC tree and 1000 randomly selected trees (including outgroups) from the BEAST analysis as input files. Based on the geographic divisions of Wen (2017), we defined four geographic areas: (A) the Hengduan Mountains and the Yunnan-Guizhou Plateau, (B) the Shan Plateau region, (C) the southern Truong Son Mountain Range, (D) all other regions except A, B, and C. Additionally, we generated 50 biogeographic stochastic maps in BioGeoBEARS (Matzke, 2013) to evaluate uncertainty in ancestral range estimation and biome transition dynamics (Appendix S1: Table S6).

In total, we collected data for 187 occurrences of *E. spicata* complex sites from GBIF, CVH, and new field records (Appendix S1: Table S7), excluding localities with

inaccurate or missing coordinates. For each record, we extracted the elevational variable from the calibration dataset for global land areas at 1-km resolution (Fick and Hijmans, 2017) using the function raster in the R package raster (Hijmans, 2020). We calculated the mean and boundaries of the 95% elevational range for each species. After confirming normality and homogeneity of variances, we used an ANOVA to assess interspecific differences in elevation ranges. IBM SPSS Statistics v22.0 (IBM, Armonk, NY, USA) was used for all calculations and box plots. A squared-change parsimony method is appropriate for analyzing continuous characters (Paradis et al., 2004), and so we inferred the ancestral elevation of the *E. spicata* complex with this method in Mesquite 3.61 (Maddison, 2019), using the MCC tree, with vertical lines of different colors representing the three genetic clusters. For comparative purposes, we also inferred the ancestral elevation using the phylogenetic independent contrast (PIC) method, a Brownianmotion based estimator. We conducted the PIC analysis using the function ace in the R package ape (Paradis et al., 2004). We also employed the phylogenetic generalized least squares (PGLS) method to analyze the ancestral elevation of the E. spicata complex because this method incorporates phylogenetic relationships to account for evolutionary relatedness among species (Grafen, 1989; Martins and Hansen, 1997).

RESULTS

Genetic diversity of plastid DNA and nrDNA

We sequenced a total of 778 samples from the E. spicata complex, yielding 724 plastid DNA sequences and 674 ITS sequences. The concatenated alignment of five plastid DNA spacers spans 4647 bp, revealing 51 haplotypes across all samples. The ITS consisted of 755 bp, leading to the identification of 31 ribotypes (Appendix S1: Table S1). We observed high levels of haplotype diversity (Hd = 0.9664) and ribotype diversity (Hd = 0.8675) across all regions. However, the subregional plastid DNA and ITS diversity values were lower than the overall diversity for all regions, and group A had higher diversity than groups B and C. We summarize the basic genetic data for all species across the major regions in Appendix S1 (Table S1). Variation in plastid haplotype and nuclear ribotypes within populations was greater than reported for E. roxburghiana and E. fenzelii (within the same genus) (Meng et al., 2022b): populations Y02, M02, Y03, Y05, Y35, Y43, L09, L13, L14, L15, A02, A03, A05, Y10, Y12, and Y18 each had two haplotypes present (the three-letter population codes for each population are defined in Appendix S1: Table S1). Populations M02, Y31, Y34, Y35, Y38, Y41, S08, Y42, L13, A02, A03, Y10, Y12, Y48, and Y49 each had two ribotypes (Appendix S1: Table S1). Most populations were fixed for a single plastid haplotype or nuclear ribotype (see the three-letter codes with an "H" or "R" prefix, respectively, Appendix S1: Table S1),

and some common haplotypes and ribotypes are shared among different populations, for example, plastid haplotypes H01, H02, H05, H06, H11, H15, H16, H17, H19, H20, H23, H25, H26, H29, H34, H37, and H44, and nuclear ribotypes R01, R02, R03, R04, R08, R10, R12, and R23 (Appendix S1: Table S1). The AMOVA based on plastid DNA haplotypes revealed significant and extensive genetic population differentiation ($\Phi_{ST} = 0.91862$, P < 0.001), with 49.86% of the genetic variation partitioned among the three regions and 41.99% among populations within those regions (Appendix S1: Table S8). The ITS ribotype Φ_{ST} value was 0.90452 (P < 0.001), with 31.80% of the genetic variation partitioned and 58.65% among populations within the regions. The estimates of Tajima's D and Fu's F_S are significant for all plastid DNA clades and subclades of the E. spicata complex (Appendix S1: Table S9). The observed mismatch distributions of plastid DNA for each clade/subclade were multimodal and/or highly ragged (Appendix S1: Figure S1), leading to rejection of the spatial expansion model in most cases.

Phylogenetic relationships

The ML tree of plastid DNA haplotypes revealed three distinct lineages, consistent with both BI trees (Appendix S1: Figure S2). Group A comprised populations from the Yunnan-Guizhou Plateau (China) and northern Myanmar, distributed primarily in the high-altitude Hengduan Mountains. Group B included populations predominantly located in the Shan Plateau and adjacent areas of Thailand, Laos, Vietnam, and parts of southern China. Group C comprised populations from the southern section of the Truong Son Mountain Range. The distribution of the plastid DNA phylogeographic structure exhibited a three-tiered geographic genetic pattern, characterized by high-elevation northern regions (the Hengduan Mountains and the Yunnan-Guizhou Plateau), intermediate-elevation central plateaus (the Shan Plateau), and low-elevation southern ranges (the southern Truong Son Mountain Range) (Figure 1). In contrast, nuclear ribotypes had no clear phylogeographic structure (Appendix S1: Figure S3).

Divergence time estimation

Divergence time estimation suggests that *E. fenzelii* and *E. roxburghiana* diverged from their sister lineage within *Engelhardia* by 45.06 Ma (plastid DNA; node ii, 95% HPD: 41.09–49.18 Ma; Table 1; Figures 3B) or 44.82 Ma (ITS; node ii, 95% HPD: 40.84–48.95 Ma; Table 1; Appendix S1: Figure S4). The *E. spicata* complex diverged from its sister lineage, *E. hainanensis* approximately 32.16 Ma (plastid DNA; node iii, 95% HPD: 22–41.97 Ma; Table 1; Figure 3B). The stem age of *E. spicata* group C was around 26.62 Ma (plastid DNA; node iv, 95% HPD: 17.24–36.23 Ma; Table 1;

TABLE 1 Median age and 95% highest posterior density (HPD) estimates in million year ago (Ma) for selected nodes based on plastid DNA and ITS.

| | Median age (HPD), Ma | |
|-----------------------------------|----------------------|---------------------|
| Node | Plastid DNA | ITS |
| (i) *Fossil calibration point | 52.22 (52.01–52.43) | 52.22 (52-52.43) |
| (ii) *Fossil calibration point | 45.06 (41.09–49.18) | 44.82 (40.84–48.95) |
| (iii) | 32.16 (22-41.97) | |
| (iv) | 26.62 (17.24–36.23) | |
| (v) | 23.03 (14.79–32.34) | |
| (vi) | 18.13 (10.72–26.92) | |
| (vii) | 15.5 (7.44–25.14) | |
| (viii) | 10.78 (1.13-23.78) | |

Figure 3C). The stem age of group A and group B was 23.03 Ma (plastid DNA; node v, 95% HPD: 14.79–32.34 Ma; Table 1; Figure 3C). In contrast, ITS failed to resolve intraspecific divergence events within the *E. spicata* complex, possibly due to incomplete lineage sorting or gene flow. Divergence timelines for major clades based on the plastid data are summarized in Figure 3C and Table 1.

Ancestral range reconstructions

Our ancestral range reconstruction suggests that the most recent common ancestor of the E. spicata complex occupied the Indo-Burma ecoregion (node iv; Figure 3B). Within the E. spicata complex, two vicariance events occurred (Figure 3C). One led to the divergence of group C in the southern Truong Son Mountain Range, and the most recent common ancestor of group A and group B (node v; Figure 3C). Another vicariance event separated group B in the Shan Plateau region from group A in the Hengduan Mountains and the Yunnan-Guizhou Plateau. Results from the two analytical approaches (BioGeoBEARS and RASP) had strong congruence, supporting the robustness of our reconstructions (Appendix S1: Figure S5). The BioGeo-BEARS stochastic mappings (DEC model; 50 replicates) indicated 108.1 ± 1.5 geographic range shifts. Of these, 69 ± 0 were cladogenetic (branching) events compared to 39.1 ± 1.45 anagenetic (within a single branch) dispersal events. Of the 69 branching events, in 52.96 ± 1.32 events, sister species maintained their ancestral biome ("narrow sympatry"), and in 9.96 ± 1.65 events, one sister species maintained its ancestral biome while the other sister occupied only a subset of the ancestral range ("subset sympatry"). The remaining branching events were made up of 6.08 ± 1.1 vicariance events in which the sister species split their ancestral range. Group C in the southern Truong Son

Mountain Range represented the largest source of outward dispersal (11.78 ± 5.7 events), followed by group A in the Hengduan Mountains and the Yunnan-Guizhou Plateau (10.32 ± 5.48 events; Figure 3D). In contrast, the largest sink for inward dispersals was group B in the Shan Plateau region (10.86 ± 4.82 events). The most significant directional transition occurred from group C to group B (4.72 ± 1.87 events; Figure 3D and Appendix S1: Table S6). Furthermore, network analysis of plastid DNA supported three major lineages of the *E. spicata* complex (Appendix S1: Figure S6). The network highlighted the complex genetic relationships between the haplotypes from the southern Truong Son Mountain Range (group C) and those from the other two clades, suggesting potential gene flow and incomplete lineage sorting within this group (Zhang et al., 2020).

Elevation differentiation

The altitude of the study area ranged from approximately sea level to 5000 m a.s.l. and had a stepped distribution (Figure 4A, B). The ancestral elevation reconstructions using the PIC and PGLS methods yielded highly consistent results (Appendix S1: Figure S7). We focus on the PIC results here. The E. spicata complex occurs at various elevations, categorized into group A (1351.66 m; 95% HPD: 1289.30-1414.03), group B (1048.65 m; 95% HPD: 880.75-1216.56) and group C (786.88 m; 95% HPD: 650.52-923.25). A variance homogeneity test indicated comparable within-group dispersion (P > 0.05), but the Kruskal-Wallis analysis revealed strong elevational differentiation among groups (P < 0.05). The strong elevational differentiation among groups (P < 0.05) suggests that altitude differentiation among the E. spicata complex was significant (Appendix S1: Table S10). Additionally, the reconstruction analysis of ancestral elevation demonstrated that the altitudes of the three groups also exhibited significant elevational differentiation (Figure 4D), i.e., group A at a mean altitude of 1291.6 m (95% HPD: 1281.86-1301.34), group B 1050.03 m (95% HPD: 1041.62-1058.44), and group C 763.5 m (95% HPD: 754.4-772.6).

DISCUSSION

Spatiotemporal evolution of *E. spicata* complex

Tectonic events have profoundly influenced biodiversity and geographical distribution patterns in the Indo-Burma ecoregion (Janssens et al., 2016; Herrera et al., 2021). Furthermore, the evolutionary diversification of organisms may mirror geological and climatic changes (Che et al., 2010; Xiang et al., 2021; Zhang et al., 2023; Zhang et al., 2025). Our molecular dating and geological analyses here not only offer a novel horizon to understanding the evolution of the *E. spicata* complex, but also provide broader insights into the historical biogeography reconstruction of the Indo-Burma ecoregion.

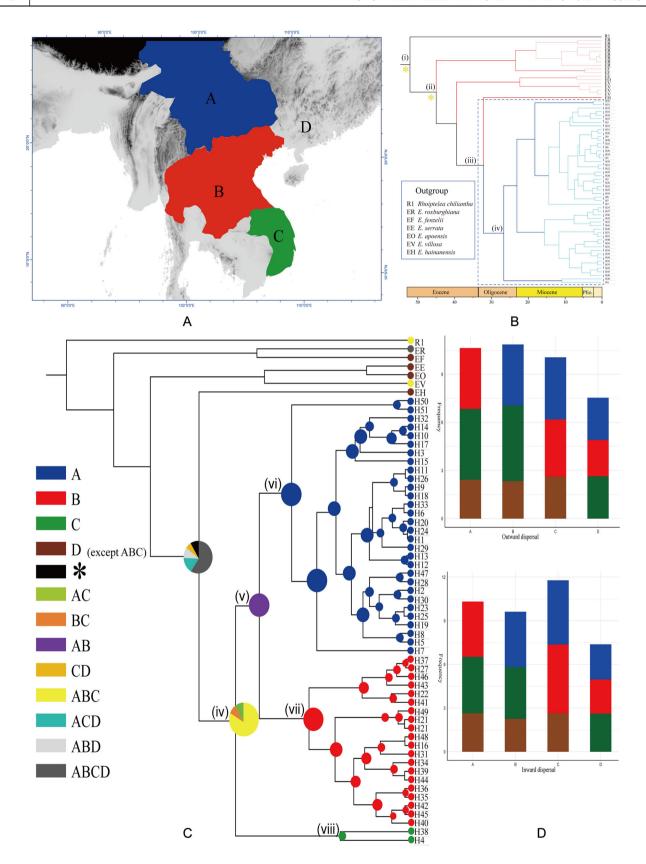


FIGURE 3 Combined chronogram of divergence times from plastid DNA, ancestral area reconstructions, and geological events in the Indo-Burma ecoregion and parts of southern China. (Panel A) Map showing major floristic divisions A–C in the Indo-Burma ecoregion and parts of southern China according to the geographical divisions of Wen (2017), and the related phylogenetic reconstruction in our study: (A) the Hengduan Mountains and the Yunnan-Guizhou Plateau; (B) the Shan Plateau region; (C) the southern Truong Son Mountain Range. (Panel B) Divergence time estimates for all *Engelhardia* species. (Panel C) Ancestral area reconstructions of the *Engelhardia spicata* complex. (Panel D) Bar plot showing the frequency and composition of outward and inward transitions. Colors correspond to different source or sink regions, consistent with panel C.

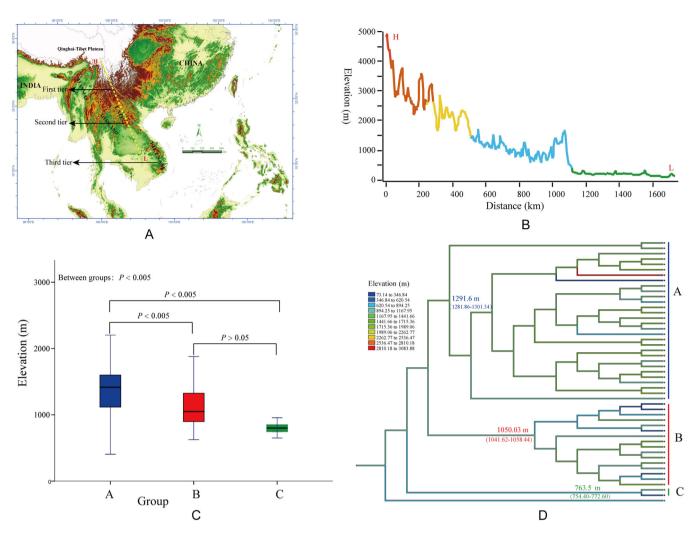


FIGURE 4 The three-tiered altitudinal landform structures of the Indo-Burma ecoregion and elevation differentiation of the *Engelhardia spicata* complex. (Panel A) Depiction of the ecoregion's three-tiered landform, as described in Figure 1. Dotted yellow lines indicate elevation. (Panel B) Modern topographic profile along the transect H–L shown in Panel A. (Panel C) Box and whisker plot indicating the current elevational range for each genetic group of the *Engelhardia spicata* complex. The differences between groups A and B and between groups A and C were statistically significant (P < 0.05); groups B and C did not differ significantly (P > 0.05) (Appendix S1: Table S10). (Panel D) Ancestral reconstruction of the mean and boundaries of the 95% elevational range using Mesquite 3.61 (Maddison and Maddison, 2019). The branch color represents elevation. Numbers above the branches are the mean and boundaries of the 95% elevational range (m a.s.l.) for ancestral nodes.

Our results indicate that the *E. spicata* complex diverged from its sister lineage, E. hainanensis around 32.16 Ma (95% HPD: 22-41.97; Figure 3B, Table 1), which aligns with previous studies that estimated this divergence at ~37.86 Ma (Meng et al., 2022b). This finding may reflect a lateral shift in the Indo-Burma ecoregion, potentially initiated by a significant collision estimated at 30 ± 5 Ma (Wang, 2017). Molecular studies of other lineages have produced similar results, such as the divergence of Roscoea (Zingiberaceae) from Cautleya (Zingiberaceae) at approximately 32 Ma (Zhao et al., 2016), and the isolation of the central Vietnam lineage of Stedocys (Araneae, Scytodidae) from the northern Vietnam and southwestern China lineages at approximately 30 Ma (Luo and Li, 2018). In addition, geological studies report that the left-lateral extrusion of Indochina began at approximately 35 Ma (Schärer et al., 1994; Leloup et al., 2001; Li et al., 2020) and was driven by significant

lithospheric delamination in the Ailao Shan-Red River shear zone (Feng et al., 2021). We propose that the lateral displacement of the Indo-Burma ecoregion occurred as early as the Early Oligocene, with a divergence time estimate of approximately 32.16 Ma. Our integrated results of molecular dating, ancestral range reconstruction, and biogeographic stochastic mapping (BSM) analyses (Figure 3) indicate that the ancestor of the *E. spicata* complex was distributed in the low-elevation region, with subsequent vicariance and dispersal events shaping its current distribution pattern.

The earliest lineages of the complex diverged in the lower-elevation southern region of the Truong Son Mountain Range, marking the initial stage of its diversification. The split between group C and its northern Asian sister group at 26.62 Ma coincides with major patterns of genetic structure within the *E. spicata* complex in the Truong Son Mountain

Range between northern and southern populations (Figure 3C, Table 1). This divergence may reflect far-field effects of the India-Eurasia collision. By the Late Oligocene, the continuous uplift of the QTP, along with the ongoing extension of the Ailao Shan-Red River shear zone (Che et al., 2010) shaped the modern Truong Son Mountain Range, whose complex topography has acted as a barrier to gene flow. The BSM analysis revealed group C as the primary source of outward dispersal (11.78 events), reflecting the ecological connectivity of the Truong Son Mountain Range during the Oligocene. However, subsequent tectonic uplift led to habitat fragmentation, restricting population connectivity. Similar patterns were observed for sympatric species such as Engelhardia roxburghiana and E. fenzlii (Meng et al., 2022b) and the spiny frog clade (Paini tribe; Che et al., 2010).

Molecular dating revealed the divergence of group A in the Hengduan Mountains and the Yunnan-Guizhou Plateau, and group B in the Shan Plateau, to be around 23.03 Ma (Figure 3C, Table 1). This timing is consistent with significant splits observed in alpine ginger lineages, which are estimated to have occurred approximately 23 Ma (Zhao et al., 2016). It also corresponds to the Oligocene-Miocene boundary (~23 Ma), as suggested by studies on spiny frogs (Che et al., 2010). This time point is generally considered to coincide with a major phase of the ongoing India-Asia collision, which caused the southeastward displacement of Indochina relative to South China along the Ailao Shan-Red River shear zone (Zhao et al., 2016). Geological evidence further supports this scenario, with peak rock metamorphism in the eastern Himalayan syntaxis 23 Ma (Zhang et al., 2012), suggesting that the Asian lithosphere underwent significant alteration around the Oligocene-Miocene boundary. The BSM analysis revealed distinct dispersal patterns among the major clades of E. spicata. Group A in the Hengduan Mountains and the Yunnan-Guizhou Plateau was a significant source of outward dispersal, with a total of 10.32 events recorded, reflecting its diverse habitats and historical connectivity. In contrast, group B in the Shan Plateau region was the largest sink for inward dispersals (10.86 events), likely due to its geographic connectivity.

Our findings highlight the interplay between geological events, ecological isolation, and dispersal dynamics in shaping the distribution and genetic structure of the *E. spicata* complex. The simultaneous uplift, strike-slip extrusion, and climatic fluctuations during the Oligocene likely created the conditions necessary to explain the observed patterns of vicariance and dispersal in the complex.

Spatial distribution of genetic diversity and the three-tiered landform pattern

The multimodal and ragged distribution generally indicate that the populations examined here are stable (Appendix S1: Figure S1). The wind-pollinated species has limited long-distance dispersal abilities (Hedrén and Lorenz, 2019;

Kotilínek et al., 2020; da Silva and Abrahão, 2021). This limitation, combined with the rapid uplift of the region, further supports the hypothesis that passive uplift is a primary driver of the elevation distribution of the species complex of *E. spicata*. Consequently, the likely stable populations and their distribution within the Indo-Burma ecoregion make the *E. spicata* complex a valuable model for studying geological evolution and terrain structure in this region.

The Engelhardia spicata complex exhibits a pronounced phylogeographic structure. Specifically, the initial divergence between the North Asian group, namely clades A and B and its sister group C, occurred at about 26.62 Ma; subsequently, the split between clades A and B took place around 23.03 Ma. The most recent common ancestor of group A is believed to have emerged in the Hengduan Mountains and Yunnan-Guizhou Plateau approximately 18.13 Ma. The estimated ancestral elevation of group A is approximately 1291.6 m (95% HPD: 1281.86-1301.34), closely corresponding to its current distribution altitude of about 1351.66 m (95% HPD: 1289.3-1414.03) (Figure 4; Appendix S1: Table S10). These findings suggest that the Hengduan Mountains and the Yunnan-Guizhou Plateau had already reached an altitude similar to present-day levels by the early Miocene, establishing the first tier of a threetiered landform structure. The most recent common ancestor of group B traces back to the Shan Plateau region around 15.5 Ma. The estimated ancestral elevation of group B is about 1050.03 m (95% HPD: 1041.62-1058.44), closely aligning with its current distribution altitude of about 1048.65 m (95% HPD: 880.75-1216.56) (Figure 4; Appendix S1: Table S10). The close alignment between the ancestral (1050.03 m) and current (1048.65 m) elevations indicates that the Shan Plateau achieved an altitude comparable to the present-day levels by the middle Miocene, thus forming the second tier of the landform structure. Furthermore, the most recent common ancestor of group C is thought to have appeared in the southern Truong Son Mountain Range around 10.78 Ma. The estimated ancestral elevation of group C is approximately 763.5 m (95% HPD: 754.4-772.6), which is close to its current distribution altitude of about 786.88 m (95% HPD: 650.52-923.25) (Figure 4; Appendix S1: Table S10). The congruence between ancestral (763.5 m) and contemporary (786.88 m) elevation estimates suggests that the southern Truong Son Mountain Range reached an altitude similar to its present-day levels by the late Miocene, establishing the third tier of the landform structure. Thus, our research supports that the three-tiered landform structure in mainland Southeast Asia had already formed by at least the late Miocene, coinciding with significant geological and climatic changes in the region. This preliminary finding supports the idea that the unique topography of the Indo-Burma ecoregion played a crucial role in shaping its rich biodiversity. However, these conclusions are based on our analysis of limited genetic markers and geological data from one species complex, and so further research is imperative to fully understand the timing and mechanisms

of the formation of the three-tiered landform structure. Future studies incorporating ecological data, such as historical climate records, habitat maps, and species interaction models, will be essential for unraveling the complex interplay between geological events, ecological dynamics, and the evolutionary history of the *E. spicata* complex and of other co-existing lineages.

Terrain refers to the undulating shape of the land resulting from the combined action of internal and external forces acting on Earth. This shape reflects the internal structure, tectonics, composition, and development of the crust at the surface (Zhang et al., 2014). Palaeo-altimetric studies generally agree that substantial surface uplift occurred in eastern Tibet at least 20 Ma before the onset of deep river incision (Ding et al., 2022). The southeastern margin of the QTP experienced significant crustal uplift during the Cenozoic era, with paleoelevation data from various periods indicating differences in the rates and magnitudes of uplift across different regions and time frames (van Hinsbergen et al., 2012; Ding et al., 2022; Lai et al., 2025a, b). Considerable evidence, including structural deformation, sedimentation, and thermal chronology, suggests that orogenic events during the Oligocene were widespread, extending beyond the southern margin of the plateau in the Himalayas and Gangdese Mountains to include the marginal areas of the plateau. These orogenic events resulted in the uplift of the Himalayas, Gangdese Mountains, Longmen Mountains, Tianshan, Ailao Mountains, and East Kunlun Mountains (Wang, 2017). Based on the analysis presented in the previous sentences and the findings of this study, it can be inferred that geological evolution since the Late Oligocene, along with tectonic movements and interactions among the Eurasian, Pacific, and Indian plates in the Indo-Burma ecoregion, were the primary drivers of the north-high-south-low terrain (Fanget et al., 2020; Garcés et al., 2020), and the three-tiered landform. These geological events likely triggered climate change, which in turn led to biotic reorganization across Asia (Meng and Song, 2023). This perspective aligns with the insights of Che et al. (2010), who attributed the emergence of elevated northern terrain and lower southern regions in the Himalayas and QTP to uplift processes that began in the late Oligocene. Similarly, Ding et al. (2022) proposed that the topographically induced flow of a weak lower crust played a crucial role in uplifting the eastern and southeastern QTP, contributing to a gradual decrease in elevation from the plateau's interior to Southeast Asia.

Our molecular dating results from a single species complex reveal, for the first time, the three-tiered landform structure of the Indo-Burma ecoregion based on botanical data. The first tier consists of the Hengduan Mountains and the Yunnan-Guizhou Plateau. The Hengduan Mountains are a series of north-south trending mountain ranges in western Sichuan and Yunnan provinces. This region encompasses several major ranges, including the Daxue Mountains, Min Mountains, and Jinsha River watershed, which originate from the southeastern margin of the QTP

(Wang et al., 2022). Elevations in this area range from 3000 to 6000 m a.s.l. with the highest peak being Kawagebo at 6740 m (Yang et al., 2022). The Hengduan Mountains are characterized by rugged terrain, steep slopes, and deep valleys dominated by alpine landscapes (Xiang et al., 2024; Chen et al., 2025). The climate is alpine with cold temperatures and heavy precipitation (Xiang et al., 2024). The Hengduan Mountains represent the first tier of the stepped topography in the Indo-Burma ecoregion, not only due to their high altitude but also because of the major rivers in this hotspot, such as the Mekong, Salween, Irrawaddy, and Red Rivers (Wen, 2017). The second tier of topography in mainland Southeast Asia is represented by the plateau region in eastern Myanmar, specifically the Shan Plateau. This area is characterized by extensive mountain ranges, high elevations, and complex terrain (Aung, 2012). The core of the third-tier terrain consists of a coastal river alluvial plain, primarily composed of three major plains and low hills interspersed among them. These include the Red River delta plain in the northeast (Larsen et al., 2017), the Ayeyarwady River downstream plain in the southwest (Chen et al., 2020), and the Mekong Delta in the south (Nguyen et al., 2000; Sakamoto et al., 2006). In a previous study, the three-tiered landform structure of mainland Southeast Asia was mainly explained from cultural and political perspectives (Wen, 2017). In contrast, our study clarifies the stepped terrain of the Indo-Burma ecoregion from a biological perspective. This research contributes to a better understanding of the topographical structure of the Indo-Burma ecoregion, its regional terrain patterns, and its evolutionary history, while positively impacting species conservation efforts.

CONCLUSIONS

Our comprehensive understanding of the evolutionary history of the E. spicata complex in the Indo-Burma ecoregion provides significant insights into the geological history and biogeographic structure of this region. Our data support the southeastward extrusion of Indochina along the Ailao Shan-Red River shear zone, which played a crucial role in the formation of the Truong Son Mountain Range during the Late Oligocene. Our research highlights a distinct altitudinal gradient across the Indo-Burma ecoregion, characterized by higher elevations in the north and lower areas in the south. This topographical gradient is reflected in the genetic diversity and population structure of the E. spicata complex, which corresponds to the tiered topography of the Indo-Burma ecoregion. Furthermore, the three-tiered landform in mainland Southeast Asia (tiered by altitude) was likely established by at least the Late Miocene, shaping the region's unique topography and influencing the evolution of its biodiversity. The tiered landscape begins with the Hengduan Mountains and the Yunnan-Guizhou Plateau as the first level, transitions to the Shan Plateau as the second level, and concludes with the coastal alluvial

plains as the third level. Our study represents a biologically informed delineation of tertiary tiered topography of the Indo-Burma ecoregion. This novel perspective offers valuable insights into the biogeographic patterns of Southeast Asia and underscores the importance of understanding the complex interplay between geological events and biological diversity.

AUTHOR CONTRIBUTIONS

C.Y.Z.: conceptualization, investigation, writing original draft, review, editing, visualization. G.L.C.: investigation, review, editing, visualization. J.L.H., P.H.H., M.L., R.P.S., O.Y.F., X.C., Y.G.S., G.X.H., K.Q.X.: review, editing. L.L. and S.S.Z.: investigation, resources. Y.H.T.: writing original draft, investigation, resources. H.H.M.: conceptualization, investigation, resources, writing original draft, review, editing, visualization, funding acquisition, and supervision. J.L.: conceptualization, supervision.

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DATA AVAILABILITY STATEMENT

All data required to replicate the results are included in the Supporting Information files. Haplotype sequences are deposited in GenBank as accessions PQ142373–PQ142622 and ribotype sequences as PQ154494–PQ154524. The original data sets are available via FigShare at https://doi.org/10.6084/m9.figshare.29069411.

ORCID

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SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

Appendix S1. Supplementary figures and tables.

Figure S1. Distribution of the number of pairwise nucleotide differences for ITS and plastid DNA of the *E. spicata* complex.

Figure S2. Phylogenies inferred using maximum likelihood (ML) and Bayesian inference (BI) based on the combined plastid DNA haplotypes.

Figure S3. Phylogenies inferred using maximum likelihood (ML) and Bayesian inference (BI) based on nuclear ITS data.

Figure S4. Bayesian divergence time estimates of the *Engelhardia* species based on the ITS data set.

Figure S5. Ancestral range reconstruction using BioGeo-BEARS and the DEC model.

Figure S6. Networks of the *E. spicata* complex from plastid DNA; each circle indicates a single haplotype drawn in proportion to its frequency.

Figure S7. Ancestral reconstruction utilizing the squared-change parsimony method (mapped on the plastid phylogeney), phylogenetic independent contrast (PIC) method and phylogenetic generalized least squares (PGLS) method analyses; the mean elevation and 95% confidence interval boundaries are noted.

Table S1. Details of *E. spicata* complex populations used in molecular analysis, sample size (N), and plastid DNA haplotypes (H) observed.

Table S2. Primers for the amplification of DNA regions in various populations of *E. spicata* complex.

Table S3. Comparison of four clock models in BEAST analyses via Bayes factors.

Table S4. Comparison of two tree prior processes in BEAST analyses via Bayes factors.

Table S5. Comparison of the fit of six models of biogeographical range evolution and model-specific estimates for different parameters.

Table S6. Summary of biogeographic stochastic map (BSM) from the DEC model.

Table S7. Distribution records used for species elevation analysis.

Table S8. Analysis of molecular variance (AMOVA) for plastid DNA data and ITS from three regions and all regions.

Table S9. Molecular diversity indices and neutral tests from three biogeographic regions and all biogeographic regions.

Table S10. Analysis of elevation variance (ANOVA) for the *E. spicata* complex across the three regions.

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