




Article

Dry-Season Fog Water Utilization by Epiphytes in a Subtropical Montane Cloud Forest of Southwest China

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Abstract: Fog water is generally considered to be an important water source for epiphytes in cloud forests because they cannot directly access ground-level water sources. However, the water use proportions of potential water sources and water use efficiency of epiphytes in the subtropical montane cloud forests (MCF) remain to be further explored. In this study, we investigated the water use pattern in the dry season and the intrinsic water use efficiency (WUE_i) of four epiphyte groups (i.e., epiphytic lichens, epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants) using stable isotope ($\delta^2\text{H}$, $\delta^{18}\text{O}$, and $\delta^{13}\text{C}$) techniques. Our results indicated that the water sources of epiphytes were significantly different among groups and species. The contribution proportions of fog water to epiphytic lichens, epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants were 83.2%, 32.7%, 38.8% and 63.7%, respectively. Epiphytic lichens and epiphytic seed plants mainly depended on fog water whereas the epiphytic bryophytes and epiphytic ferns relied on both fog water and humus. This may be due to their differences in morphological and structural traits (e.g., thallus or leaves, rhizoid or roots). Additionally, the difference in WUE_i was also significant among epiphyte groups and species, which could be related to their different water acquisition patterns. In conclusion, our study reveals the differentiation of water utilization in epiphytes and confirms the importance of fog water for epiphytes during the dry season.

Keywords: fog water; stable isotopes; water source; intrinsic water use efficiency; differentiation; drought



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1. Introduction

In the context of global climate change, the frequency, duration, and intensity of drought will increase in many regions of the world [1,2]. There is a widespread concern that such drought events will weaken the current forest carbon sink and threaten the biodiversity of various forest ecosystems [3]. Epiphytes, which grow on living or residual host trees, are one of the most diverse groups in the forest ecosystem. These plants play an important role in maintaining biodiversity and biomass and promoting the processes of nutrient and water cycling [4,5]. Epiphytes lack direct access to the ground soil and rely heavily on atmospheric resources (e.g., water, nutrients), which makes them exceptionally sensitive to changes in environmental conditions and good biological indicators of global climate change [6,7]. Understanding the water use strategies of different epiphytes is critical for correctly predicting the impacts of predicted changing rainfall patterns on them.

Montane cloud forest (MCF) is generally characterized by persistent, frequent incidences of fog and low cloud cover at the canopy level, which usually harbors an abundant epiphyte community [8–10]. The role of epiphytes in the water cycle of MCF has been studied in the Neo-tropical regions (e.g., Costa Rica, Guatemala, Mexico and Ecuador) [11–14]. These studies indicate that epiphytes contribute substantially to canopy dynamics by stabilizing canopy organic soils (i.e., humus) that affect the interception of atmospheric moisture (e.g., fog and rainfall). In the subtropical MCF of Southwest China, also known as the montane moist evergreen broadleaved forest, the diversity of epiphytes is extremely high (>600 species, including epiphytic lichens, epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants) [15,16]. However, a six-month-long dry season lasts from November to the subsequent April, implying that the epiphytes need to overcome seasonal drought stress [7,17]. Moreover, the epiphytic habitats are usually harsh and heterogeneous with significant variations in water availability at small spatiotemporal scales, and short-term drought occurs even in rainy seasons [17,18]. Mechanisms on adaptation strategies of epiphytes to forest canopies with limited and unstable water availability remain one of the most captivating questions in plant ecology [19,20].

Multiple morphological and structural traits allow epiphytes to cope with the water deficits, such as early dormant, water storage stems and leaves, rhizomes, and air moisture absorption [21–23]. Recent studies confirm that epiphytes could take up atmospheric water (e.g., water vapor, rainwater, and fog water) directly throughout their surface structures, especially for epiphytic lichens that lack roots and an outer waxy cuticle [24,25]. Atmospheric water enhances the growth of epiphytes by alleviating the desiccation degrees and lengthening their photosynthetic activities [26]. Among them, Liu et al. [13] indicated that the *Ficus tinctoria* in the epiphytic stage would utilize a combination of fog water and rainwater present in canopy humus. Similarly, Wu et al. [27] found that fog water is a crucial supplemental source (contributing ratio ~87%) to maintain the water budgets of epiphytes during the dry season. To adapt to the water deficits, epiphytes was suggested to exhibit higher intrinsic water use efficiency (WUE_i) than the ground-rooted plants [28]. However, far too little attention has been paid to the quantitative relationship between the potential water sources and the water WUE_i among different groups of epiphytes.

Epiphytes are usually divided into different groups (namely epiphytic lichens, epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants) based on their systematic positions, life cycles, and biological characteristics. Epiphytic lichens are complex life forms that live in a symbiotic relationship with algae and fungi, which can attach to tree trunks only with the help of thalli base plates [29,30]. Epiphytic bryophytes, with no true roots (only rhizoid) and vascular systems (only rib), usually grow on barks or thick mats of humus [31–33]. As vascular plants, however, both epiphytic ferns and epiphytic seed plants have developed true roots and networks of the vascular system, allowing efficient water uptake, water and nutrient transport, and photosynthates [34,35], although roots and vascular systems in the former are much simpler than the latter. Due to these differences in structure, water uptake and transport, different epiphytic groups are expected to show differences in their water sources and water use efficiencies, which has not been well-quantified. Early findings suggested that the WUE_i of epiphytic bryophytes were lower than the vascular plants [36]. Although the water use of several epiphyte species has been studied, investigating the water use characteristics of different epiphyte groups remains scarce and challenging.

With the potential aggravation of drought predicted by future climate models [37], epiphytes will face more severe water stress in the dry season due to their high sensitivity to water fluctuations [7,38]. Understanding how different epiphytes obtain water and maintain normal life activities in the dry season is urgently needed to predict their potentially different responses to changes in the rainfall pattern. To fill the knowledge gap mentioned above, 12 dominant species belonging to four epiphyte groups were selected from a subtropical MCF in SW China to study their water use at the peak of the dry season. The specific objectives of this study are to quantify the water sources of different

epiphyte groups in the dry season based on stable isotope techniques ($\delta^2\text{H}$ and $\delta^{18}\text{O}$), and explore the intrinsic water use efficiency (WUE_i) of various epiphyte groups using $\delta^{13}\text{C}$. We expected that the water use patterns of epiphytes show group-specific and interspecific differentiations, and fog water plays a crucial role in maintaining the water absorption of epiphytes during the dry season.

2. Materials and Methods

2.1. Study Site

The study was conducted at the Ailaoshan Station for Subtropical Forest Ecosystem Studies (ASSFE, $23^\circ 35' - 24^\circ 44' \text{ N}$, $100^\circ 54' - 101^\circ 01' \text{ E}$), located at the Ailao Mountain, Jingdong County, Yunnan Province, SW China [39]. Co-affected by the western monsoon from India-Pakistan and the southeast monsoon, the seasonal distribution of precipitation is uneven, with only approximately 16% of the precipitation occurring in the dry season (November to April) (Figure 1a). The mean annual relative humidity (RH) was 84%, and the mean air temperature was 11.6°C [9]. However, fogs are frequent in this research region (212 days per year). The foggy days (Visibility $< 1000 \text{ m}$, Rainfall = 0 mm) in the dry season are 12 days per month on average [40]. Meanwhile, the average annual evaporation and annual net radiation are 1126 mm and 2444 W/m^2 , respectively (Figure 1b).

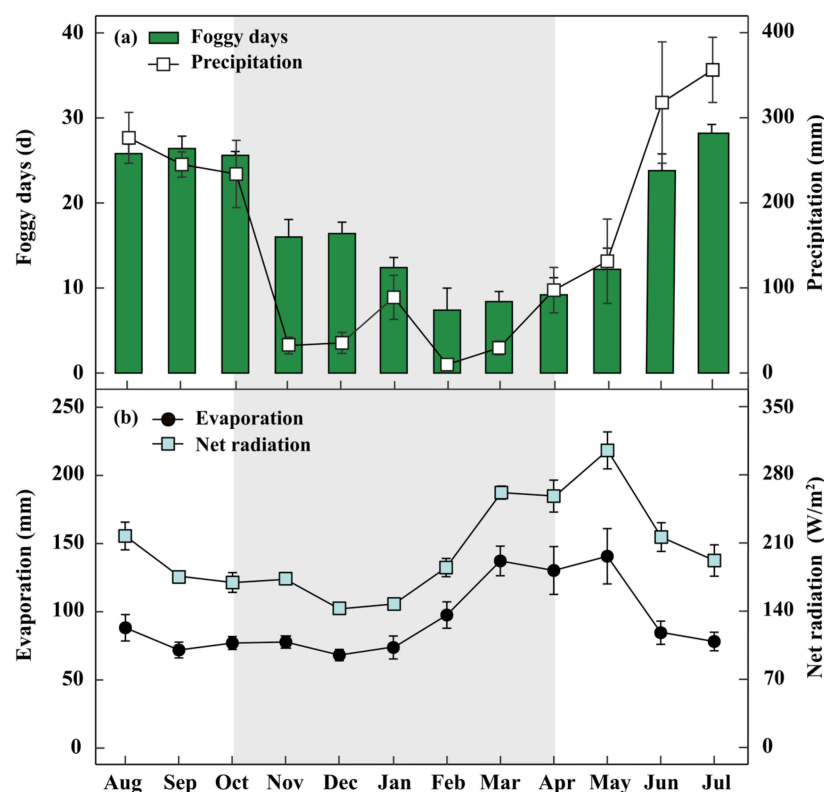


Figure 1. Meteorological conditions of the study site. Average monthly foggy days and precipitation (during 2015–2019; (a)), as well as average monthly number of evaporation and net radiation (b) at the study site. The dry seasons from November to April are remarked by grey bars. Each vertical bar represents an average ($\pm \text{SE}$) for that month.

These meteorological data were obtained from the Ailaoshan weather station, which is located near the study site. All data were collected using a suite of micro-meteorological sensors, including air temperature (HMP45, Vaisala Inc., Helsinki, Finland), radiation (CNR-1, Kipp and Zonen Inc., Delft, The Netherlands), and rainfall amount (52203, RM Young Inc., Traverse City, MI, USA). An evaporation pan (E-601, Weinasa Inc., Sichuan, China) with 0.62 m in diameter and 0.69 m in height was used to monitor daily evaporation. The water depth in E-601 was 0.6 m and the surface of the water is at ground level. Further

details about the site are provided in Song et al. [9] and Zhang, et al. [41]. This region has abundant epiphytic communities, including more than 217 species of epiphytic lichens [42], 176 species of bryophytes [16], 93 species of ferns, and 125 species of seed plants [43,44].

2.2. Isotopic Sampling

Hydrogen ($\delta^2\text{H}$) and oxygen ($\delta^{18}\text{O}$) stable isotopes are widely used to trace the water sources of plants, which rest on the fact that the isotopic composition of plant non-photosynthetic tissues remains unchanged during the root water uptake process and long-distance water transport, until it reaches leaves or non-suberized stems [45,46]. Moreover, the stable isotope of carbon ($\delta^{13}\text{C}$) is an effective approach to reflect the intrinsic water use efficiency (WUE_i) of epiphytes [47]. This method is based on the positive correlation between bulk leaf $\delta^{13}\text{C}$ of plants and their WUE_i [48].

Because the epiphytes can absorb atmospheric water inputs and humus simultaneously through leaves, roots, and rhizoid (except epiphytic lichens) [13,27], we assumed that sampling water of epiphytes was a combination of rainwater, canopy humus, and intercepted fog. Epiphytes may be subjected to long-term drought stress during the rainless dry season due to their high dependences on atmospheric water inputs. Also, different epiphytic groups may show differences in responding to the seasonal water deficits. Therefore, we studied four groups of epiphytes at the peak of the dry season (from 10 to 23 January 2019) to infer their potential responses to changes in the rainfall pattern. Species used included two epiphytic lichens (*Nephromopsis pallescens*, *Lobaria retigera*), four epiphytic bryophytes (*Homaliodendron montagneanum*, *Plagiochila assamica*, *Bazzania himalayana*, *Thuidium cymbifolium*), four epiphytic ferns (*Asplenium indicum*, *Lepisorus loriformis*, *Hymenophyllum polyanthos*, *Loxogramme chinensi*), and two epiphytic seed plants (*Aeschynanthus buxifolius*, *Agapetes mannii*). All the species are C_3 plants, and thus we can use carbon isotopic ratio $\delta^{13}\text{C}$ to compare their WUE_i [35].

For different groups of epiphytes, the samples of epiphytic seed plants ($n = 4$) and epiphytic ferns ($n = 4$) were collected from the non-photosynthetic basal culm tissue, while the samples of epiphytic bryophytes ($n = 4$) and epiphytic lichens ($n = 4$) were collected from shoots and thalli separately. All samples of the epiphyte species were collected from different host trees in the morning (9:00–11:00) from 10 to 23 January, 2019. Considering that the epiphytes grow at different heights of the host tree, 5–10 non-photosynthetic tissues, shoots, or thalli of epiphytes were combined into a single sample and four replicates for each species were collected from the trunk (<3 m) of each host tree. If the numbers of the individuals on the target host trees were insufficient, the epiphyte samples were collected from the neighboring 3–5 trees (within the distance of 5 m of the host tree). After each sampling, liquid water, humus or litter on the surface of the epiphyte samples were gently cleaned with a filter paper one by one carefully. Then the samples (~ 10 g per sample) were retained and instantly put into 10 mL screw-cap glass vials, sealed with parafilm, frozen (-4°C) in the portable fridge. After getting back from the field, all samples were moved to the refrigerator instantly until water extraction using cryogenic vacuum distillation [45]. While collecting the plant samples, the humus samples ($n = 4$) were collected at the sampling site of epiphyte species, except for epiphytic lichens that grow directly on the bark of the host trees. The collection process of humus samples was the same as epiphyte samples.

Fog water and rainwater were collected at the Ailaoshan weather station throughout the period of the experiment for water source determination and throughout 2018 and 2019 for the analysis of the local meteoric water line (LMWL). Fog water was collected using a self-made V-shaped collector (Figure S1a). The collector was set up at 1 m height and a circular rain shield (0.6 m in diameter) was mounted on the top of the fog collector to prevent vertical precipitation (i.e., rainwater) from entering the collector [49]. Fog water samples were collected during a dense fog event between 8:00 to 9:00 before the isotope fractionation happens from re-evaporation [50,51]. Meanwhile, the rainwater was collected using a cylindrical collector measuring 0.65 m in height and 0.2 m in diameter (Figure S1b),

with a 15 cm diameter funnel-shaped draining to a 1 L polyethylene bottle. All rainwater samples were collected immediately after a rainfall event or early in the morning following overnight rainfall. During the experiment period, the fog water and rainwater samples were collected daily on days with fog and rain events (a total of seven fog events and five rainfall events occurred) in the study site. All the fog and rain samples were stored immediately in 2 mL screw-cap glass vials, sealed with parafilm, and frozen (-4°C) in the refrigerator until water extraction using cryogenic vacuum distillation. In addition, samples for all rainfall events between March 2018 to March 2019 were collected after the precipitation events (8:00) to analyze the LMWL.

For each repetition of epiphyte species for $\delta^{13}\text{C}$ analysis, all healthy and fully expanded mature leaves (10–20) were collected from a host tree on the dates of plant tissue collection. The selected epiphyte samples were collected on the same day of sample collection for water source analysis on the same day. The humus or litter on the surface of the leaves were gently cleaned superficially with a filter paper, oven-dried at 70°C for 48 h to a constant mass, homogenized and ground to fine powder to pass through a 100-mesh sieve and stored immediately in 2 mL screw-cap glass vials, sealed with parafilm, then subsampled until $\delta^{13}\text{C}$ analysis in the laboratory [52].

2.3. Isotope Measurements

The liquid water from plant samples and humus (0.5–1 mL per sample) was extracted using the ultra-low temperature (-196°C) automatic vacuum condensation and extraction system (LI-2100, Lica United Technology Limited Inc., Beijing, China). All samples were filtered using an injection syringe fitted with a filter (pore size $0.22\ \mu\text{m}$) during this process. Then, the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of liquid water (including plant tissues water, humus, fog water and rainwater) were determined using the DELTA-V-Advantage isotope ratio mass spectrometer (Thermo Fisher Scientific, Bremen, Germany) combined with a high-temperature conversion elemental analyzer. To avoid any “memory effect”, each sample was analyzed four times with the last three injections used for calculations [53]. The determination of $\delta^{13}\text{C}$ in leaf samples was carried out using a flash combustion elemental analyzer (Flash EA) coupled with the DELTA-V-Advantage isotope ratio mass spectrometer (Thermo Fisher Scientific, Bremen, Germany) in the Thermo Fisher Scientific, Shanghai, China. The precision of the $\delta^2\text{H}$, $\delta^{18}\text{O}$, and $\delta^{13}\text{C}$ measurements were 1‰ , $\pm 0.2\text{‰}$, and $\pm 0.15\text{‰}$, respectively.

Isotope composition ($\delta^2\text{H}$, $\delta^{18}\text{O}$, and $\delta^{13}\text{C}$) of unknown samples were addressed to the Vienna-Standard Mean Ocean Water (V-SMOW) and the Vienna-standard Pee Dee Belemnite (V-PDB):

$$\delta^2\text{H}\left(^{18}\text{O}\right)_{\text{sample}} = (R_{\text{sample}}/R_{\text{VSMOW}} - 1) \times 1000 \quad (1)$$

$$\delta^{13}\text{C}_{\text{leaf}} = (R_{\text{leaf}}/R_{\text{PDB}} - 1) \times 1000 \quad (2)$$

where R_{sample} is the isotope ratio ($^2\text{H}/^1\text{H}$, $^{18}\text{O}/^{16}\text{O}$) of a water sample, R_{VSMOW} is the isotope ratio ($^2\text{H}/^1\text{H}$, $^{18}\text{O}/^{16}\text{O}$) of the Vienna Standard Mean Ocean Water (VSMOW) standard, R_{leaf} is the isotope ratio ($^{13}\text{C}/^{12}\text{C}$) of plant bulk leaf, R_{PDB} is the isotope ratio ($^{13}\text{C}/^{12}\text{C}$) of the Vienna Pee Dee Belemnite (VPDB) standard.

2.4. Statistical Analysis

The MixSIAR model was used to evaluate the relative contribution of potential water sources to the water used by plants [54]. This Bayesian mixing model combines a variety of sources of uncertainty including multiple sources, spatiotemporal variability, and isotope fractionation [55,56]. Regardless of whether multiple sources are logically related to each other, or whether the isotopic signatures were significantly different, the model can accurately calculate the contribution range of each source to the mixture by priori or posteriori aggregation methods [57]. In this study, input data of the MixSIAR model were the individual isotope values ($\delta^2\text{H}$ and $\delta^{18}\text{O}$) of epiphytes (i.e., non-photosynthetic

tissues, shoots, or thalli of epiphytes in this study, $n = 4$) and the average and standard deviation (SD) of potential water sources (i.e., fog water: $n = 7$; humus: $n = 4$; rainwater: $n = 5$). The discrimination data were set to zero for both $\delta^2\text{H}$ and $\delta^{18}\text{O}$ because isotopic fractionation does not occur during plant water uptake process [45]. The humus samples were collected one-to-one correspondence to the epiphyte samples during the sampling process. In consequence of a lack of humus accumulation in their habitats, the humus was excluded from the potential water sources for the epiphytic lichens.

All statistical analysis was performed using the statistical platform R3.6.3 [58]. After checking the normality and the homogeneity of variances using the Shapiro-Wilk test and Bartlett's test respectively, the assumptions of one-way Analysis of Variance for some data (i.e., the $\delta^2\text{H}$, $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ of epiphytes, and the $\delta^{18}\text{O}$ of potential water sources) could not be satisfied even after transformation. Thus, a nonparametric Kruskal-Wallis rank-sum test, followed by the pair-wise Wilcoxon rank-sum test, were used to test for differences of $\delta^2\text{H}$, $\delta^{18}\text{O}$, and $\delta^{13}\text{C}$ in epiphytes, and the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ in potential water sources [59]. To test the effects of $\delta^2\text{H}$ and $\delta^{18}\text{O}$ on $\delta^{13}\text{C}$ (WUE_i) in different groups of epiphytes (i.e., epiphytic lichens, epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants) linear mixed model (LMM) was used with the R package 'lme4' [60], in which the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of different groups were treated as fixed effects. The species of each group was incorporated as a random effect to account for the potential influence of different species across the result.

3. Results

3.1. Isotopic Compositions of Water Sources and Epiphytes

The isotope compositions ($\delta^2\text{H}$ and $\delta^{18}\text{O}$) of epiphytes and their potential water sources were presented to appraise the isotopic fractionation processes during the peak dry season (Figure 2). As a reference, the local meteoric water line (LMWL: $\delta^2\text{H} = 6.23 + 7.55 \delta^{18}\text{O}$, $R^2 = 0.86$, $p < 0.001$) was shown based on the rainfall data throughout 2018 and 2019. Compared with the global meter water line (GMWL: $\delta^2\text{H} = 10 + 8 \delta^{18}\text{O}$), the slope of LMWL was slightly lower than GMWL. The mean $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of fog water were above the LMWL and GMWL, indicating that the fog water experienced lower evaporative enrichment than the canopy humus and rainwater. Among the four groups of epiphytes, the isotopic signatures of epiphytic lichens and epiphytic seed plants were close to the fog water. In addition, the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of epiphytic bryophytes and epiphytic ferns were identical to humus.

The $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of fog water were higher ($p < 0.05$) than those of humus and rainwater (Figure 3 and Table S1). However, no significant difference was found between the humus and rainwater. Specifically, the average $\delta^2\text{H}$ and $\delta^{18}\text{O}$ values were $-27.4 \pm 4.9\text{‰}$ and $-5.93 \pm 0.55\text{‰}$ for fog water, $-70.8 \pm 3.1\text{‰}$ and $-8.80 \pm 0.46\text{‰}$ for humus, and $-88.9 \pm 13.7\text{‰}$ and $-11.89 \pm 1.71\text{‰}$ for rainwater. The $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of epiphytic lichens were significantly higher than epiphytic bryophytes ($p < 0.01$), epiphytic ferns ($p < 0.01$), and epiphytic seed plants ($p < 0.05$) (Figure 4). Meanwhile, we also found a significant difference in $\delta^2\text{H}$ and $\delta^{18}\text{O}$ between epiphytic bryophytes and epiphytic seed plants ($p < 0.01$). There was no significant difference between the epiphytic bryophytes and the epiphytic ferns. The average $\delta^2\text{H}$ and $\delta^{18}\text{O}$ values were $-34.7 \pm 4.0\text{‰}$ and $-3.38 \pm 0.92\text{‰}$ for epiphytic lichens, $-71.7 \pm 2.0\text{‰}$ and $-8.42 \pm 0.29\text{‰}$ for epiphytic bryophytes, and $-63.9 \pm 4.2\text{‰}$ and $-7.16 \pm 0.59\text{‰}$ for epiphytic ferns, and $-44.5 \pm 2.2\text{‰}$ and $-6.75 \pm 0.45\text{‰}$ for epiphytic seed plants. There were also interspecific differences ($p < 0.05$) among the epiphytic ferns. The $\delta^2\text{H}$ and $\delta^{18}\text{O}$ values of epiphytic ferns ranged from -77.33‰ to -46.46‰ and from -9.22‰ to -5.66‰ , respectively.

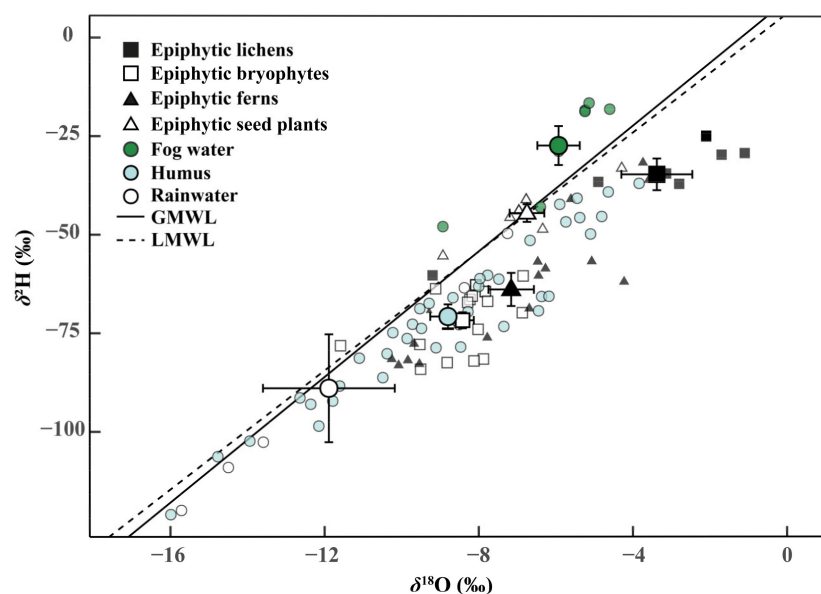


Figure 2. Average hydrogen and oxygen isotope ratios ($\delta^2\text{H}$ and $\delta^{18}\text{O}$) of epiphytes (Epiphytic lichens, $n = 4$ species; Epiphytic bryophytes, $n = 4$; Epiphytic ferns, $n = 4$; Epiphytic seed plants, $n = 4$) and water sources (Fog water, $n = 7$; humus, $n = 4$; and rainwater, $n = 5$) in the dry season (January 2019). The solid and segmented lines represent the global meteoric water line (GMWL: $\delta^2\text{H} = 10 + 8 \delta^{18}\text{O}$) and the local meteoric water line (LMWL: $\delta^2\text{H} = 6.23 + 7.55 \delta^{18}\text{O}$, $R^2 = 0.86$, $p < 0.001$), respectively. The LMWL was calculated by linear regression of the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of local precipitation data from 2018 to 2019. Error bars represent mean \pm SE of epiphytes and water sources.

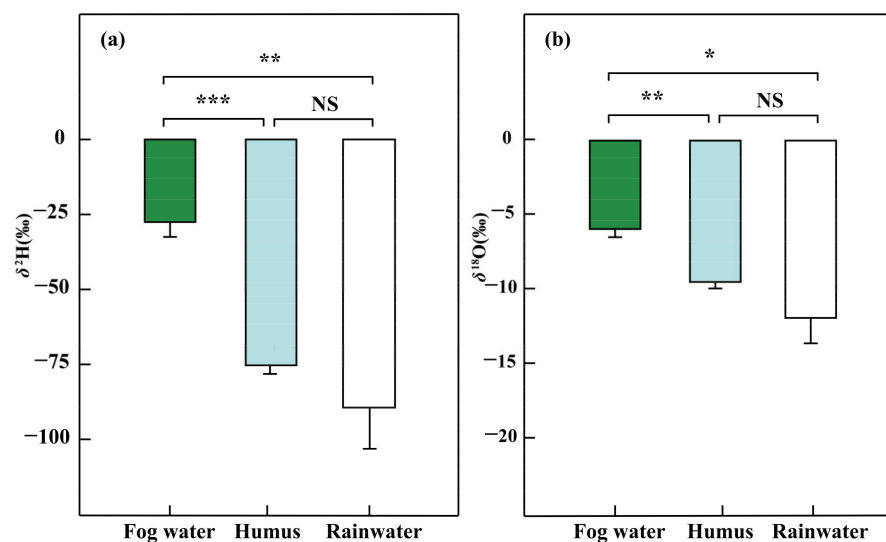


Figure 3. The $\delta^2\text{H}$ (a) and $\delta^{18}\text{O}$ (‰) (b) of different water sources (Fog water, $n = 7$; humus, $n = 4$; and rainwater, $n = 5$) in the dry season, January 2019. Wilcoxon rank sum test is used to verify the differences of water source samples (NS > 0.05 , * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$); Error bars represent means \pm SEs of different water sources.

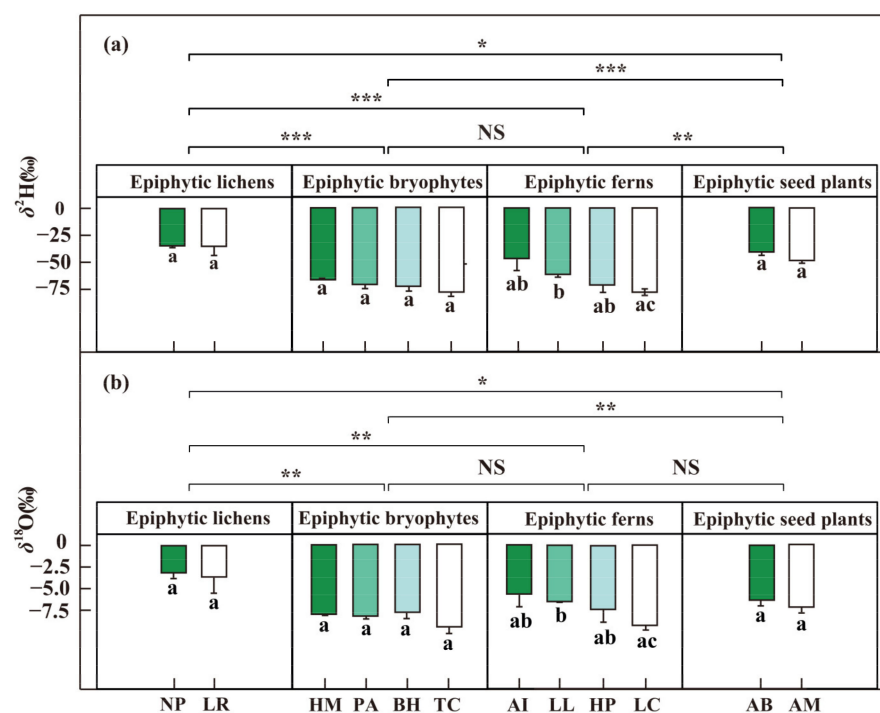


Figure 4. The $\delta^2\text{H}$ (a) and $\delta^{18}\text{O}$ (‰) (b) of epiphytes from different groups. Epiphytic lichens ($n = 4$): NP, *Nephromopsis pallescens*; LR, *Lobaria retigera*. Epiphytic bryophytes ($n = 4$): HM, *Hamaliiodendron montagneanum*; PA, *Plagiochila assamica*; BH, *Bazzania himalayana*; TC, *Thuidium cymbifolium*. Epiphytic ferns ($n = 4$): AI, *Asplenium indicum*; LL, *Lepisorus loriformis*; HP, *Hymenophyllum polyanthos*; LC, *Loxogramme chinensis*. Epiphytic seed plants ($n = 4$): AB, *Aeschynanthus buxifolius*; AM, *Agapetes mannii*. in the dry season, January 2019. Wilcoxon rank sum test is used to verify the differences of epiphyte samples (NS > 0.05, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$); Error bars represent mean \pm SE, and different letters with bars represent significant differences for each species ($p < 0.05$).

3.2. Partitioning of Water Sources for Epiphytes

The MixSIAR model showed that all the epiphytes could use fog water as their water sources (Figure 5). Because the epiphytic lichens had only two potential water sources (see Section 2.4), the contributions of fog water to *Nephromopsis pallescens* (NP) and *Lobaria retigera* (LR) were up to $86.3 \pm 9.9\%$ and $80.1 \pm 15.0\%$, respectively. With three potential water sources (i.e., fog water, humus, and rainwater), the epiphytic bryophytes (HM: *Hamaliiodendron montagneanum*; PA: *Plagiochila assamica*; BH: *Bazzania himalayana*; TC: *Thuidium cymbifolium*) acquired $24.0 \pm 11.7\%$ to $38.7 \pm 10.2\%$ of their water from fog water. Meanwhile, the epiphytic bryophytes of PA, BH, and TC could absorb $35.6 \pm 15.2\%$ to $38.6 \pm 16.3\%$ of their water from humus. Similarly, $46.4 \pm 12.6\%$ to $51.5 \pm 16.6\%$ of the water utilized by the epiphytic ferns of *Asplenium indicum* (AI) and *Lepisorus loriformis* (LL) was from fog water. Besides this, the contribution of fog water toward *Hymenophyllum polyanthos* (HP) and *Loxogramme chinensis* (LC) accounted for $31.0 \pm 13.2\%$ and $26.2 \pm 11.7\%$, respectively. However, the other two epiphytic ferns (HP: *Hymenophyllum polyanthos*; LC: *Loxogramme chinensis*) used humus as their major ($37.0 \pm 17.7\%$ to $38.1 \pm 15.9\%$) water source ($p < 0.05$). On the contrary, the epiphytic seed plants *Aeschynanthus buxifolius* (AB) and *Agapetes mannii* (AM) were highly dependent on fog water. The contribution percentage of fog water to AB and AM ranged from $57.7 \pm 11.1\%$ to $69.7 \pm 12.0\%$.

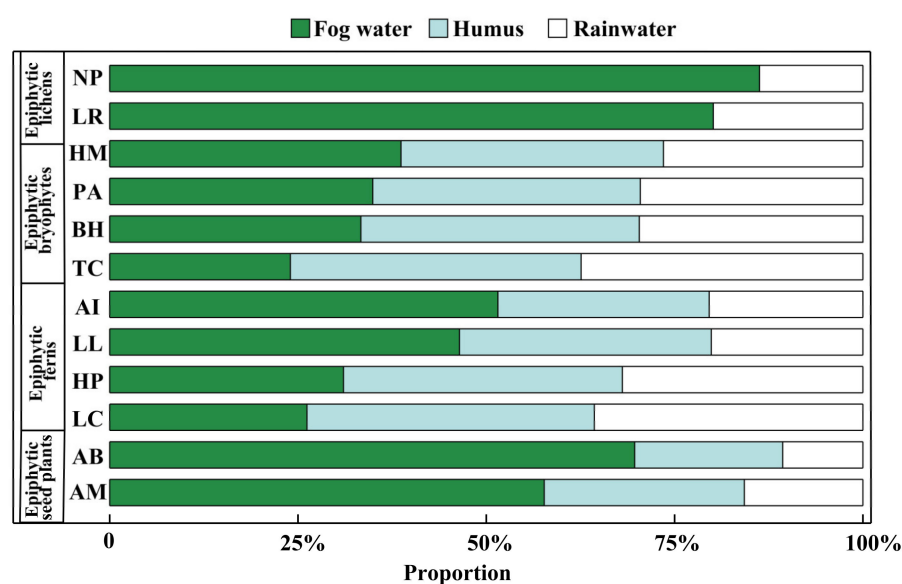


Figure 5. Partitioning of potential water sources of epiphytes from different groups by MixSIAR model in a subtropical montane cloud forest in the dry season. The abbreviations of species names are the same as in Figure 4.

3.3. Leaf Carbon Isotope Ratios

The $\delta^{13}\text{C}$ of epiphytic lichens were significantly higher ($p < 0.001$) than the other epiphyte groups (Figure 6 and Table S1), but there was no difference among epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants. The average $\delta^{13}\text{C}$ values were -23.77 ± 0.34 ‰ for epiphytes lichens, -29.59 ± 0.36 ‰ for epiphytic bryophytes, -30.96 ± 0.66 ‰ for epiphytic ferns, and -32.29 ± 0.26 ‰ for epiphytic seed plants. Meanwhile, the $\delta^{13}\text{C}$ values of epiphytic lichens were positively correlated with the $\delta^2\text{H}$ ($p < 0.05$) and interactions of $\delta^2\text{H} \times \delta^{18}\text{O}$ in water sources (Table 1). The $\delta^{13}\text{C}$ values of epiphytic bryophytes were different significantly ($p < 0.05$) among the four species; the average values of $\delta^{13}\text{C}$ in *Hamaliiodendron montagneanum* (HM: -27.57 ± 0.36 ‰) were significantly higher than other epiphytic bryophytes: -29.46 ± 0.29 ‰ for *Plagiochila assamica* (PA), -30.38 ± 0.23 ‰ for *Bazzania himalayana* (BH), and -30.96 ± 0.39 ‰ for *Thuidium cymbifolium* (TC). There were also interspecific differences ($p < 0.05$) among the four species of epiphytic ferns, which ranged from -27.81 ± 0.31 ‰ to -34.33 ± 0.40 ‰. These results indicated that the group-specific and interspecific differentiation of WUE_i (i.e., $\delta^{13}\text{C}$) were obvious among epiphytes.

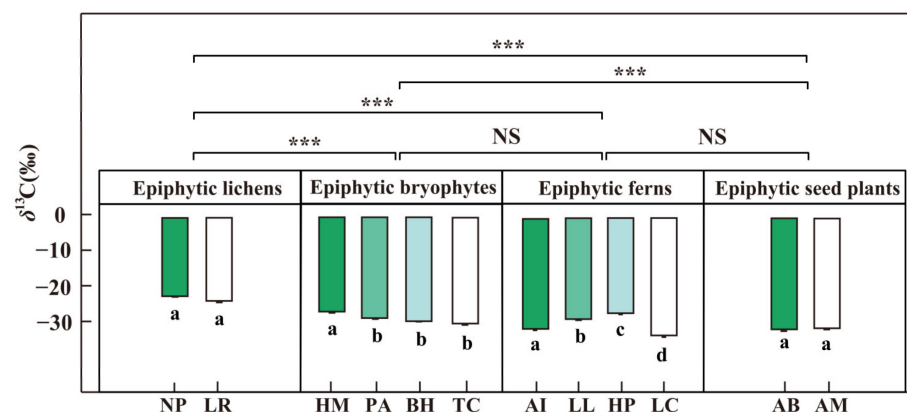


Figure 6. The $\delta^{13}\text{C}$ (‰) of mature leaves of epiphytes from different groups in the dry season, January 2019. Wilcoxon rank-sum test is used to verify the differences of epiphyte samples (NS > 0.05 , *** $p < 0.001$); Error bars represent mean \pm SE and different letters with bars represent significant differences for each species ($p < 0.05$). The abbreviations of species names are the same as in Figure 4.

Table 1. Linear mixed model analysis of the effects of $\delta^2\text{H}$, $\delta^{18}\text{O}$, and their interactions on WUE_i . ($\delta^{13}\text{C}$) of epiphytes from different groups (* $p < 0.05$). The species of each group was incorporated as a random effect, but were not shown here for simplicity.

Fixed Effect	SE	Epiphytic Lichens		SE	Epiphytic Bryophytes		SE	Epiphytic Ferns		SE	Epiphytic Seed Plants	
		χ^2	p		χ^2	p		χ^2	p		χ^2	p
$\delta^2\text{H}$	0.115	4.624	0.032 *	0.177	0.188	0.665	0.091	1.406	0.236	0.300	1.459	0.227
$\delta^{18}\text{O}$	0.091	2.394	0.122	0.197	0.002	0.965	0.077	0.065	0.065	0.194	3.644	0.056
$\delta^2\text{H} \times \delta^{18}\text{O}$	1.283	9.236	0.026 *	2.935	4.258	0.235	0.601	0.139	0.139	1.698	5.048	0.168

4. Discussion

Our study reveals the importance of fog in maintaining the water balance of epiphytes from a subtropical montane cloud forest in the dry season as well as the differentiation in water acquisition pattern of different epiphyte groups. The contribution of fog water to the water sources could be as high as 83.2% in epiphytic lichens. While epiphytic lichens and epiphytic seed plants mainly relied on fog water, the epiphytic bryophytes and epiphytic ferns were depending on both fog water and humus. We also found significant differences in WUE_i among epiphytic groups and species. The clear differentiation of water utilization among epiphytes provides a possible explanation for their coexistence. Our results also provide important implications for their responses to projected climate change patterns including a decline in fog persistence in the region.

4.1. Water Conditions in the Subtropical MCF

In our study area, we found that the slope of LMWL ($\delta^2\text{H} = 6.23 + 7.55 \delta^{18}\text{O}$, $R^2 = 0.86$, $p < 0.001$) was slightly lower than that in GMWL ($\delta^2\text{H} = 10 + 8 \delta^{18}\text{O}$). This result indicated that, in the dry season, there is a certain degree of fractionation during the rainfall process, which could be seen in other montane forests [61]. Moreover, the $\delta^2\text{H}$ and $\delta^{18}\text{O}$ of fog water were located above the LMWL, suggesting that fog water was more enriched compared with canopy humus and rainwater. Earlier studies also indicate that the isotopic signature of fog tends to be more enriched in heavier isotopes $\delta^2\text{H}$ and $\delta^{18}\text{O}$ than rainwater due to different condensation temperatures and histories [62]. In contrast, the water isotopes of canopy humus represent a mixture of evaporation and atmospheric water inputs (i.e., fog water and rainwater), which leads to the isotope signatures of $\delta^2\text{H}$ and $\delta^{18}\text{O}$ in humus falling in between fog water and rainwater [46].

4.2. Water Partitioning of the Four Epiphyte Groups

Generally, epiphytes could obtain fog water through root uptake of coalesced water that drips into the soil or by direct foliar water uptake (FWU) [63]. However, there are large differences among different groups and species in the ability of FWU. Although both epiphytic lichens and epiphytic bryophytes were probably some of the earliest land plants, they showed distinct structural differences. These differences can most likely be attributed to the evolution in plant morphology and structure. Epiphytic lichens are a symbiosis composed of fungus and algae [30]. Epiphytic lichens found in our study region can only attach to tree trunks and mainly absorb fog water (80.1% to 86.3%) during the dry season. However, it should be noted that in some other regions (e.g., the Pacific NW of the USA), lichens thrive in moist habitats with a bit of humus development [64], so the water sources of lichens from different forest ecosystems need to be analyzed further. Epiphytic bryophytes are true multicellular organisms with rhizomes [31,32]. They could absorb water from fog water (24.0% to 38.7%), rainwater (26.5% to 37.4%) and humus (35.6% to 38.6%). Our study also found that epiphytic seed plants absorb more fog water than epiphytic ferns. The epiphytic seed plants *Aeschynanthus buxifolius* (AB) and *Agapetes mannii* (AM) make full use of fog water (ranged from $57.7 \pm 11.1\%$ to $69.7 \pm 12.0\%$) during the dry season (Figure 5). This is most probably because epiphytic seed plants have complete roots, stomata, and the vascular system, which could absorb water sources more flexibly than

epiphytic ferns [33,65]. There was also an obvious interspecific differentiation in water acquisition among the epiphyte species (Figure 4). Wu et al. [27] pointed out that there was a divergence of water sources among four epiphytic orchids (*B. delitescens*, *C. sumatranus*, *C. viscosa*, and *P. weinmanniifolia*). Some studies further found that the leaf morphology and anatomy (e.g., leaf thickness and stomata) of epiphytes may account for a significant part of the variation in water uptake at the interspecific level [66,67].

It is obvious that the epiphytes in this study are C_3 species based on the carbon isotope data. The average $\delta^{13}C$ values of all epiphytes were below -20‰ (Figure 6), which is commonly interpreted as evidence for C_3 -photosynthesis [17]. Meanwhile, we found obvious group-specific and interspecific differentiation in WUE_i (i.e., $\delta^{13}C$) (Figure 6 and Table 1), which could be related to their different water acquisition patterns. Through the LMM model, we further found that the $\delta^{13}C$ of epiphytic lichens were correlated with their $\delta^2H / \delta^{18}O$ (Table 1). Thus, the difference in water acquisition patterns of epiphytes probably has an effect on WUE_i . This relationship has also been found in other studies [36,48]. Moreno-Gutierrez et al., [48] indicated that the large variation in WUE_i of coexisting species was affected by the stability of water sources. Because the habitats lack humus accumulation, epiphytic lichens in our study area can only absorb the unstable atmospheric water inputs (i.e., fog water and rainwater). Therefore, they exhibited higher WUE_i compared to other groups to cope with the relatively unstable water input during the dry season (Figures 5 and 6). The apparent differentiation of water utilization and WUE_i may facilitate the coexistence of different epiphytes in the canopy with the intermittent water supply.

The present study confirmed that all epiphytes can use fog water in various degrees (Figure 5). For the epiphytic lichens, we found that fog water accounted for 80.1–86.3% of their water acquisitions. The average contribution proportions of fog water to epiphytic bryophytes, epiphytic ferns, and epiphytic seed plants were 32.7%, 38.8%, and 63.7%, respectively. The dependence on fog water of epiphytes was possible because of the frequent fog events in the MCF [8,68]. Gehrig-Downie et al. [8] found that the frequent input of fog water enhances the growth of four epiphyte groups (bryophytes, lichens, pteridophytes, angiosperms) growth by shortening the desiccation periods. Several early studies in cloud montane forests also found that the canopy drip of throughfall could exceed the rainfall by 1.2–3.2 times, which confirmed the importance of fog as an extra input of water [8,51]. For epiphytes, fog water is more important because there are no vascular connections to their host plants (or ground) [24,27]. Zheng and Feng [69] pointed out that the fog water absorptions of the epiphytic orchids *C. occultata* and *S. dawsonianus* were higher than that in other non-epiphytes. In addition, some studies also suggested that fog water moving horizontally can be intercepted and accumulated on the epiphytic surfaces [24,70]. For instance, Song et al. [9] showed that fan bryophytes (*H. flabellatum*, *P. arbuscula*, and *P. assamica*) could intercept and absorb the fog water because of the flattened surfaces.

Notably, here in the present study, the δ^2H and $\delta^{18}O$ fog and rainfall water collected in open areas were used to represent the values of fog and rainwater used by epiphytes. This may raise some methodological concerns, such as the difference in physical location of the trees and rain/fog collectors, and the sampling timing [71,72]. However, there is no or neglectable evaporation and isotope discrimination when the fog moves from an open area to the forest and from the top canopy to the site of the epiphytes because the relative humidity (RH) is equal to or close to 100% when there is fog [50]. There might be isotope fractionation when the rainwater is intercepted by canopy leaves, evaporates for a while after the rain, and then drops to epiphytes [72,73]. However, the contribution of this leaf-intercepted water should be very limited as the remaining water on the leaf surface after the rainfall event will most likely be lost through evaporation or be directly absorbed by the tree leaves (FWU) [74]. In addition, the δ^2H and $\delta^{18}O$ of humus are changing overtime after a rain or fog event due to evaporation. Nevertheless, our study suggested that the effect of evaporation on canopy humus is limited because the isotope signatures of

humus are close to ($p > 0.05$) that of rainwater (Figure 3). Actually, the isotopic composition ($\delta^2\text{H}$ and $\delta^{18}\text{O}$) of canopy humus varies slightly within the forest. In order to reduce the spatial uncertainty, the humus samples we collected were one-to-one correspondence to the epiphyte samples during the experiment period. Therefore, we collected humus while we were collecting epiphyte samples, which should be the best and most practical approach because the isotope signatures of humus collected should be close to that of the water absorbed by epiphytes from humus during collection. These potential technical uncertainties apply to all water source determination studies using $\delta^2\text{H}$ and $\delta^{18}\text{O}$ [56,75], and can be further tested with a study quantifying the spatial-temporal variation of $\delta^2\text{H}$ and $\delta^{18}\text{O}$ in a high sampling resolution [76].

4.3. Ecological Implications

High fog dependence of epiphytes indicates that predicted declines in fog persistence due to climate change may negatively impact their performance and survival. Fog water has been suggested to be an important water input for epiphytes in forest ecosystems [51,63]. The present study further confirmed this and indicated a significant differentiation of fog water utilization among epiphytes. With the intensification of global warming [2], rising cloud bases will intensify the solar radiation received by MCF, which may further decrease the fog frequency and duration [38]. The reduction of fog events and/or duration will probably result in a high mortality of fog-dependent species (especially epiphytes) under projected climate change scenarios [51]. Therefore, the epiphyte community in MCF may experience a change in composition and functioning. Previous results also suggested that decreases in fog persistence will negatively affect the productivity and longevity of epiphytes [62,77]. For example, Nadkarni and Solano [77] found that the decline of fog persistence significantly increased leaf mortality, reduced leaf production, and shortened the longevity of epiphytes. Because epiphytes play crucial roles in hydrological and nutrient cycles, and promote the survival of other plants and animals in the canopy habitat [7,12], the potential deterioration of epiphytes due to decreasing fog persistence may have cascading effects on the whole forest ecosystem. Therefore, more conservation attention should be paid to epiphytes in montane forest ecosystems under climate change.

5. Conclusions

Our results indicated that epiphytic lichens and epiphytic seed plants depended mainly on fog water whereas the epiphytic bryophytes and epiphytic ferns relied on both fog water and humus. Also, there were obvious variations in WUE_i among groups and species, and the WUE_i of epiphytic lichens were appreciably higher than the other epiphyte groups. The variation in WUE_i was probably related to the differentiation of water resources and water use strategies. Despite the fact that water sources of epiphytes were significantly different among groups and species, this study clearly showed that all epiphytes rely on fog water heavily in the dry season. Thus, our results suggest a dim future for epiphytes as fog persistence is predicted to decline. Conservation efforts may be needed to retain the high diversity of epiphytes in this region. The response of these epiphytes to increasingly severe drought also needs to be studied to better understand their drought response mechanisms and to develop mitigation strategies.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/w13223237/s1>, Figure S1: Photograph of self-made V-shaped fog collector (0.3 m × 1 m) (a) and cylindrical rain collector (0.2 m × 0.65 m) (b) at the study site, Table S1: The $\delta^2\text{H}$, $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ (‰) (mean ± SE) of water sources and epiphytes from different groups in a subtropical montane cloud forest in the dry season. The abbreviations of species names are the same as in Figure 4.

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