



Plant Signaling & Behavior

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/kpsb20

Functions of silicon and phytolith in higher plants

Rui Xu, Jianfeng Huang, Huijun Guo, Changming Wang & Hui Zhan

To cite this article: Rui Xu, Jianfeng Huang, Huijun Guo, Changming Wang & Hui Zhan (2023) Functions of silicon and phytolith in higher plants, Plant Signaling & Behavior, 18:1, 2198848, DOI: <u>10.1080/15592324.2023.2198848</u>

To link to this article: <u>https://doi.org/10.1080/15592324.2023.2198848</u>

© 2023 The Author(s). Published with license by Taylor & Francis Group, LLC.



0

Published online: 09 Apr 2023.

(Y)	٢	
ك	L	

Submit your article to this journal 🕑





View related articles 🗹



View Crossmark data

REVIEW

Taylor & Francis

OPEN ACCESS Check for updates

Functions of silicon and phytolith in higher plants

Rui Xu^{a,b#}, Jianfeng Huang^{d#}, Huijun Guo^c, Changming Wang^{a,b,e}, and Hui Zhan^{a,b,e}

^aKey Laboratory for Sympodial Bamboo Research, Southwest Forestry University, Kunming, China; ^bScience and Technology Innovation Team of National Forestry and Grassland Administration, Southwest Forestry University, Kunming, China; ^cYunnan Academy of Biodiversity/College of Biodiversity and Conservation, Southwest Forestry University, Kunming, China; ^dCAS Key Laboratory of Tropical Forest Ecology, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Mengla, China; ^eCollege of Forestry, Southwest Forestry University, Kunming, China;

ABSTRACT

Silicon (Si) is abundant in the lithosphere, and previous studies have confirmed that silicon plays an important role in plant growth. Higher plants absorb soluble silicon from soil through roots which is deposited in plant tissues mainly in the form of phytoliths. Based on previous studies, the research progress in silicon and phytoliths in the structural protection, enhancement on photosynthesis and transpiration of plants and plant growth and stress resistance was reviewed. Meanwhile, gaps in phytolith research, including phytolith morphology and function, impact of diverse environmental factors coupling with phytoliths, phytolith characteristics at different stages of plant development and phytoliths in regional vegetation are identified. The paper intends to promote the wider application of phytolith research findings and provides reference for further research on phytoliths.

ARTICLE HISTORY

Received 7 February 2023 Revised 1 March 2023 Accepted 4 March 2023

KEYWORDS

Silicon; phytolith; functions; higher plants

Introduction

Silicon is the second richest element in the earth's crust, and it is one of the key elements in the cycle of earth's materials¹. Silicon is absorbed by higher plant roots as soluble silica (Si $(OH)_4$) which was transported to different parts of the plant system through the vascular system. After a series of physiological and biochemical activities, it is silicified as phytoliths in various plant organs in specific forms depending on protocells and intercellular spaces^{2–4}. Phytoliths are ubiquitous in the leaves, branches, shoots, flowers, fruits, stems and roots of higher plants. They are the main form of silicon in higher plants, accounting for more than 90% of total silicon in plant^{5,6}.

Due to the unique advantages of phytoliths, viz., wide spread presence, high temperature and corrosion resistance, morphological stability and in situ sedimentation, the phytolith analysis is irreplaceable and widely used in botany, zoology, archeology, geology, paleoecology, pedology, medicine and agriculture. Silicon research has expanded as an Earth-life science superdiscipline⁷. Phytoliths have attracted increasing attention in recent years, mainly involving plant morphology classification⁸, plant community productivity⁹, wildlife feeding habits¹⁰, early evolution of plants and animals¹¹, topsoil phytolith analysis¹², stratum confirmation¹³, evidence of forest and grassland fire history¹⁴, reconstruction of ancient vegetation and environment¹⁵, propagation routes of cultivated plants¹⁶ and the origin of crops¹⁷. Based on previous research, this paper mainly summarized the positive effects of silicon and phytoliths on plant growth and physiological functions. In

addition, the research trends and issues of the relationship between phytoliths and plants were summarized, which can provide a valuable reference for future research on phytoliths.

Structural protection function of silicon and phytoliths

Phytoliths play a structural protective role in plants. The sessile and almost immobile characteristics of plants make them in a passive state when being vulnerable to insects, herbivores or pathogens. The whole life of plants is fraught with crises, which directly affect their survival, forcing them to exhibit various unique survival strategies to cope with the adverse environment and predators to achieve the purpose of gaining advantages and avoiding disadvantages^{18,19}. The expansion of grassland and the evolution of herbivores have also accelerated the evolution of phytoliths, and their co-evolution has formed a complex positive feedback network²⁰. Silicon and phytoliths play a multitude of significant roles in plant and global ecology²¹.

A large number of phytoliths produced significantly improve the mechanical stability of plants. Phytoliths exhibit better structural support performance than carbon-based compounds²² which contributed to develop erect and hard defensive plant structures such as stems, branches and leaves through complex biochemical pathways²³. Silicon deposition increases the number of phytoliths on the surface of stems, which significantly increases the cell wall thickness of stem sclerenchyma²⁴. In the leaves of Gramineae plants, a large

© 2023 The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The terms on which this article has been published allow the posting of the Accepted Manuscript in a repository by the author(s) or with their consent.

CONTACT Hui Zhan 🔯 zhanhui99@swfu.edu.cn; Changming Wang 🔯 forestwcm@swfu.edu.cn 🗈 Key Laboratory for Sympodial Bamboo Research, Southwest Forestry University, Kunming, China

[#]These authors contributed equally to this work.

This article has been corrected with minor changes. These changes do not impact the academic content of the article.

number of phytoliths are closely arranged, which better protect the vascular tissues of leaves and significantly enhance their resistance²⁵. The uptake and accumulation of silicon by plants also affects the properties of plant communities and ecosystems, and thus affecting ecological functions and response to environmental changes²⁶.

In addition, phytolith has a relatively high hardness, which is even higher than that of enamel. It can wear the mouthparts of insects and the teeth of herbivores and increase their feeding pressure on plants in terms of food cost and digestion difficulty^{10,27}. This effect of phytoliths in plants treated with higher silicon concentration is more obvious²⁸. Another anti-herbivory defensive mechanism of silicon is the use of sharp silica needles. The needlelike phytoliths enter the tissues of herbivores through the oral cavity and digestive system, bringing in microscopic pathogens (bacteria, fungi, viruses) and causing infections and cancer. Internal micro spines can wound large herbivores, insects and other small herbivores^{29–31}. Thus, silicon and phytoliths in plant tissue can act as a defense against vertebrate and invertebrate herbivores³².

When plants are damaged to a certain extent by insects or herbivores, a large number of phytoliths are produced to resist the damage³³. Structures such as micro hairs, bristles and hook hairs on the leaf epidermis of plants are frequently silicified. Hair density and hardness are used as physical defenses to directly "resist" herbivores, slow down or prevent the invasion of fungi and other pathogenic bacteria, and greatly enhance the ability of plants to resist erosion^{34,35}.

Generally, the deposition of phytoliths in vascular bundle and other conducting tissues and lignified cell wall in plants greatly enhance their mechanical strength and physical properties, decrease the mouthfeel of herbivores in feeding, thereby effectively reducing or avoiding the damage by herbivores and enhance the ability of plants to resist biotic and abiotic stresses in different ways.

Silicon and phytoliths enhance plant photosynthesis and transpiration

Photosynthesis and transpiration responsible for biomass production and overall growth and development are the most important physiological processes in plant. However, they are very sensitive to changes in environmental factors³⁶. Silicon and phytoliths sustain normal photosynthesis and transpiration of plants under changing environment³⁷. They regulate plant growth and physiological processes through photosynthesis and transpiration to alleviate the effects of stress on plants.

Exogenous silicon application increases the absorption and accumulation of silicon in plants, promotes the absorption of nutrient elements such as nitrogen, phosphorus, kalium, etc., thickens leaves and stems to enhance the average weight and significantly increases the chlorophyll concentration in leaves and the leaf arrangement, which has a positive effect on intercepting light energy, thereby improving the photosynthetic rate of rice, wheat, sugarcane, banana, cucumber, amaranth and other crops^{38–41}. Wang et al.⁴² found that the application of silicon fertilizer increases the volume of chloroplasts in mesophyll cells and increases the lamellar structure and grana of rice, which is conducive to the progress of photosynthetic

phosphorylation, reduces the concentration of intercellular CO₂, delays the decline in chlorophyll content in rice, and increases the photosynthetic rate and stomatal conductance. The research of Detmann et al.⁴³ supported the above viewpoint. They found that the application of exogenous silicon in the reproductive growth stage of rice increased the photosynthetic rate, attributing to nonstomatal limitation. The increase in silicon enhanced the leaf thickness, chloroplast surface area and mesophyll cell wall thickness, thus enhancing mesophyll conductance and promoting photosynthesis. Avila et al.44 found that silicon supplementation reduced the effects of water deficiency on leaf potential, instantaneous carboxylation efficiency and morphometry of the root system by increasing root growth, thus reducing the adverse effects of drought on photosynthesis. Silicon has been observed to ameliorate abiotic and biotic stresses in plants through gene regulation and interacting with other physiological processes to improve photosynthesis, such as absorption of macro and micronutrients, and phytohormones which also influence photosynthetic activity^{45,46}.

Silicon protects rice from overtranspiration under the stress conditions. Rice absorbs silicon and accumulates around the epidermis to form a special structure, which affects the permeability of plant tissues, reduces water loss from the epidermis. It precipitates in plant tissues and organs, responding to environmental stimulation in the form of stomatal movement to reduce transpiration and water evaporation on the plant surface⁴⁷⁻⁴⁹. Silicon deposition in the cell wall forms cuticle double layer in the epidermis of leaves; meanwhile, the stomatal conductance is reduced through the modification of the cell wall and reduces the transpiration of stratum corneum to improve the water status of plants under drought stress⁵⁰. The application of exogenous silicon also significantly reduces the transpiration rate of maize under drought stress, and with the extension of stress time, the transpiration rate gradually decreased⁵¹. The silicon absorbed by plants is deposited in cell walls, which prevents plant tissue cells from swelling and cracking due to excessive water absorption and stabilizes the osmotic pressure of plant tissue cells to a certain extent. Meanwhile, the phytoliths formed by silicon deposition in plants contain water, which increases or decreases in response to environmental changes, thus playing a passive role in storing and regulating the water in plants⁵².

Silicon promotes plant growth and fruit quality

The imperative role of silicon in triggering plant development has been identified. It is pivotal in regulating overall physiological and metabolic characteristics of the plants⁵³. The yield of rice is significantly reduced due to the lack of silicon during its growth^{54–56}. Supplementing silicon promotes the growth of rice roots, increases rooting sprouting (by 20%–30%), enhances the root vitality (by 1.12–1.13 times)^{57,58} and prolongs the functional period of roots to avoid premature senescence. In addition, silicon supplementation increases the number of mitochondria in cells, which is conducive to oxidative phosphorylation, thus increasing the respiration rate and the content of ATP (adenosine triphosphate) in roots, and improving the absorption of water and nutrients by roots⁵⁹.

Some studies also show that silicon promotes cell elongation, significantly increases the length and cell wall extensibility of leaf epidermal cells, and promotes the growth of rice seedlings. Silicon supplementation increases the number of phytoliths on the surface of plant stems, which thickens vascular bundles, and increases the cell wall thickness and leaf thickness of stem sclerenchyma^{60,61}. Silicon spraying on rice leaves activates and releases available soil nutrients, significantly increasing the total nitrogen content in rice leaves and the total phosphorus content in rice stems, sheaths, leaves and panicles, as well as the rice grain quality, e.g. the aspect ratio, brown rice rate and protein content⁶². Silicon enhances the nitrogen distribution in plants efficiently, reducing the nitrogen content in stems and leaves but increasing the nitrogen content in panicles, so as to promote the formation of fruits and seeds⁶³. Through complex physiological and biochemical processes, silicon increases rice yield by increasing its height, root size, vascular bundle number in stems, leaf weight and nutrient content⁶⁴⁻⁶⁶. Silicon also affects the growth of wheat. Silicon promotes the germination of wheat, improves the activities of various enzymes in seed, and promotes the transformation of nutrients and development of wheat embryos⁶⁷. In the growth stage of wheat seedlings, silicon application increases the root length, plant weight and chlorophyll content, enhances plant growth and development and increases the spike length, spikelet number and grain number to different degrees, thus increasing the wheat grain yield⁶⁸. It also improves the tensile properties of wheat dough evaluated from dry and wet gluten content, flour water absorption, dough breaking time and other processing quality indicators⁶⁹. Silicon application not only increases the photosynthetic pigment, biomass and growth of wheat but also improves the grain quality and yield⁷⁰. Moreover, silicon application plays a positive role in the growth of wheat in all types of soil⁷¹.

In addition, Suriyaprabha et al.⁷² found that the silicon absorbed by maize roots was carried and accumulated in leaves and other parts of maize, which increased the total protein content in the maize leaves and significantly enhanced the absorption of trace elements (copper, iron, manganese, zinc, etc.), so as to improve the growth potential of maize. Pei et al.⁷³ found that foliar application of nanosilicon fertilizer promoted the absorption and utilization of nitrogen, phosphorus, potassium and other nutrient elements in amaranth, increased the chlorophyll content in leaves, and enhanced photosynthesis, thus promoting the growth of amaranth; The fresh weight, dry weight, biomass and soluble sugar content of amaranth were obviously increased, thus increasing the yield of amaranth (by 11%–31%). Zhang⁷⁴ found that silicon fertilizer application significantly increased the plant height and yield of tomato (Solanum lycopersicum), increased the soluble sugar and vitamin C content, but decreased the organic acid content in tomato fruit. Fitiyani and Haryanti⁷⁵ also reported that the growth potential of tomato seedlings treated with nanosilicon fertilizer was obviously improved, and the plant height, number of new leaves, root length and other indicators were significantly higher than those of the control. Gong et al.⁷⁶ applied silicon fertilizer to Achnatherum extremiorientale seedlings and found that the root length, plant height, fresh weight and relative water content of the seedlings were significantly increased. Alsaeedia et al.⁷⁷ found that silicon promoted the absorption of nutrient elements such as nitrogen and potassium by cucumber plants and regulated the cell ion and osmotic balance, thus not only retarding the damage caused by salt and alkali on cucumber seedlings but also promoting growth and increasing cucumber yield.

Silicon and phytoliths improve the abiotic and biotic stress resistance of plants

Plants may survive through various stress factors, including biological stresses viz., pests and diseases caused by both fungi and bacteria in different plant species, and abiotic stresses viz., temperature stress, water stress, light stress, metal toxicity, and soil salinization, etc. Previous studies have shown the beneficial effects of silicon for plant growth, particularly under stress conditions⁷⁸. Silicon can alleviate the adverse impact of different abiotic and biotic stresses by different mechanisms including morphological, physiological, biochemical and genetic changes.

Silicon enhances the resistance to biotic stresses. Previous studies have shown that the improved mechanical strengthening of the plant as a result of silicon fertilization, thus directly improving plant constitutive defense ability against various folivores, borers, phloem and xylem feeder pests⁷⁹, including stem borer, brown planthopper, rice green leafhopper, and white-backed planthopper⁸⁰, greenbug Schizaphis graminum (Rondani)⁸¹, Thrips palmi Karny⁸², Myzus persicae⁸³, Scirpophaga incertulas (Walker)⁸⁴ and Asian citrus psyllid⁸⁵, etc. Silicon is as effective as conventional fungicides in resisting diseases. Under the condition of increasing silicon supply, the occurrence of stem rot, sheath brown rot, cucumber Fusarium wilt, crown spot disease, grain discoloration, gray leaf spot, dollar spot and brown patch besides rice blast and powdery mildew can be reduced⁸⁶. Silicon is as effective as conventional fungicides in resisting diseases. Under the condition of increasing silicon supply, besides rice blast and powdery mildew, it can also reduce the occurrence of stem rot, leaf sheath brown rot, cucumber wilt, crown spot, grain discoloration, gray leaf spot, dollar spot and brown spot. The reduction in disease symptom expression is due to the effect of silicon on some components of host resistance, including incubation period, lesion size, and lesion number⁸⁷. Silicon has also been proved to be mediated by signaling pathways during plant-pathogen interactions, viz., ET (ethylene), JA (jasmonic acid), SA (salicylic acid) and/or ROS (reactive oxygen species) (Ghareeb et al., 2011). Induced biochemical/molecular resistance stimulates the production of antibacterial compounds, regulates the complex signal pathway network, and activates the expression of defense-related genes to improve plant resistance⁸⁸. The temperature regulation of silicon mainly acts on transpiration. It regulates the water use efficiency and thereby improving the drought resistance and high-temperature resistance of plants. Meanwhile, under high-temperature stress, silicon increases the diameter of pollen grains of rice and makes the anthers of crack normally, thus ensuring the fertilization rate and seed setting rate of rice flowers⁸⁹. In the absence of silicon, the probability of rice suffering from rice blast is greatly increased ⁹⁰. Silicon fertilizer application to cucumber effectively inhibits

the probability of plant suffering from powdery mildew²⁴. Silicon also significantly improves the resistance of plants to drought. Silicon application reduces the MDA (malondialdehyde) content and relative electric permeability of rice roots under water stress, thus increasing the stability of the plasma membrane of plant roots. Silicon application also inhibits the production of peroxide, enhances the antioxidant capacity of plant roots, and alleviates the degradation of ABA (abscisic acid) in plant root cells. Under the water stress, silicon weakens the influence on the physiological activity of plant roots and maintains certain activity of roots^{51,91}. Silicon in sugarcane can reduce both freezing injury and water stress⁹². In addition, salt stress is the major threat for plant growth worldwide, and many studies have explained the regulation mechanism of silicon to alleviate salt stress from physiology, molecular genetics and genomic approaches⁹³. Silicon inhibits the transport of soil salt to the aboveground parts, thus enhancing the salt tolerance of plants to a certain extent⁹⁴.

The relationship between silicon and other elements is synergism and antagonism. On the one hand, silicon promotes the absorption and utilization of the vast majority of nutrients or beneficial elements by plants^{39,95,96}. On the other hand, silicon has an antagonistic effect on toxic and harmful elements and heavy metals to inhibit or reduce their harm to plants^{97–99}, and to some extent resists the adverse effects of radioactive elements (strontium, cesium, cadmium, etc.) on plant growth^{100,101}. There is an interaction between silicon and phosphorus. Silicon promotes the absorption of phosphorus by plants, and silicate promotes the release of phosphorus in soil, thus improving the bioavailability of phosphorus¹⁰². The relationship between silicon and calcium is very close but complicated. Different plant species, soil components, ecological environments, etc., affect the content of silicon and calcium with significant variation¹⁰³. Increasing the silicon absorption in plants reduces the stress of excessive iron and manganese. Silicon oxidizes iron and manganese ions in the roots of plants and turns them into hard-to-absorb deposits adhering to the surface of plant roots^{104,105}. Silicon reduces the bioavailability of aluminum by adjusting the pH and absorbing silicic acid, thus reducing the toxicity of aluminum¹⁰⁶. Silicon also precipitates heavy metals in plants, and improves the hardness of plants. Silicon promotes the precipitation of heavy metal ions in plants or precipitates in the form of silicate complexes, so as to reduce the concentration and mobility of active heavy metal ions in plant tissues. Moreover, silicon reacts with heavy metal ions in the outer cortex cells of plant roots to produce insoluble heavy metal salt complexes attached to the surface of roots, preventing the upward transportation of heavy metals in plants, thus weakening the damage of heavy metals to plants^{97,107}.

Summary and prospect

The silicon and phytoliths play a positive role in plant. They protect and support plant development by enhancing mechanical structure and their ability to resist herbivores and pathogens, adjusting the intensity of photosynthesis and transpiration and promoting the utilization of beneficial elements in soil by plants. They also inhibit the toxicity of heavy metals and alleviate the stress of soil salinization. Silicon reduces the damage to plants caused by water, temperature, light, air pollution and radiation and secures the transportation of water and nutrients, achieving physiological balance in plants from various aspects. Silicon enhances the ability of plants to resist biotic and abiotic stresses from different ways, thus promoting the growth and development. As an independent branch of micropaleontology, phytoliths provide a new perspective for extensive phytolith research. Gaps in the research on the relationship between silicon, phytoliths and plants remain to be filled:

- (1) Different plants produce different morphotypes of phytoliths, and the phytoliths in different organs of the same plant species also differ. The morphotypes of phytoliths reflect the structure and function of plants to some extent. The development mechanism of phytoliths is worth exploring, viz., morphology, structure, and function of phytoliths in different life stages of plants.
- (2) The responses of plants to different environmental factors differ. More attention needs to be addressed to the research of phytoliths in live plants and the variation of phytoliths type, size and yield from the perspective of single or complex environmental factors such as temperature, light, water and soil, etc.
- (3) The demand for silicon differs in various stages of plant development. Systematic research on the silicon demand for plants at different development stages may provide practical support for crop cultivation, intensive agricultural production and the development of silicon fertilizers.
- (4) Regional vegetation has different abilities to resist different biotic and abiotic stresses. Research on the physiological mechanism of silicon and phytoliths in plants, particularly the vegetation in ecologically fragile areas and its response to different stress may contribute to shaping the conservation strategies for vegetation restoration.

Acknowledgments

This work was jointly funded by Yunnan Provincial Department of Education Foundation (NO. 2022Y604), National Natural Science Foundation of China (NO. 32160415), Natural Science Foundation of Yunnan Province (NO. 202001AT070108) and Yunnan Provincial Joint Special Project for Basic Research in Agriculture (NO. 202101BD070001-114).

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

The work was supported by the National Natural Science Foundation of China [NO. 32160415]; Natural Science Foundation of Yunnan Province [NO. 202001AT070108]; Yunnan Provincial Joint Special Project for Basic Research in Agriculture [NO. 202101BD070001-114]; Yunnan Provincial Department of Education Foundation [NO. 2022Y604]

References

- Höhn A, Sommer M, Kaczorek D, Schalitz G, Breuer J. Silicon fractions in histosols and gleysols of a temperate grassland site. J Soil Sci Plant Nutr. 2008;171(3):409–418. doi:10.1002/jpln. 200625231.
- 2. Parr JF, Sullivan LA. Soil carbon sequestration in phytoliths. Soil Biol Biochem. 2005;37(1):117–124. doi:10.1016/j.soilbio. 2004.06.013.
- Prychid CJ, Rudall PJ, Gregory RM. Systematics and biology of silica bodies in monocotyledons. Botan Rev. 2003;69(4):377–440. doi:10.1663/0006-8101(2004)069[0377:SABOSB]2.0.CO;2.
- 4. Wang YJ, Lu HY. An introduction of plant opal and it's use. Adv Mar Biol. 1989;2:66–68.
- Twiss PC, Suess E, Smith RM. Morphological classification of grass phytolith. Soil Sci Soc America J. 1969;33(1):109–114. doi:10.2136/ sssaj1969.03615995003300010030x.
- Wang YJ, Lu HY. Phytolith study and it's application. Beijing: Ocean Press; 1993. p. 20–32.
- Nawaz MA, Zakharenko AM, Zemchenko IV, Haider MS, Ali MA, Imtiaz M, Chung G, Tsatsakis A, Sun S, Golokhvast KS. Phytolith formation in plants: from soil to cell. Plants (Basel). 2019;8(8):249. doi:10.3390/plants8080249.
- 8. Li ZM. Research on the potential of phytolith sequestration carbon from atmosphere in soil-plant ecosystem of typical wetland. Linan: Zhejiang Agriculture and Forestry University; 2013.
- 9. Yao L, Luo WH, Yang YZ, Zhang JZ, Li WY, Cao S, Mo LH. Study on plant microremains on the tooth surface of animals unearthed from Laosicheng site in Yongshun, Hunan. Agri Archaeol. 2015;6:1–7.
- Zhang X, Gélin U, Spicer RA, Wu F, Farnsworth A, Chen P, Del RC, Li S, Liu J, Jian H, et al. Rapid eocene diversification of spiny plants in subtropical woodlands of central Tibet. Nat Commun. 2022;13:1. doi:10.1038/s41467-022-31512-z.
- Fan B. The disaster event response to environmental changes recorded by phytolith in chaohu area. Shanghai: East China Normal University; 2006.
- Piperno DR. Phytolith taphonomy and distributions in archeological sediments from Panama. J Archaeol Sci. 1985;12(4):247–267. doi:10.1016/0305-4403(85)90032-9.
- Meng M, Jie DM, Gao GZ, Gao T, Xu SH, Lian YT, Xu HQ, Li TY, Wang JY, Niu HH, et al. Characteristics of burned phytolith from representative plants in Northeast China and implications for paleo-fire reconstruction. Rev Palaeobot Palynol. 2022;300:104628. doi:10.1016/j.revpalbo.2022.104628.
- Flenley JR, King A, Jackson J, Chew CW, Teller JT, Prentice ME. The late quaternary vegetational and climatic history of Easter Island. J Quat Sci. 1991;6(2):85–115. doi:10.1002/jqs.3390060202.
- Zou XJ, Li Q, Ma ZK, Jiang LP, Yang XY. Fan-shaped phytoliths reveal the process of rice domestication at Shangshan site, Zhejiang Province. Quat Sci. 2014;34:106–113.
- Zuo XX, Dai JQ, Wu W, Jin JH, Ge W, Wang YP, Ren L, Lin YJ, Pei YY, Xie H. Microfossil evidence of rice cultivation on the Southeast China coast 7500 years ago. Sci China Earth Sci. 2022;65(11):2115– 2126. doi:10.1007/s11430-022-9995-3.
- Wu Y. The application and improvement of phytolith analysis method. Hubei: China University of Science and Technology; 2008.
- Moles AT, Westoby M. Seed size and plant strategy across the whole life cycle. Oikos. 2006;113(1):91–105. doi:10.1111/j.0030-1299.2006.14194.x.
- Pierce SJ, Vianelli A, Cerabolini BE. From ancient genes to modern communities: the cellular stress response and the evolution of plant strategies. Funct Ecol. 2005;19(5):763–776. doi:10.1111/j.1365-2435.2005.01028.x.
- Vandevenne FI, Barão AL, Schoelynck J, Smis A, Ryken N, Damme PM, Meire P, Struyf E. Grazers: biocatalysts of terrestrial silica cycling. Proc Royal Soc London B Biolog Sci. 2013;280 (1772):20132083. doi:10.1098/rspb.2013.2083.
- Katz O. Silica phytoliths in angiosperms: phylogeny and early evolutionary history. New Phytol. 2015;208(3):642–646. doi:10. 1111/nph.13559.

- Ma JF, Yamaji N. Functions and transport of silicon in plants. Cellul Molecul Life Sci. 2008;65(19):3049–3057. doi:10.1007/ s00018-008-7580-x.
- Quigley KM, Anderson TM. Leaf silica concentration in Serengeti grasses increases with watering but not clipping: insights from a common garden study and literature review. Front Plant Sci. 2014;5:1–10. doi:10.3389/fpls.2014.00568.
- Miyake Y, Takahashi E. Effect of silicon on the growth of solutioncultured cucumber plant. Soil Sci Plant Nutr. 1983;29(1):71–83. doi:10.1080/00380768.1983.10432407.
- Reynolds OL, Keeping MG, Meyer JH. Silicon-augmented resistance of plants to herbivorous insects: a review. Ann Appl Biol. 2009;155(2):171–186. doi:10.1111/j.1744-7348.2009.00348.x.
- Katz O. Silicon content is a plant functional trait: implications in a changing world. Flora. 2019;254:88–94. doi:10.1016/j.flora. 2018.08.007.
- Strömberg CA, Stilio VS, Song Z, De Gabriel J. Functions of phytoliths in vascular plants: an evolutionary perspective. Funct Ecol. 2016;30(8):1286–1297. doi:10.1111/1365-2435.12692.
- Mir SH, Rashid I, Hussain B, Reshi ZA, Assad R, Sofi IA. Silicon supplementation of rescuegrass reduces herbivory by a grasshopper. Front Plant Sci. 2019;10. doi:10.3389/fpls.2019.00671.
- Hodson MJ, Smith RJ, van Blaaderen A, Crafton T, O'neill CH. Detecting plant silica fibres in animal tissue by confocal fluorescence microscopy. Ann Occup Hyg. 1994;38(2):149–160. doi:10. 1093/annhyg/38.2.149.
- Lev-Yadun S, Malka H. External and internal spines in plants insert pathogenic microorganisms into herbivore's tissues for defense. Micro Ecolog Res Trends. 2008;155–168.
- Lev-Yadun S, Malka H. Extended phenotype in action. Two possible roles for silica needles in plants: not just injuring herbivores but also inserting pathogens into their tissues. Plant Signal Behav. 2019;14(7):1–5. doi:10.1080/15592324.2019.1609858.
- 32. Massey FP, Sue EH. Physical defences wear you down: progressive and irreversible impacts of silica on insect herbivores. J Anim Ecol. 2009;78(1):281–291. doi:10.1111/j.1365-2656.2008.01472.x.
- Hartley SE, DeGabriel JL. The ecology of herbivore-induced silicon defenses in grasses. Funct Ecol. 2016;30(8):1311–1322. doi:10. 1111/1365-2435.12706.
- Callis-Duehl K, McAuslane H, Duehl A, Levey DJ. The effects of silica fertilizer as an anti-herbivore defense in cucumber. Horticult Res. 2017;25(1):89–98. doi:10.1515/johr-2017-0010.
- Vermeij GJ. Plants that lead: do some surface features direct enemy traffic on leaves and stems? Biol J Linnean Soc. 2015;116(2):288– 294. doi:10.1111/bij.12592.
- 36. Kalaji HM, Rastogi A, Živčák M, Brestic A, Daszkowska-Golec K, Sitko KY, Alsharafa R, Lotfi P, Samborska IA, Kusaka M. Prompt chlorophyll fluorescence as a tool for crop phenotyping: an example of barley landraces exposed to various abiotic stress factors. Photosynthetica. 2018;56(3):953–961. doi:10.1007/s11099-018-0766-z.
- Rastogi A, Yadav S, Hussain S, Kataria S, Hajihashemi S, Kumari P, Yang X, Brestic M. Does silicon really matter for the photosynthetic machinery in plants ... ? Plant Physiol Biochem. 2021;169:40–48. doi:10.1016/j.plaphy.2021.11.004.
- Deren CW, Datnoff LE, Snyder GH. Variable silicon content of rice cultivars grown on everglades histosols. J Plant Nutr. 1992;15 (1):2363–2368. doi:10.1080/01904169209364480.
- Huang HR, Xu L, Bokhtiar SM, Srivastava MK, Li YR, Yang LT. Effect of calcium silicate fertilizer on soil characteristics, sugarcane nutrients and its yield parameters. J Southern Agricult. 2011;42 (7):756–759. doi:10.3969/j.issn.2095-1191.2011.07.017.
- Ke YS, Huang JM, Xiao CM, Lun XJ, Shang ZF. Study on silicon and nitrogen-silicon linkage in paddy soil of Guangdong Province. Guangdong Agricul Sci. 1993;6:22–24. doi:10.1004/ 874x.1993.06.009.
- Pei FY, Dong CW, Chen WZ, Yang Y, Fang QF, Duan JX, Huang PZ, Wang HD. Preparation and the effect of nano-silicon fertilizer on the growth of Amaranth. Horticult Seed 2015;6:12–17. doi:10. 16530/j.cnki.cn21-1574/s.2015.06.005.

- 42. Wang X, Zhang GL, Huo ZY, Xiao YC, Xiong F, Zhang HC, Dai QG. Effects of application of nitrogen combined with silicon on the photosynthesis and activities of nitrogen metabolic enzyme of rice leaf. J Yangzhou Univ (Agricult Life Sci Edition). 2010;31(3):44–49. doi:10.16872/j.cnki.1671-4652.2010.03.010.
- 43. Detmann KC, Araujo WL, Martin SC, Sanglard LM, Reis JV, Detmann E, Rodrigues FA, Nunes-Nesi A, Fernie AR, DaMatta FM. Silicon nutrition increases grain yield, which, in turn, exerts a feedforward stimulation of photosynthetic rates via enhanced mesophyll conductance and alters primary metabolism in rice. New Phytol. 2012;196(3):752–762. doi:10.1111/j.1469-8137.2012.04299.x.
- 44. Avila RG, Magalhães PC, da Silva EM, Júnior CCG, de Paula Lana UG, de Alvarenga AA, de Souza TC. Silicon supplementation improves tolerance to water deficiency in sorghum plants by increasing root system growth and improving photosynthesis. Silicon. 2020;12(11):2545–2554. doi:10.1007/s12633-019-00349-5.
- 45. Hussain S, Shuxian L, Mumtaz M, Shafiq I, Iqbal N, Brestic M, Shoaib M, Sisi Q, Li W, Mei X, et al. Foliar application of silicon improves stem strength under low light stress by regulating lignin biosynthesis genes in soybean (Glycine max (L.) Merr.). J Hazard Mater. 2021;401:401. doi:10.1016/j.jhazmat.2020.123256.
- 46. Song A, Li P, Fan F, Li Z, Liang Y, Araujo WL. The effect of silicon on photosynthesis and expression of its relevant genes in rice (Oryza sativa L.) under high-zinc stress. PLoS One. 2014;9(11): e113782. doi:10.1371/journal.pone.0113782.
- Agarie S, Uchida H, Agata W, Kubota F, Kaufman PB. Effects of silicon on transpiration and leaf conductance in rice plants (*Oryza* sativa L.). Plant Prod Sci. 1998;1(2):89–95. doi:10.1626/pps.1.89.
- Ma JF, Takahashi E. Interaction between calcium and silicon in water-cultured rice plants. Plant Soil. 1993;148(1):107–113. doi:10. 1007/BF02185390.
- Yoshida S, Ohnishi Y, Kitagishi K. Histochemistry of silicon in rice plant. Soil Sci Plant Nutr. 1962;8(1):36–41. doi:10.1080/00380768. 1962.10430980.
- Zhu YX, Gong HJ. Beneficial effects of silicon on salt and drought tolerance in plants. Agron Sustain Develop. 2014;34(2):455–472. doi:10.1007/s13593-013-0194-1.
- Zou CQ, Gao XP, Zhang FS. Effects of silicon application on growth and transpiration rate of maize. Chinese J Eco- Agricult. 2007;15(3):55-51.
- Zhang GQ. Effect of silicon on the growth and some physiological characteristics of ginger. Taian: Shandong Agricultural University; 2008.
- Souri Z, Khanna K, Karimi N, Ahmad P. Silicon and plants: current knowledge and future prospects. J Plant Growth Regul. 2021;40 (3):906–925. doi:10.1007/s00344-020-10172-7.
- 54. Ma TS. Research status of silicon content and silicon fertilizer use in paddy soil in China. Advan Pedol. 1990;4:1–5.
- 55. Sun X. Soil nutrition-plant nutrition and rational fertilization. Agricult Publish House. 1983;15–36.
- 56. Zhou Q, Pan GQ, Shi ZJ. Effect of silicon fertilizer on population quality and yield of wheat. Jiangsu Agricult Sci. 2001;3:47–52.
- 57. Gong JL, Zhang HC, Long HY, Hu YQ, Dai QG, Huo ZY, Xu K, Wei HY, Gao H. Progress in research of nutrition functions and physiological mechanisms of silicon in rice. J Plant Physiol. 2012;48 (1):1–10. doi:10.13592/j.cnki.ppj.2012.01.001.
- Matichenkov V, Bochamikova E. Chapter 13 the relationship between silicon and soil physical and chemical properties. Studies Plant Sci. 2001;8:209–219. doi:10.1016/S0928-3420(01)80017-3.
- 59. Ke YS, Huang XH, Zhang ZT, Xiao CM, Wu LF, Li YZ, Jian HQ, Wu YQ. Effect of silicon fertilizer on nitrogen, phosphorus and potassium nutrition of rice and analysis of reasons for increasing production. Guangdong Agricult Sci. 1997;5:25–27. doi:10.16768/j. issn.1004-874x.1997.05.011.
- 60. Li BL. Research on phytolith carbon sequestration in bamboo and its relationship with species and lithology. Linan: Zhejiang Agriculture and Forestry University; 2014.
- Wang YM. Effects of silicon fertilizer on the growth and development, yield and quality of paddy rice. Chongqing: Southwest University; 2007.

- Ding WM. The effect of nano silica soil on nutrient absorption and quality of rice. Harbin: Northeast Agricultural University; 2015.
- 63. Gong Y. Effect of silicon on growth and development and absorption of nitrogen in apple trees. Master thesis, Shandong Agricultural University; 2012.
- Qu TG, Ding JM. Effect of silicon fertilizer on stress resistance and yield of direct seeding rice. Tillage Cultivat. 2003;2:57–58.
- Wang XY, Zhang YL, Zhang HM, Yu N. Research advance of silicon biochemical functions on crop. Soil Fertilizer Sci China. 2007;6:6–9.
- Wen CB, Gao HL, Cai DL, Chen CY. Present study situation of the silicon fertilizer application to rice. Areal Res Develop. 2003;3:79–81.
- Liu CB, Ma CC, Zhang DL. The effect of silicon on emerging metabolism of wheat. J Anhui Agricult Sci. 2001;29(4):502–503. doi:10.13989/j.cnki.0517-6611.2001.04.038.
- Mushtaq A, Jamil N, Riaz M, Hornyak L, Ahmed N, Rana SSA, Naeem M, Najam M, Malghani K. Synthesis of silica nanoparticles and their effect on priming of wheat (*Triticum aestivum* L.) under salinity stress. Biolog Forum. 2017;9:150–157.
- Yu LH, Gao JL. Effect of silicon yield and grain quality of wheat. J Triticeae Crops. 2012;32:469–473.
- Muhammad T, Waqar I, Hua Z. Promising role of silicon to enhance drought resistance in wheat. Commun Soil Sci Plant Anal. 2018;49(22):2932–2941. doi:10.1080/00103624.2018. 1547394.
- Bao SD, Yang XR, Li YQ, Zhang MJ. Silicon nutrition of wheat in calcareous soil and the effects of silicon, zinc and manganese fertilizers on wheat yield. Soils. 1996;06:311–315. doi:10.13758/j. cnki.tr.1996.06.008.
- Suriyaprabha R, Karunakaran G, Yuvakkumar R, Rajendran V, Kannan N. Silica nanoparticles for increased silica availability in maize (*Zea mays* L.) seeds under hydroponic conditions. Curr Nanosci. 2012;8(6):902–908. doi:10.2174/157341312803989033.
- Pei FY, Dong CW, Duan JX, Huang PZ, Chen ZQ, Wang DH. Effect of different silicon fertilizer on the growth, quality and yield of Amaranth. J Anhui Agricult Sci. 2015;43(12):76–78. doi:10. 13989/j.cnki.0517-6611.2015.12.028.
- 74. Zhang Z. Effects of different fertilization practice on plant growth and fruit quality of tomato and melon. Master thesis, Hebei Agricultural University; 2015.
- Fitiyani HP, Haryanti S. The effect of using nanosilica fertilizer on growth tomato plant (*Solanum lycopersicum* L.). Bullet Anatomy Physiol. 2016;24:34–41.
- Gong SF, Liu Y, Su XY, Jiang TT, Wang JG. Influence of nano silicon fertilizer on osmotic stress in achnatherum extremiorientale. Pratacult Sci. 2018;35(12):2924–2930. doi:10.11829/j.issn. 1001-0629.2018-0109.
- 77. Alsaeedia A, El-Ramadyb H, Alshaalb T, El-Garawani M, Elhawat N, Al-Otaibi A. Exogenous nanosilica improves germination and growth of cucumber by maintaining K+/Na+ ratio under elevated Na+ stress. Plant Physiol Biochem. 2018;125:164–171. doi:10.1016/j.plaphy.2018.02.006.
- Mandlik R, Thakral V, Raturi G, Shinde S, Nikolić M, Tripathi DK, Sonah H, Deshmukh R, Singh V. Significance of silicon uptake, transport, and deposition in plants. J Exp Bot. 2020;71(21):6703– 6718. doi:10.1093/jxb/eraa301.
- 79. Connick VJ. The impact of silicon fertilisation on the chemical ecology of grapevine, Vitis vinifera constitutive and induced chemical defenses against arthropod pests and their natural enemies [Ph.D. thesis]. Albury-Wodonga, NSW: Charles Sturt University; 2011.
- Ma JF. Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses. Soil Sci Plant Nutr. 2004;50(1):11–18. doi:10. 1080/00380768.2004.10408447.
- 81. Moraes JC, Goussai MM, Basagli MAB, Carvalho GA, Ecole CC, Sampaio MV. Silicon influence on the tritrophic interaction: wheat plants, the greenbug Schizaphis graminum (Rondani) (Hemiptera: Aphididae), and its natural enemies, Chrysoperla externa (Hagen) (Neuroptera: Chrysopidae) and Aphidius colemani Viereck

(Hymenoptera: Aphidiidae). Neotrop Entomol. 2004;33:619–624. doi:10.1590/S1519-566X2004000500012.

- 82. De Almeida GD, Pratissoli D, Zanuncio JC, Vicentini VB, Holtz AM, Serrão JE. Calcium silicate and organic mineral fertilizer applications reduce phytophagy by thrips palmi karny (Thysanoptera: thripidae) on eggplants (Solanum melongena L.). Interciencia. 2008;33:835–838.
- Ranger CM, Singh AP, Frantz JM, Cañas L, Locke JC, Reding ME, Vorsa N. Influence of silicon on resistance of zinnia elegans to myzus persicae (Hemiptera: aphididae). Environ Entomol. 2009;38:129–136. doi:10.1603/022.038.0116.
- Jeer M, Telugu UM, Voleti SR, Padmakumari AP. Soil application of silicon reduces yellow stem borer, scirpophaga incertulas (walker) damage in rice. J Appl Entomol. 2017;141(3):189–201. doi:10.1111/jen.12324.
- Ramírez GA, Puentes PG, Restrepo DH. An evaluation of the use of calcium, potassium and silicon for the management of diaphorina citri populations in Tahiti lime trees. Notulae Botanicae Horti Agrobotanici Cluj-Napoca. 2018;46(2):546–552. doi:10.15835/ nbha46211152.
- Elizabeth AG, Lawrence ED. Silicon in turfgrass: a review. Crop Sci. 2021;61(6):3861–3876. doi:10.1002/csc2.20591.
- Daniel D, Fabrício AR, Lawrence ED. Silicon's role in abiotic and biotic plant stresses. Annu Rev Phytopathol. 2017;55(1):85–107. doi:10.1146/annurev-phyto-080516-035312.
- Wang M, Gao LM, Dong SY, Sun YM, Shen QR, Guo SW. Role of silicon on plant-pathogen interactions. Front Plant Sci. 2017;8:1– 14. doi:10.3389/fpls.2017.00701.
- Cao BL, Xu K, Shi J, Xin GF, Liu CY, Li X. Effects of silicon on growth, photosynthesis and transpiration of tomato. J Plant Nutri Fert. 2013;19(2):354–360. doi:10.11674/zwyf.2013.0211.
- Bonman JM, Estrada BA, Bandong JM. Leaf and neck blast resistance in tropical lowland rice cultivars. Plant Dis. 1989;73(5):388– 390. doi:10.1094/pd-73-0388.
- Ming DF, Yuan HM, Wang YH, Wang YH, Gong HJ, Zhou WJ. Effects of silicon on the physiological and biochemical characteristics of roots of rice seedlings under water stress. Scientia Agricultura sinica. 2012;45(12):2510–2519. doi:10.3864/j.issn. 0578-1752.2012.12.021.
- Majumdar S, Prakash NB. An overview on the potential of silicon in promoting defense against biotic and abiotic stresses in sugarcane. J Soil Sci Plant Nutr. 2020;20(4):1969–1998. doi:10.1007/ s42729-020-00269-z.
- Zhu YX, Gong HJ, Yin JL. Role of silicon in mediating salt tolerance in plants: a review. Plants (Basel). 2019;8(6):147. doi:10.3390/ plants8060147.

- Zhang YL, Yang D, Liu MD, Li J. Combination effects of N, P and Si on rice yield in acid paddy soil of Eastern liaoning. Chinese J Soil Sci. 2003;05:432–435. doi:10.19336/j.cnki.trtb.2003.05.012.
- 95. Wu Y, Wei D, Gao SH. Study on the nutritional function and effective conditions of silicon for rice. Soil Fert. 1992;03:25–27.
- 96. Zhang XM, Zhang ZY, Duan KS, Li GL, Duan ZX, Liu YG. Study on the available silicon content and the relationship between it and the properties of physics and chemistry of soils. J Heilongjiang Bayi Agricult Univ. 1996;02:42–45.
- Chen CF, Zhong JH, Li SY. Effect of silicon on growth and antistress ability of Chinese cabbage (*Brassica pekinensis* Rupr.) in cadmium contaminated soil. Plant Physiol J. 2007;03:479–482. doi:10.13592/j.cnki.ppj.2007.03.030.
- 98. Wang YR, Chen P. The absorption and distribution of Se and the coeffects of Se-Si in rice. J Plant Physiol. 1996;22:344–348.
- 99. Wang YR, Cheng Y, Hu ZQ, Zhou JH, Zhang YS. A study on poisoning rice (*Oryza sativa* L.) seedling with Na and Cu salts through inhibition of Si nutrition. J Plant Physiol. 1997;36:72–75.
- 100. Li ZM, Song ZL, Li BL, Cai YB. Phytolith production in wetland plants of the hangzhou xixi wetlands ecosystem. J Zhejiang Agricult Forest Univ. 2013;30(4):470–476. doi:10.11833/j.issn. 2095-0756.2013.04.002.
- 101. Zhou JH, Wang YR. Physiological studies on poisoning effects of Cd and Cr on rice (*Oryza sativa* L.) seedlings through inhibition of Si nutrition. Chinese J Appl Environ Biol. 1999;01:12–16.
- Hu KW, Yan L, Guan LZ. Interaction of silicon and phosphorus in soils. Chinese J Soil Sci. 2004;2:230–233. doi:10.3321/j.issn:0564-3945.2004.02.030.
- Zang HL. Control effect of silicon fertilizer on rice blast. Plant Prot. 1985;4:30.
- 104. Hou YL, Guo W, Zhu YG. Effect of silicon on plant and relevant mechanism under abiotic stresses. Chinese J Soil Sci. 2005;3:426– 429. doi:10.19336/j.cnki.trtb.2005.03.033.
- 105. Li YY, Liu Y, Liu SQ, Shi XM, Zhang MY, Ji JH, Feng L, Wang LY, Zhao HM. Study on the effect of silicon fertilizer application on rice in Heilongjiang province. Heilongjiang Agricult Sci. 2009;03:60–63.
- 106. Xu FF, Du JP. Effects of silicon on alleviating toxicity to rice seedlings under aluminum stress. Hybrid Rice. 2013;28(06):73– 75. doi:10.16267/j.cnki.1005-3956.2013.06.002.
- 107. Liang YC, Chen Q, Liu Q, Zhang WH, Ding RX. Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of salt-stressed barley (*Hordeum vul*gare L.). J Plant Physiol. 2003;160(10):1157–1164. doi:10.1078/ 0176-1617-01065.